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The attached photocopy is a true copy of the following document:

The specification, claims, abstract and drawings as filed with the application on the filing date indicated above.

By assignment dated 08 Sept 1999 and filed 09 Sept 1999, the application has been assigned to Biolmage A/S





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A method for preventing or treating adverse conditions which may be reduced or abolished by modulating the effectiveness of one or more I-kappaB kinases.

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SUMMARY OF THE INVENTION

This application describes a method by which to identify novel chemical entities 5 found to inhibit the activation of NF-kappaB and/or degradation of I-kappaB in living cells. Such compounds will specifically modulate activation of NF-kappaB and/or degradation of I-kappaB in a way that can be identified by detection and quantification of the I-kappaB kinase (IKK) targeting or localisation in the cells of interest using quantitative fluorescence redistribution assays. The preferred mode of 10 action being sought is dislocation or interference with the targeting of specific isoforms of the IKK from or to their anchoring sites within cells, which will comprise the I-kappaB kinase anchoring protein (IKAP) and its associated enzymes, thereby reducing their specific effectiveness, not their enzymatic capacity.

In its broadest aspect, the present application relates to a novel method for preventing or treating, in an animal in need thereof, an adverse condition which may be reduced or abolished by modulating the activity of one or more lKKs. The method comprises modulation of the specific effectiveness of IKKs by modulating their spatial distribution within cells of the animal.

The IKK is chosen from the group consisting of IKK α , IKK β , IKK γ and NIK. In one embodiment IKK\$\beta\$ is the preferred isoform. The animal with the adverse condition may be a mammal and preferably a human.

In one embodiment of the invention modulation of the specific effectiveness of the IKK is a dislocation of the IKK from a native location within the cell.

In another embodiment of the invention modulation of the specific effectiveness of the IKK involves a disruption of its targeting to a native location within the cell.

In another embodiment of the invention modulation of the specific effectiveness of the IKK involves interference with the redistribution of the IKK, the redistribution being associated with an increase or a decrease of the specific effectiveness of the IKK.

The modulation of the specific effectiveness of the IKK may involve both an upregulation or a down-regulation of the effectiveness of the IKK to perform its function within the cell.

The compounds found by this methodology are supposedly useful in the treatment of the following diseases/conditions: asthma, allergy, chronic inflammation and autoimmune diseases.

This patent application is associated with the patent application "An improved method..." enclosed hereto as appendix A. Appendix A is considered part of this application.

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BACKGROUND

Chronic inflammation is the result of unbalanced and continued production of inflammatory cytokines. Cytokines are produced in cascades, the pro-inflammatory TNFα and IL-1β often responsible for initiating a process, which leads to a more general production of further cytokines. This cascade of gene expression is largely under the control of NF-kappaB, a ubiquitous transcription factor that, by regulating the expression of multiple inflammatory and immune genes, plays a critical role in host defence and in chronic inflammatory diseases (Sen and Baltimore, 1986; Mukaida et al., 1990; Beg et al., 1993; Cogswell et al., 1993). NF-kappaB is activated not only by cytokines, but also by reactive oxygen species (ROS), viruses, and a range of other generally noxious and pathogenic stimuli (Blackwell et al., 1997; Schulzwe-Osthoff et al., 1997). Activation of NF-kappaB via ROS has been implicated in neurodegenerative disorders such as Parkinson's and Alzheimer's (Lesoualc'h et al., 1998; O'Neill et al., 1997) and also in inflammatory bowel disease (Jourd'heuil et al., 1997). Tissue inflammatory reponse to x-rays is mediated directly by NF-kappaB (Hallahan et al., 1995). Activation of NF-kappaB has been implicated in the production of atherosclerotic lesions of smooth muscle cells (Bourcier et al., 1997) and in cardiac inflammatory disorders (Hattori et al., 1997). NF-kappaB/Rel transcription factors are also known to play a role in the pathogenesis of certain tumours, especially those of haematopoetic origin (Neumann et al., 1997), and constitutive (autocrine) activation of NF-kappaB is known to promote a resistance to apoptotic stimuli (Giri et al., 1998). Inhibitors of NF-kappaB should increase the cytotoxic efficacy of anticancer chemotherapies (Bours et al., 1998).

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The inflammatory pathways are notoriously complex, yet the feasibility of reducing or eliminating inflammatory responses through modulation of NF-kappaB activity has already been demonstrated in a number of different cells (Makarov *et al.*, 1997).

The NF-kappaB/Rel group of transcription activators and their co-evolved regulatory proteins, the inhibitors of kappa B (I-kappaBs), play important roles in many cellular signalling processes in vertebrates, which include controlling communication between cells, embryo development, maintenance of cell type specific expression of genes as well as co-ordinating the inflammatory response to stressors and viral infection (Wulczyn et al., 1996). The key proteins involved in this control system divide into distinct groups: a) Those that bind DNA. These belong to the Rel family of transcription factors (Ghosh et al., 1990) and include p50, p65, p52/49, p75/Rel and RelB. Only dimers bind DNA, but these can be homodimers or heterodimers. p65/p50 heterodimer is the most abundant, and plays a more elaborate role than other factors in regulating gene expression (Baldwin, 1996). b) Those that interact with the DNAbinding subunits in cytoplasm, which include the inhibitory I-kappaB α and I-kappaB β molecules (Bauerle and Baltimore, 1988), and the precursor molecule p105 (Naumann et al., 1993). c) Those transcriptional coactivators which interact with the DNAbinding subunits in the nucleus, such as Bcl3 (Nolan et al., 1993; Watanabe et al., 1997) and Cbp/p300 (Zhong et al.,, 1998). d) Kinases which activate proteasomal destruction of I-kappaB α and β subunits - the I-kappaB kinases (Beg et al., 1993). e) Kinases which directly phosphorylate the DNA-binding subunits in cytoplasm and nucleus to modulate their activity, such as PKA (Zhong et al., 1998), casein kinase II (Bird et al., 1997) and others (Hayashi et al., 1993; Schulze-Osthoff et al., 1997).

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Inactive p65/p50 NF-kappaB dimers are held in the cytoplasm coupled to inhibitory I-kappaB molecules (α and β isoforms) via the p65 subunits. Activated I-kappaB kinases (IKK) phosphorylate the inhibitors, targeting them for ubiquitination and subsequent proteasomal digestion (Beg *et al.*, 1993). The released subunits translocate to the nucleus and there activate transcription.

The I-kappa kinases (IKK- α , IKK- β and IKK- γ) have been shown to be part of a large multi-component complex (Chen et al. 1996; Rothwarf et al., 1998). It is likely to assume that the assembly and disassembly of the IKK complex is controlled by a

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scaffold protein termed IKK-complex-associated protein, IKAP (Cohen et al. 1998). It is expected that a tight assembly of the complex is necessary for the IKKs to be activated by the NF-kappa-B-inducing kinase (NIK) and thereby induce phosphorylation of the I-kappaB subunits. Interestingly the affinity of IKK-β for IKAP diminishes upon phosphorylation of IKK-β by NIK.

Glucocorticoids (GC) are powerfully efficient modulators of inflammation, but suffer from the potential hazards of suppressing necessary protective responses to infection and decreasing some essential healing processes. They modulate cytokine expression by a combination of genomic mechanisms. The activated GC-receptor complex can (i) bind to and inactivate AP-1 or NF-kappaB, (ii) upregulate I-kappaB production via GC response elements (iii) reduce the half-life of cytokine mRNAs (Brattsand & Linden 1996). But steroid treatment broadly attenuates all cytokine production from all lymphocytes, so not only do levels of the inflammatory cytokines fall, but also that of the anti-inflammatory IL-10. Specific modulation of Th1-type pathways would be an initial goal of this project.

It is also known that some fibroblast cell NF-kappaB-mediated responses are likely governors of inflammatory progression, so inhibition of such responses could have detrimental effects (Smith et *al.*, 1997). Therapies, which maintain appropriate feedback systems, but modulate inappropriate cytokine production represent an unmet medical need.

An attractive therapeutic intervention to be used in the treatment of chronic inflammatory conditions is inhibition of the I-kappaB degradation. Blocking the ubiquitin proteasome pathway (PharmaProjects, Accession no. 023654 and 027675), can directly inhibit this degradation. Another mechanism that is being pursued is inhibition of the enzymatic activity of either of the IKKs or NIK (public statement from Signal Pharmaceuticals).

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In the present invention I-kappaB degradation is inhibited by a novel mechanism namely inhibition of the redistribution of specific IKKs (IKK- β and IKK- α). In contrast to previous interventions involving IKK the presented invention does not

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involve direct inhibition of the IKK enzymatic activity. This completely novel mechanism for inhibition of the overall effect of the IKK complex provides clear advantages as it opens for a higher IKK isoform selectivity and a higher cell specificity of the therapy.

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DETAILED DISCLOSURE

In the present specification and claims, the term "influence" covers any influence to which the cellular response comprises a redistribution. Thus, e.g., heating, cooling, high pressure, low pressure, humidifying, or drying are influences on the cellular response on which the resulting redistribution can be quantified, but perhaps the most important influence is the influence of contacting or incubating the cell or cells with a substance which is known or suspected to cause a redistribution. In another embodiment of the invention the influence could be substances from a compound drug library.

In the present context, the term "green fluorescent protein" (GFP) is intended to indicate a protein which, when expressed by a cell, emits fluorescence upon exposure to light of the correct excitation wavelength (cf. Chalfie, M. et al. (1994) Science 263, 802-805). "GFP" as used herein includes wild-type GFP derived from the jelly fish Aequorea victoria and modifications of GFP, such as the blue fluorescent variant of GFP disclosed by Heim et al. (Heim, R. et al. (1994). Proc.Natl.Acad.Sci. 91:26, pp 12501-12504), and other modifications that change the spectral properties of the GFP fluorescence, or modifications that exhibit increased fluorescence when expressed in cells at a temperature above about 30°C described in PCT/DK96/00051, published as WO 97/11094 on 27 March 1997 and hereby incorporated by reference, and which comprises a fluorescent protein derived from Aequorea Green Fluorescent Protein or any functional analogue thereof, wherein the amino acid in position 1 upstream from the chromophore has been mutated to provide an increase of fluorescence intensity when the fluorescent protein of the invention is expressed in cells. Preferred GFP variants are F64L-GFP, F64L-Y66H-GFP and F64L-S65T-GFP. An especially preferred variant of GFP for use in all the aspects of this invention is EGFP (DNA encoding EGFP which is a F64L-S65T variant with codons optimized for expression in mammalian cells is

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available from Clontech, Palo Alto, plasmids containing the EGFP DNA sequence, cf. GenBank Acc. Nos. U55762, U55763).

The terms "intracellular signalling pathway" and "signal transduction pathway" are intended to indicate the coordinated intracellular processes whereby a living cell transduces an external or internal signal into cellular responses. Said signal transduction will involve an enzymatic reaction said enzymes include but are not limited to protein kinases, GTPases, ATPases, protein phosphatases, phospholipases and cyclic nucleotide phosphodiesterases. The cellular responses include but are not limited to gene transcription, secretion, proliferation, mechanical activity, metabolic activity, cell death.

The term "second messenger" is used to indicate a low molecular weight component involved in the early events of intracellular signal transduction pathways.

The term "luminophore" is used to indicate a chemical substance which has the property of emitting light either inherently or upon stimulation with chemical or physical means. This includes but is not limited to fluorescence, bioluminescence, phosphorescence, chemiluminescence.

The term "mechanically intact living cell" is used to indicate a cell which is considered living according to standard criteria for that particular type of cell such as maintenance of normal membrane potential, energy metabolism, proliferative capability, and has not experienced any physically invasive treatment designed to introduce external substances into the cell such as microinjection.

In the present context, the term "permeabilised living cell" is used to indicate cells where a pore forming agent such as Streptolysin O or *Staphylococcus Aureus* α-toxin has been applied and thereby incorporated into the plasma membrane in the cells. This creates proteinaceous pores with a defined pore size in the plasma membranes of the exposed cells. Pores could also be made by electroporation, i.e. exposing the cells to high voltage discharges, a procedure that creates small holes in the plasma membrane by coagulating integral membrane proteins. Treatment with a mild detergent such as saponin may accomplish the same thing. Common to all these treatments is that pores are formed only in the plasma membrane without affecting the integrity of cytoplasmic structural elements and organelles. The term living in this context means

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that the permeabilised cell or cells bathed in a solution mimicking the intracellular milieu still have functional organelles, such as actively respiring mitochondria and endoplasmatic reticulum that can take up and release calcium ions, and functional structural elements. In one embodiment this method is applied so that substances that normally can not traverse the plasma membrane, but most likely exert their influence intracellularly, can be introduced and their influence studied. In another embodiment this method is used to record the response to an influence from many cells simultaneously.

In the present context, the term "permeabilisation" is intended to indicate the selective disruption of the plasma membrane barrier so that soluble substances freely mobile in the cytosol may be lost from the interior of the cells. The permeabilisation can be achieved as described above under "permeabilised living cells" or by using other chemical detergents such as Triton X-100 or digitonin in carefully titrated amounts.

The term "physiologically relevant", when applied to an experimentally determined redistribution of an intracellular component, as measured by a change in the luminescence properties or distribution, is used to indicate that said redistribution can be explained in terms of the underlying biological phenomenon which gives rise to the redistribution.

The terms "image processing" and "image analysis" are used to describe a large family of digital data analysis techniques or combination of such techniques which reduce ordered arrays of numbers (images) to quantitative information describing those ordered arrays of numbers. When said ordered arrays of numbers represent measured values from a physical process, the quantitative information derived is therefore a measure of the physical process.

The term "mammalian cell" is intended to indicate any living cell of mammalian origin. The cell may be an established cell line, many of which are available from The American Type Culture Collection (ATCC, Virginia, USA) or a primary cell with a limited life span derived from a mammalian tissue, including tissues derived from a transgenic animal, or a newly established immortal cell line derived from a mammalian tissue including transgenic tissues, or a hybrid cell or cell line derived by fusing different celltypes of mammalian origin e.g. hybridoma cell lines. The cells may optionally express one or more non-native gene products, e.g. receptors,

enzymes, enzyme substrates, prior to or in addition to the fluorescent probe. Preferred cell lines include but are not limited to those of fibroblast origin, e.g. BHK, CHO, BALB, or of endothelial origin, e.g. HUVEC, BAE (bovine artery endothelial), CPAE (cow pulmonary artery endothelial), HLMVEC (human lung microvascular endothelial cells) or of pancreatic origin, e.g. RIN, INS-1, MIN6, bTC3, aTC6, bTC6, HIT, or of hematopoietic origin, e.g. primary isolated human monocytes, macrophages, neutrophils, basophils, eosinophils and lyphocyte populations, AML-193, HL-60, RBL-1, adipocyte origin, e.g. 3T3-L1, neuronal/neuroendocrine origin, e.g. AtT20, PC12, GH3, muscle origin, e.g. SKMC, A10, C2C12, renal origin, e.g. HEK 293,

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The term "hybrid polypeptide" is intended to indicate a polypeptide which is a fusion of at least a portion of each of two proteins, in this case at least a portion of the green fluorescent protein, and at least a portion of a catalytic and/or regulatory domain of a protein kinase. Furthermore a hybrid polypeptide is intended to indicate a fusion polypeptide comprising a GFP or at least a portion of the green fluorescent protein that contains a functional fluorophore, and at least a portion of a biologically active polypeptide as defined herein provided that said fusion is not the Glucocorticoid Receptor-GFP disclosed by Carey, KL et al. and Guiliano, KA et al., respectively. Thus, GFP may be N- or C-terminally tagged to a biologically active polypeptide, optionally via a linker portion or linker peptide consisting of a sequence of one or more amino acids. The hybrid polypeptide or fusion polypeptide may act as a fluorescent probe in mechanically intact or permeabilised living cells carrying a DNA sequence encoding the hybrid polypeptide under conditions permitting expression of said hybrid polypeptide. The term hybrid polypeptide or fusion polypeptide is intended also to include the term "fluorescent probe", where the latter is used to indicate a fluorescent fusion polypeptide comprising a GFP or any functional part thereof which is N- or C-terminally fused to a biologically active polypeptide as defined herein, optionally via a peptide linker consisting of one or more amino acid residues, where the size of the linker peptide in itself is not critical as long as the desired functionality of the fluorescent probe is maintained. A fluorescent probe according to the invention is expressed in a cell and basically mimics the physiological behaviour of the biologically active polypeptide moiety of the fusion polypeptide.

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The term "kinase" is intended to indicate an enzyme that is capable of phosphorylating a cellular component.

The term "protein kinase" is intended to indicate an enzyme that is capable of phosphorylating serine and/or threonine and/or tyrosine in peptides and/or proteins.

The term "phosphatase" is intended to indicate an enzyme that is capable of dephosphorylating phosphoserine and/or phosphothreonine and/or phosphotyrosine in peptides and/or proteins.

The term "cyclic nucleotide phosphodiesterase" is intended to indicate an enzyme that is capable of inactivating the second messengers cAMP and cGMP by hydrolysis of their 3'-ester bond.

In the present context, the term "biologically active polypeptide" is intended to indicate a polypeptide affecting intracellular processes upon activation, such as an enzyme which is active in intracellular processes or a portion thereof comprising a desired amino acid sequence which has a biological function or exerts a biological effect in a cellular system. In the polypeptide one or several amino acids may have been deleted, inserted and/or replaced to alter its biological function, e.g. by rendering a catalytic site inactive or by disrupting the targeting sequence. In another embodiment, one or several amino acids may have been deleted, inserted and/or replaced without altering the biological function of the polypeptide, that is, it remains biologically equivalent. Preferably, the biologically active polypeptide is selected from the group consisting of proteins taking part in an intracellular signalling pathway, such as enzymes involved in the intracellular phosphorylation and dephosphorylation processes including kinases, protein kinases and phosphorylases as defined herein, but also proteins making up the cytoskeleton play important roles in intracellular signal transduction and are therefore included in the meaning of "biologically active polypeptide" herein. More preferably, the biologically active polypeptide is a protein which according to its state as activated or non-activated changes localisation within the cell, preferably as an intermediary component in a signal transduction pathway. Included in this preferred group of biologically active polypeptides are cAMP dependent protein kinase A and cyclic nucleotide phosphodiesterases.

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The term "a substance" is intended to indicate any sample which has a biological function or exerts a biological effect in a cellular system. The sample may be a sample of a biological material such as a sample of a body fluid including blood, plasma, saliva, milk, urine, or a microbial or plant extract, an environmental sample containing pollutants including heavy metals or toxins, or it may be a sample containing a compound or mixture of compounds prepared by organic synthesis or genetic techniques.

The phrase "any change in fluorescence" means any change in absorption properties, such as wavelength and intensity, or any change in spectral properties of the emitted light, such as a change of wavelength, fluorescence lifetime, intensity or polarisation, or any change in the intracellular localisation of the fluorophore. It may thus be localised to a specific cellular component (e.g. organelle, membrane, cytoskeleton, molecular structure) or it may be evenly distributed throughout the cell or parts of the cell.

The term "organism" as used herein indicates any unicellular or multicellular organism preferably originating from the animal kingdom including protozoans, but also organisms that are members of the plant kingdoms, such as algae, fungi, bryophytes, and vascular plants are included in this definition.

The term "nucleic acid" is intended to indicate any type of poly- or oligonucleic acid sequence, such as a DNA sequence, a cDNA sequence, or an RNA sequence.

The term "biologically equivalent" as it relates to proteins is intended to mean that a first protein is equivalent to a second protein if the cellular functions of the two proteins may substitute for each other, e.g. if the two proteins are closely related isoforms encoded by different genes, if they are splicing variants, or allelic variants derived from the same gene, if they perform identical cellular functions in different cell types, or in different species. The term "biologically equivalent" as it relates to DNA is intended to mean that a first DNA sequence encoding a polypeptide is equivalent to a second DNA sequence encoding a polypeptide if the functional proteins encoded by the two genes are biologically equivalent.

The term "fixed cells" is used to mean cells treated with a cytological fixative such as glutaraldehyde or formaldehyde, treatments which serve to chemically cross-link and

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stabilize soluble and insoluble proteins within the structure of the cell. Once in this state, such proteins cannot be lost from the structure of the now-dead cell. In the present context a "quantitative fluorescence redistribution assay" is intended to indicate an assay whereby it is possible to observe and quantify the subcelluar

localisation and possible redistribution of an biologically active polypeptide, or part thereof, genetically or chemically tagged with a luminophore inside an intact living cell or cells or permeabilised living cells. The subcelluar location and redistribution may be monitored using fluorescence microscopy or fluorescence imaging microscopy but is preferably monitored using a fluorescence imaging plate reader or a fluorescence plate reader for improved throughput. A more thorough description is given in Appendix A.

In the present context a "mortal cell line" is used to indicate animal cells that may grow in vitro, given the right conditions, but that have a definite life span of a number of cell divisions or days, week or months beyond which it is not at present possible to keep them alive.

In the present context an "immortalised cell line" is used to indicate cells of animal origin where the normal limitations for cell life and number of cell divisions do not apply. Essentially, such cells can live, grow and divide for an unlimited or very long (years to decades) time.

The term "targeting sequence" is used to indicate the amino-acid sequence of a biologically active polypeptide that contains the actual structure or structures necessary for association of the biologically active polypeptide with its native intracellular binding sites. The term "targeting sequence" is also used to indicate the amino-acid sequence of a protein that contains the actual structure or structures necessary for association of a biologically active polypeptide with the protein. The term "targeting" is used to indicate the process whereby a spatially distributed protein is directed to the intracellular sites and maintained at the intracellular sites to which it is normally anchored or associated. These anchoring sites are normally assumed to be the intracellular sites where the protein has its optimal function for the 30 cell.

The term "dislocate" and derivatives thereof is used to indicate the process whereby an intracellularly spatially distributed protein is forced to detach from its normal anchoring or association structures in the cells due to intercalation of another,

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preferably smaller, compound at the site of anchoring or association. This usually means that the optimal function of the protein within the cell is lost or reduced and that a larger portion of the protein molecules are freely mobile within the cytoplasm. In the present context a "screening assay" is intended to mean any measurement protocol, including materials, cells, instruments, chemicals, reagents, detection units, calibration and quantification procedures used to measure a response from mechanically intact or permeabilised living cells relevant to influences on an intracellular pathway.

In the present context a "primary screening assay" is used to indicate the first screening assay in a discovery project that is used to select and sort all compounds available to the project according to the quantified effect of the compounds in the assay.

In the present context a "counterscreen" is intended to mean a screening assay that is relevant to a phenomenon that is undesirable seen from the point of view of the discovery project.

In the present context a "discovery project" is intended to mean the process whereby general or specific ideas about ways of how to modulate an intracellular signalling pathway are exploited in order to find new chemical compounds that can be used to modulate the intracellular signalling pathway and thereby treat, reduce or abolish symptoms associated with a condition or a disease that is lethal, degenerative, performance-reducing or just uncomfortable to an animal, preferably a human being. The aim of the discovery project is to produce drug candidates that can be tested as potential drugs in an animal, preferably in human beings. The term "discovery project" also encompasses the actual group of individuals, screening assays, tests, machinery, cells, animals and compounds involved in different aspects of the project. The term "tagging" is used to indicate the process whereby a luminophore is genetically or chemically attached to the protein, or part of the protein, of interest to

the discovery project.

The term "primary hit" is used to indicate compounds identified in the primary screening assay as having at least the minimum level of desired effect that has been specified in the discovery project.

The term "primary lead compound" is used to indicate a primary hit that has at least the minimal level of desired potency and specificity predetermined by the discovery project.

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The term "dose-response relationship" is in the present context intended to mean a clear correlation between the quantified response of cells in a screening assay to application of an influence, such as a compound, and the concentration of the applied influence. The response to the influence may be both an up-regulation and a down-regulation of the quantitated parameter used in the screening assay.

In the present context, the term "potency" is intended to mean the ability of an influence to affect the process under study. The process under study may be, for

example a screening assay or a specific physiological or pathophysiological response in an animal.

In the present context, the term "selectivity" is intended to mean the difference in potency on the desired process, such as a screening assay, and an undesired process, such as a counterscreen, with the view of the discovery project. An influence or a compound is said to display selectivity if the potency for the desired process is higher than for the undesired process.

In the present context, the term "structure-activity relationship" or "SAR" is intended to mean the situation where a direct relationship exists between a compound and modifications made to the compound and the activity of the compound and the modifications made to the compound in one or more screening assays. The process of building a SAR may be used to direct the chemical construction of new compounds with higher potency and selecivity than the original compound.

The term "drug candidate lead" is used to indicate compounds that may be pursued by

a discovery project as potential candidates for the final outcome of the project. In the present context, the term "efficacy" is intended to mean the ability of a compound to affect the process or condition under study. It is closely related to the term "potency" but is in the present context used when relating to effects of a compound on more complex screening assays than the primary screening assay or counterscreens and when relating to effects of a compound in animals.

In the present context, the term "toxicity" is intended to mean that a compound in some way is toxic to cells, tissues or animals. The toxicity means that the cells, tissues or animals will in some way be harmed if the compound is applied at a sufficient concentration. The effects may ultimately lead to cell, tissue or animal death or a limited life compared to the normal condition.

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In the present context, the term "physiology" is intended to mean the normal function of biological and biochemical processes inside cells, between cells and in the whole organism or animal.

In the present context, the term "pathophysiology" is intended to mean deviations from the normal function of biological and biochemical processes inside cells, between cells and in the whole organism or animal that may be part of a condition or disease.

In the present context, the term "pathogenesis" is intended to mean the process, be it genetical, biological, biochemical, chemical or environmental, that ultimately may explain, at least in part, the apparent patophysiology associated with a condition or disease in an animal.

In the present context, the term "fractionated cells" is intended to mean the outcome of a simple division of initially mechanically intact living cells into two fractions, particulate (the components that can be sedimented by centrifugation at more than 10 000xg and not more than 100 000xg for 10 minutes) and soluble fraction (the soluble components and small membrane fragments that do not sediment), after subjecting the cells to plasma membrane disruption either mechanically with some form of homogeniser or sonicator or osmotically (hypoosmotic shock) or through some kind of permeabilisation of the plasma membrane with detergents, toxins or electroporation.

The term "parenteral route of administration" is used to indicate the administration of a drug or compound in solution to an animal, such as a mammal or a human, by injection or infusion of the drug or compound into the bloodstream of the animal via an injection needle iserted into one of the animals blood vessels, preferably a vein.

The term "oral route of administration" is used to indicate the administration of a drug or compound in solution or as a solid to an animal, such as a mammal or a human, by placing the drug or compound in the mouth of the animal so that the animal itself can swallow the drug or compound or have it delivered to the stomach or intestine by intubation. When the drug or compound enters the stomach and intestine it will be taken up over the mucosa into the bloodstream and administered via the blood stream to the tissues and organs where it is to exert its effect, or it will be acting locally in the stomach and intestine.

The term "pulmonary route of administration" is used to indicate the administration of

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a drug or compound as an aerosol with either solid or liquid particles to an animal, such as a mammal or a human, by placing the drug or compound container close to or in contact with the mouth and/or nose of the animal so that the animal itself can inhale the drug or compound aerosol. When the drug or compound enters the peripheral bronchioloi and alveoli it will be taken up over the alveolar membrane, either into the bloodstream and administered via the blood stream to the tissues and organs where it is to exert its effect or it will act locally in the lungs on lung, vessel and muscle cells as well as any other cell type present there.

The term "cutaneous route of administration" is used to indicate the administration of a drug or compound in solution or as a solid to an animal, such as a mammal or a human, by placing the drug or compound on the skin of the animal. The drug can then enter the blood vessels under the skin as it is permeaing the skin and thereby be taken up into the bloodstream and administered via the blood stream to the tissues and organs where it is to exert its effect. It may also exert an effect locally on the site of application on the skin.

The term "rectal route of administration" is used to indicate the administration of a drug or compound in solution or as a solid to an animal, such as a mammal or a human, by placing the drug or compound in the rectal cavity of the animal. When the drug or compound enters the rectum and parts of the large intestine it will be taken up over the mucosa into the bloodstream and administered via the blood stream to the tissues and organs where it is to exert its effect, or it will act locally in the rectum and parts of the large intestine.

Several IKKs are known. When setting up a program to identify pharmacological agents that affect the intracellular distribution of a target IKK, it is first necessary to choose the target from the IKKs known. This may be done according to various criteria. A first criterion is that it is imperative that the target IKK be present in the tissue or cell type(s) where the pharmacological agent is to exert its effect. A second criterion is that it is desirable that the target not be present in tissues or cell types where no pharmacological effects are desired. A third criterion is that the target IKK displays a non-random pattern of intracellular distribution.

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Establishing the expression patterns of IKKs in relation to tissues and cell types is best done using the methods of detection of mRNA, e.g. Northern analysis, which is a well established procedure. Briefly, mRNA isolated from a given source is probed with a labelled nucleotide, whose sequence is complementary to the mRNA or a region in a mRNA of interest. The assay allows the investigator to determine the stringency of the probing, i.e. to correlate the resulting signal(s) with sequence similarities.

As a first step, the nucleotide sequences of IKKs are compiled and inspected to identify regions that are unique to specific IKKs as well as regions that are shared among several, many, or all IKKs. Nucleotide sequences may be found in a depository of genetic information, e.g. GenBank, which is a wellknown resource. The inspection of the sequences may be aided by using computer programs that were developed to align several or many sequences, and in so doing highlighting regions of similarity or lack of the same. Many of these are presented and explained in great detail in e.g. Sequence Data Analysis Guidebook /edited by S.R.Swindell, Methods in Molecular Biology vol. 70 (1997), from Humana Press Inc. Totowa, New Jersey.

When sequences have been identified that are unique to an IKK, or respectively shared by several or many IKKs, oligonucleotide probes based on these sequences may be designed and synthesized. The use of such probes to detect mRNA is well established in the research community, see e.g. Basic DNA and RNA Protocols/edited by A.J.Harwood, Methods in Molecular Biology vol. 58 (1996), from Humana Press Inc. Totowa, New Jersey.

for a detailed description, and many commercial suppliers of biological research materials offer to synthesize specified oligonucleotides, e.g. Life Technologies.

In addition to oligonucleotide probes, mRNA extracted from the tissues and cell types of interest is required, preferably in a form ready to use in Northern analysis. Several companies offer such material, e.g. Invitrogen and Clontech. Briefly, they provide RNA extracted from a great many human and non-human tissues or cell types immobilized on membranes, as an array or size-fractionated.

In a next step, a detectable label needs to be attached to the oligonucleotide probe(s). The label is traditionally in the form of a radioactive isotope, but may to advantage be a chemiluminescent reagent or a fluorescent agent. See e.g. DNA Probes by Keller and Manak (1993), from Macmillan Publishers. Several companies offer reagents to

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label nucleotide probes, e.g. Ambion (Austin, Texas) and Molecular Probes (Eugene, Oregon).

The actual probing procedure involves contacting the immobilized mRNA (s) with the probe(s), washing away unbound probe(s) and detecting the signal(s) from the probe(s) that bound under the conditions tested, a positive signal indicating that the target(s) of the probe(s) was present in the sample(s) subjected to the test. In its simplest form, the test is "one-to-one", i.e. each sample of mRNA is exposed to each probe. However, it may be advantageous to exploit the sequence hierarchy of the IKKs, by first probing arrays of mRNA from multiple sources with family-specific probes, then examining first positives with isotype-specific probes, and then examining the secondary positives in detail with very specific probes. One could also multiplex the probing by adding different distungishable fluorescent labels to the probes, thus obtaining information from several probes in one experiment.

The outcome of the analysis is information regarding the expression pattern(s) of IKKs.

Based on their expression pattern(s) specific IKKs are then selected for further study, and genetic probes are constructed.

In general, a genetic probe, i.e. a "GeneX"-GFP fusion or a GFP-"GeneX" fusion, is constructed using PCR with "GeneX"-specific primers followed by a cloning step to fuse "GeneX" in frame with GFP. The fusion may contain a short vector derived sequence between "GeneX" and GFP (e.g. part of a multiple cloning site region in the plasmid) resulting in a peptide linker between "GeneX" and GFP in the resulting fusion protein.

The fusion may be made using ploymerase chain reaction techniques, which are common laboratory procedures, see e.g. PCR Protocols/edited by B.A.White, Methods in Molecular Biology vol. 15 (1993), from Humana Press Inc. Totowa, New Jersey.

30 In more detail, the steps involved include:

- Design of gene-specific primers. Inspection of the sequence of the gene allows design of gene-specific primers to be used in a PCR reaction. Typically, the top-strand primer encompasses the ATG start codon of the gene and the following ca. 20

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nucleotides, while the bottom-strand primer encompasses the stop codon and the ca. 20 preceding nucleotides, if the gene is to be fused behind GFP, i.e. a GFP-"GeneX" fusion. If the gene is to be fused in front of GFP, i.e. a "GeneX"-GFP fusion, a stop codon must be avoided. Optionally, the full length sequence of GeneX may not be used in the fusion, but merely the part which localizes and redistributes like GeneX in response to a signal.

In addition to gene-specific sequences, the primers contain at least one recognition sequence for a restriction enzyme, to allow subsequent cloning of the PCR product. The sites are chosen so that they are unique in the PCR product and compatible with sites in the cloning vector. Furthermore, it may be necessary to include an exact number of nucleotides between the restriction enzyme site and the gene-specific sequence in order to establish the correct reading frame of the fusion gene and/or a translation initiation concensus sequence. Lastly, the primers always contain a few nucleotides in front of the restriction enzyme site to allow efficient digestion with the enzyme.

-Identifying a source of the gene to be amplified. In order for a PCR reaction to produce a product with gene-specific primers, the gene-sequence must initially be present in the reaction, e.g. in the form of cDNA. The results of the extensive expression analysis performed previously will provide clear information regarding what tissue(s) are useful as source material. cDNA libraries from a great variety of tissues or cell types from various species are commercially available, e.g. from Clontech (Palo Alto), Stratagene (La Jolla) and Invitrogen (San Diego). Many genes are also available in cloned form from The American Type Tissue Collection (Virginia).

- Optimizing the PCR reaction. Several factors are known to influence the efficiency and specificity of a PCR reaction, including the annealing temperature of the primers, the concentration of ions, notably Mg²⁺ and K⁺, present in the reaction, as well as pH of the reaction. If the result of a PCR reaction is deemed unsatisfactory, it might be because the parameters mentioned above are not optimal. Various annealing temperatures should be tested, e.g. in a PCR machine with a built-in temperature gradient, available from e.g. Stratagene (La Jolla), and/or various buffer compositions should be tried, e.g. the OptiPrime buffer system from Stratagene (La Jolla).

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- Cloning the PCR product. The vector into which the amplified gene product will be cloned and fused with GFP will already have been taken into consideration when the primers were designed. When choosing a vector, one should at least consider in which cell types the probe subsequently will be expressed, so that the promoter controlling expression of the probe is compatible with the cells. Most expression vectors also contain one or more selective markers, e.g. conferring resistance to a drug, which is a useful feature when one wants to make stable transfectants. The selective marker should also be compatible with the cells to be used.

The actual cloning of the PCR product should present no difficulty as it typically will be a one-step cloning of a fragment digested with two different restriction enzymes into a vector digested with the same two enzymes. If the cloning proves to be problematic, it may be because the restriction enzymes did not work well with the PCR fragment. In this case one could add longer extensions to the end of the primers to overcome a possible difficulty of digestion close to a fragment end, or one could introduce an intermediate cloning step not based on restriction enzyme digestion. Several companies offer systems for this approach, e.g. Invitrogen (San Diego) and Clontech (Palo Alto).

Once the gene has been cloned and, in the process, fused with the GFP gene, the resulting product, usually a plasmid, should be carefully checked to make sure it is as expected. The most exact test would be to obtain the nucleotide sequence of the fusion-gene.

Once a DNA construct for a probe has been generated, its functionality and usefulness may be tested by subjecting it to the following tests:

- Transfecting it into cells capable of expressing the probe. The fluorescence of the cell is inspected soon after, typically the next day. At this point, two features of cellular fluorescence are noted:

The intensity and the sub-cellular localization.

The intensity should usually be at least as strong as that of unfused GFP in the cells. If it is not, the sequence or quality of the probe-DNA might be faulty, and should be carefully checked.

The sub-cellular localization is an indication of whether the probe is likely to perform well.

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If it localizes as expected for the gene in question, e.g. is excluded from the nucleus, it can immediately go on to a functional test. If the probe is not localized soon after the transfection procedure, it may be because of overexpression at this point in time, as the cell typically will have taken of very many copies of the plasmid, and localization will occur in time, e.g. within a few weeks, as plasmid copy number and expression level decreases. If localization does not occur after prolonged time, it may be because the fusion to GFP has destroyed a localization function, e.g. masked a protein sequence essential for interaction with its normal cellular anchor-protein. In this case the opposite fusion might work, e.g. if GeneX-GFP does not work, GFP-GeneX might, as two different parts of GeneX will be affected by the proximity to GFP. If this does not work, the proximity of GFP at either end might be a problem, and it could be attempted to increase the distance by incorporating a longer linker between GeneX and GFP in the DNA construct.

If there is no prior knowledge of localization, and no localization is observed, it may be because the probe should not be localized at this point, because such is the nature of the protein fused to GFP. It should then be subjected to a functional test.

In a functional test, the cells expressing the probe are treated with at least one compound known to perturb, usually by activating, the signalling pathway on which the probe is expected to report by redistributing itself within the cell.

If the redistribution is as expected, e.g. if prior knowledge tell that it should translocate from location X to location Y, it has passed the first critical test. In this case it can go on to further characterization and quantification of the response.

If it does not perform as expected, it may be because the cell lacks at least one component of the signalling pathway, e.g. a cell surface receptor, or there is species incompatibility, e.g. if the probe is modelled on sequence information of a human geneproduct, and the cell is of hamster origin. In both instances one should identify other cell types for the testing process where these potential problems would not apply.

If there is no prior knowledge about the pattern of redistribution, the analysis of the redistribution will have to be done in greater depth to identify what the essential and indicative features are, and when this is clear, it can go on to further characterization and quantification of the response.

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If no feature of redistribution can be identified, the problem might be as mentioned above, and the probe should be retested under more optimal cellular conditions.

Libraries for cloning of cDNA libraries in the present discovery plan are naturally related to the target tissues of the projects. For ultimately finding lead compounds useful in the treatment of astma the cloning libraries should preferably be obtained from one ore more of the following tissue or cells types: Bronchial smooth muscle, Lung microvascular endothelial cells, Eosinophil granulocytes, Th1 or 2 lymphocytes and alveolar macrophages. For ultimately finding lead compounds useful in the treatment of chronic inflammatory diseases the cloning libraries should preferably be obtained from one ore more of the following tissue or cell types: Th1 or 2 lymphocytes, T-lymphocytes, B-lymphocytes, Monocytes, Eosinophil granulocytes, Neutrophil granulocytes, Basophil granulocytes, Tissue specific macrophages (such as the liver Kupffer cells and skin Langhans cells), microvascular endothelial cells, vascular endothelial cells, antigen presenting cells, joint connective and synovial cells. For ultimately finding lead compounds useful in the treatment of depression the cloning libraries should preferably be obtained from one or more of the following tissue and cell types: Noradrenergic neurons from the brain, neurons form the brain. For ultimately finding lead compounds useful in the treatment of hyper- and hypotension the cloning libraries should preferably be obtained from one or more of the following tissue or cell types: vascular smooth muscle, vascular smooth muscle from resistance vessels on the arterial side of the vascular system, vascular smooth muscle from capacitance vessels on the venous side of the vascular system, vascular smooth muscle cells from small arteries, arterioles, venules or veins, smooth vascular cells lines such as T/G HA-VSMCA10 and A7r5.

The cells should always be of animal origin, most likely of mammalian origin and preferably of human origin. The cells could be derived from normal tissue or from tissue of an individual animal having a disease or condition of interest for the project. The cells may also be a mortal or immortalised cell line where the initial cell clone has been derived from a tissue or cell type as described above. Depending on the discovery project the cells of interest for screening assays will vary but may be chosen from the above mentioned categories.

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Once a genetic construct containing the protein of interest and the luminophore, from here on referred to as "the original fluorescent probe", has been transfected into a relevant cell type, as described above under 'preferred cell types for cloning libraries' the cells are monitored for the appearance of spatially distributed or randomly distributed intracellular fluorescence. Based on prior knowledge regarding the distribution of the actual protein different patterns can be expected. If for example previous studies have found the protein associated only with the particulate fraction of fractionated cells, it can be expected to find a spatial distribution of the original fluorescent probe to the plasma membrane, internal membrane/organelle structures or structural cytoplasmic elements such as microtubules and microfilaments. If on the other hand previous studies report that the protein has been found mostly in the soluble fraction of fractionated cells one can expect to find a homogenous or nonhomogenous distribution of the original fluorescent probe throughout the cytoplasm and perhaps also in the nucleus. For proteins where previous studies have found a mixed localisation to both the particulate and soluble fraction of fractionated cells any mixture in the two distribution patterns mentioned above for the original fluorescent probe can be expected. For proteins where no prior knowledge is at hand a simple cell fractionation and Western Blotting can be made, one can use immunohistochemistry of fixed cells of relevance or one can decide to rely on the distribution observed for the original fluorescent probe. At this stage of the project, a normal distribution pattern of the original fluorescent probe may be established after such studies as outlined above. The effects of physiologically important and relevant cellular activation on the distributed pattern of the original fluorescent probe is also established. It will also become evident if the pattern of distribution changes, i.e. if a redistribution of the original fluorescent probe occurs as a consequence of applying a physiologically important and relevant influence.

When a specific subcellular distribution of a GFP-based IKK probe has been identified, it may be advantageous to narrow down which part of the IKK is responsible for this effect. The advantage is twofold: It may suggest the design of peptide leads, and it may eventually aid in defining the binding partner. Knowledge of both partners involved in specific binding may aid in the selection of compound libraries to screen for inhibition of the specific binding.

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To identify the region of the IKK involved in specific binding, one may make GFP-based fusions with progressively shorter parts of the IKK, and examine the cellular distribution of these constructs. If there is prior knowledge of functional domains, one may start with the domain believed to confer specific binding to a subcellular structure. The generation of constructs to test may consist of selecting a particular part of the IKK to fuse to GFP, or it may involve the generation of in-frame deletions in the IKK part of the fusion. Both approaches have been widely used in molecular genetic studies.

When a region has been identified that appears responsible for conferring a specific subcellular distribution upon an IKK, the amino acid residues most important for this trait may be identified by a more detailed analysis, e.g. substituting them one by one with e.g. an alanin residue, a socalled Ala-scan, which also has been used extensively in molecular genetic studies.

To identify the identity of the cellular protein partaking in the specific distribution of the IKK, one may exploit the knowledge about the region of the IKK responsible for the subcellular distribution. E.g. one may use the region of the IKK as bait in a genetic two hybrid screen to pull out its binding partner. Several companies offer two hybrid systems, e.g. Life Technologies.

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The knowledge about the normal distribution of the original fluorescent probe is used to establish which part or which parts of the terminal (or entire) amino-acid sequence that is important for the attachment of this fluorescent probe to subcellular structures, giving it its specific spatially distributed pattern in the cell or cells, when such a pattern has been established as the normal distribution of this fluorescent probe. This is accomplished by creating new fluorescent probes where a systematic deletion of short N- or C-terminal or internal sequences (number of DNA bases) of the original fluorescent probe are made. These new shorter variants of the of the original fluorescent probe construct are transfected into the cells of interest and then the cells are examined for spatial distribution of the new fluorescent probes as described above for the original fluorescent probe. In those cells where the new fluorescent probe distribution pattern is different from the original fluorescent probe distribution pattern it is evident that part of the, or the entire, targeting sequence has been deleted. The

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intracellular binding sites.

DNA- or amino-acid sequence of the missing part therefore contains the structural information necessary for association of the original fluorescent probe with its

Peptides for inhibition of the established normal distribution of the original fluorescent probe are designed according to the hypothesis, that the deduced targeting sequence, or sequences, in the original fluorescent probe amino-acid sequence are the important sequences for the actual spatial distribution of the original fluorescent probe in intact living cells, is tested. This is done by producing peptides of identical amino-acid sequence as the deduced targeting sequence or parts thereof and introducing them into the cytoplasm, either by microinjection or transient or permanent permeabilisation, of cells containing the original fluorescent probe and thereafter monitoring the spatial distribution of the original fluorescent probe in the cells. If the deduced targeting sequence or sequences are of importance for the actual spatial distribution of the original fluorescent probe in intact living cells, the introduced peptides will self-associate with the anchoring sites for the original fluorescent probe and thereby disrupt the normal distribution of the original fluorescent probe. In order to have this effect, the introduction of the peptides should change the original distribution pattern so that a decrease in fluorescence of 10% or more, compared to the pattern before their introduction, can be detected. This is done by observing the same cells before and after administration of the peptides. When peptides that fulfil this criterion have been found they are called 'peptide leads' and will hereafter be referred to using this expression. These peptide leads can now be used as a basis for the design of organic molecules that can be used eventually to disrupt the spatial distribution of the original fluorescent probe but also as control compounds in screening assays.

In parallel to the above mentioned step wherein peptide leads are defined, the distribution pattern found for the original fluorescent probe is compared to the naturally occurring spatial distribution of the protein on which the original fluorescent probe is based. This may be accomplished by fixation of primary cells separated or within the tissue of interest and fixation of cells that contain the original fluorescent probe. Thereafter the protein is stained using ordinary immunocytochemical or

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immunohistochemical methods and the spatial distribution revealed by this staining procedure is compared to the spatial distribution of the original fluorescent probe. It is desirable, but not required, that a high degree of correlation between the two patterns obtained in this step can be observed.

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Establishment of a primary screening assay is normally done by making use of the cells of interest containing the original fluorescent probe as the basis for a screening assay. Depending on the knowledge acquired about the behaviour of the original fluorescent probe when subjecting the cells to physiologically relevant influences the assay procedure can be chosen: 1. If the fluorescent probe normally is targeted to specific sites and stay associated with these sites during stimulation of the intracellular pathway the assay should preferably be designed to detect dislocation of the original fluorescent probe from the targeting sites in mechanically intact or permeabilised living cells. This is an assay where the dislocation can be detected within minutes after application of an influence and the time frame for the detection and time for exposing the cells to an influence should be chosen to match this. 2. If the desire is to disrupt the actual targeting event rather than dislocate already targeted fluorescent probe the influence may need hours to produce a detectable response. The actual measurement, still of a change in the fluorescence or luminescence distribution pattern compared to the normal distribution pattern for the original fluorescent probe, may be made at two time points; before and after the influence has exerted any effect it may have. This is an assay where the effect of an influence may require several hours to produce a detectable response and the time frame for the detection and time for exposing the cells to an influence should be chosen to match this. 3. If the fluorescent probe normally redistributes between two intracellular sites upon activation of the intracellular pathway one may either want to disrupt the initial targeting or dislocate the original fluorescent probe from its initial or resting anchoring site. In this case procedure no. 1 above may be used. If the desire instead is to inhibit the association of the original fluorescent probe with the site it redistributes to during activation of the intracellular pathway the targeting sequence of this site should be in focus for the lead peptide generation. This is an assay where the redistribution may be detected within minutes after application of an influence and the time frame for the detection and time for exposing the cells to an influence should be chosen to match this. Furthermore,

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any influence applied to inhibit the targeting of the original fluorescent probe upon its redistribution may need to be added to the cells before activation of the intracellular pathway.

5 While the original fluorescent probe and peptide leads will be used in the actual primary screening assay, it is also desirable to have a counterscreen or counterscreens directed at protein isoforms that one does not wish to affect. In order to accomplish this, constructs are made for new fluorescent probes encoding the protein isoforms tagged with GFP. These constructs are subsequently transfected into the cells of interest. When the new fluorescent probes are expressed in the cells, some of the cells are chosen as the basis for new cell lines that can be used in the counterscreen or counterscreens.

Suitable probes for this purpose comprise DNA constructs encoding fusion polypeptides comprising forms of IKK α , IKK β , IKK γ or NIK and GFP.

In a preferred embodiment the DNA constructs will encode fusion polypeptides comprising isoforms of IKK β and GFP.

The cell lines established for the primary screen and the counterscreen or 20 counterscreens are used to establish peptide leads that more specifically dislocate the desired isoform of the protein of interest compared to other isoforms of the same protein. The peptide leads are introduced into the cells as described above and the changes in spatial distribution of the original and counterscreen fluorescent probes are quantified and dose-response relationships are established for each lead peptide. 25 Thereafter the dose-response relationships are compared. A peptide lead is considered specific for the original fluorescent probe if the dose of the peptide required to dislocate at least 10% of the fluorescent probes in the counterscreen or conterscreens are at least two times higher than the dose required to dislocate 10% of the original fluorescent probe. The lead peptides with the biggest dose difference when comparing 30 the primary and the counterscreen dose-response relationships are chosen as the basis for the next step in the discovery project.

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In one embodiment the primary screening assay and counterscreen or counterscreens are used to define specificity of the peptide leads by using a procedure that compares their ability to cause a dislocation, disruption of targeting or inhibition of redistribution of the original fluorescent probe in the primary screening assay to their ability to cause a dislocation, disruption of targeting or inhibition of redistribution of the new fluorescent probes in the counterscreen or counterscreens.

In a preferred embodiment the dose of a peptide lead required to cause a quantified dislocation, disruption of targeting or inhibition of redistribution of the original fluorescent probe of at least 10% in the primary screening assay is 50% or less of the dose required to cause a quantified dislocation, disruption of targeting or inhibition of redistribution of the new fluorescent probes of at least 10% in the counterscreen or counterscreens.

The invention provides for a specificity index which may be constructed describing a numerical relationship, with the primary screening asay result first, of the dose required to produce half-maximal effect in the primary assay compared to the dose required to produce half-maximal effect in the counterscreen or counterscreens.

In one embodiment the peptide leads chosen for further use in the discovery project have a specificity index of 1 to 2.

In another embodiment the peptide leads chosen for further use in the discovery project have a specificity index between 1 to 2 and 1 to 10.

In a further embodiment the peptide leads chosen for further use in the discovery project have a specificity index between 1 to 11 and 1 to 100.

In yet a further preferred embodiment the peptide leads chosen for further use in the discovery project have a specificity index better than 1 to 100.

Lead peptides are used to create and select libraries of small organic molecules that can be useful in screening assays to find bioactive substances useful as drugs to treat the condition or disease of interest for the project. In this step the amino-acid sequence information and other structural information about the lead peptide or peptides is used to extract information useful for finding and/or defining and synthesising bioactive organic molecules that can mimic the effect of the lead peptides on the normal spatial distribution pattern of the original fluorescent probe. Peptide leads selected by the

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discovery project are used to design and assemble compound libraries based on the structural and chemical information inherent in the lead peptides using prior chemical knowledge and computational chemistry approaches so that the compounds have a structure that give them the ability to interact with or bind to the targeting sequence of IKKB, thereafter testing the compound libraries at a concentration of 10 or 100 micromolar of each compound in the primary screening assay.

When the libraries of compounds have been defined and are at hand it is time to initiate primary screening. In this procedure, cells containing the original fluorescent probe are contacted with the compounds. The compounds are all tested at just one or a few concentrations, typically 10 and 100 micromolar, in a highly parallel fashion using a quantitative fluorescence redistribution assay. Compounds that cause a change in the quantitated response (the response scale defined by the range 0 (no change in redistribution) - 100%) of the assay by more than a predetermined value, typically between 10 and 100%, are considered to be "primary hits". The primary hits are then further characterised: 1. for potency by establishing a dose-response relationship compared to the lead peptide(s) using the primary screening assay 2. for selectivity by establishing a dose-response relationship in the counterscreen or counterscreens. Primary hits that have low potency, typically when the half-maximal effect of the compound in the primary assay is achieved at a concentration of the compound between 10 and 100 micromolar, may not need testing in the counterscreen or counterscreens since the likelihood that they will be used beyond this step in the discovery project is small. Primary hits that have equal or lower potency in the primary screening assay compared to the counterscreen or counterscreens are regarded as non-selective and the likelihood that they will be used beyond this step in the discovery project is small. Primary hits that display some degree of selectivity, typically half maximal effect in the primary screening assay at a concentration 50% or less of the concentration that gives half maximal effect in the counterscreen or counterscreens are considered interesting as the basis for further chemical synthesis or construction of new libraries of compounds and will hereafter be referred to as "primary lead compounds".

Compounds that cause a change in the quantitated response, with a response scale from 0 to 100% based on the absence of a response and the maximal response

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observed with the peptide leads in the primary screening assay, of the assay by more than a predetermined value are selected and called "primary hits".

In one embodiment the predetermined value is 10%.

In another embodiment the predetermined value is 50%.

5 In yet another embodiment the predetermined value is 70%.

In one embodiment the primary hits are further characterised for potency (as defined herein) and maximal effect by establishing a dose-response relationship (as defined herein) and comparing that to the effects of the lead peptides using the primary screening assay and for selectivity (as defined herein) by establishing a dose-response relationship in the counterscreen or counterscreens.

Primary hits may be deselected by the discovery project when they display a half-maximal potency at a dose corresponding to a concentration of more than 10 micromolar or because they display a selectivity index less than 1 to 2.

Primary hits may be selected by the discovery project when they display a half-maximal potency at a dose corresponding to a concentration of 10 micromolar or less or because they display a selectivity index higher than 1 to 2, the compounds hereafter also referred to as "primary lead compounds".

A Structure-Activity Relationship is built by iterations of compound library composition and screening to define drug candidate leads. This step is included to further improve the possibilities of finding bioactive compounds with desirable properties for treatment of the diseases or conditions of interest to the project. The primary lead compounds are here used to provide chemical structural information that can be used as the basis for composition or chemical synthesis of new, directed, compound libraries. By systematic chemical modification of part of the structure of one or more primary lead compounds new libraries are assembled. These new libraries of compounds are also investigated using the primary screening assay and counterscreen or counterscreens. Preferably, dose-response relationships are recorded for each chemical modification of the primary lead compound and compared to the primary lead compound itself. Thereby, a structure-activity relationship, hereafter referred to as "SAR", is established. Among the new compounds, the ones that in this step has the best combination of potency and specificity are chosen either as the basis for a new round of compound library synthesis or composition or, as the final step of

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the SAR building process, as compounds that will be further for actual pharmacoloical effects in assay systems and animals that are relevant to the underlying physiological and pathophysiological processes of interest to the project. The latter compounds will hereafter be referred to as "drug candidate leads".

In one embodiment drug candidate leads have a half-maximal potency at a dose corresponding to a concentration of less than 1 micromolar and a selectivity index higher than 1 to 2.

In one embodiment the drug candidate leads have a half-maximal potency at a dose corresponding to a concentration of less than 1 micromolar and a selectivity index higher than 1 to 10.

In one embodiment the drug candidate leads have a half-maximal potency at a dose corresponding to a concentration of less than 1 micromolar and a selectivity index higher than 1 to 100.

In one embodiment the drug candidate leads have a half-maximal potency at a dose corresponding to a concentration of less than 0,1 micromolar and a selectivity index higher than 1 to 2.

In a preferred embodiment the drug candidate leads have a half-maximal potency at a dose corresponding to a concentration of less than 0,1 micromolar and a selectivity index higher than 1 to 10.

In another preferred embodiment the drug candidate leads have a half-maximal potency at a dose corresponding to a concentration of less than 0,1 micromolar and a selectivity index higher than 1 to 100.

Drug candidate leads may be further characterised *in vitro* in tissue based, cell based and biochemical assays for efficacy and toxicity. There are many ways to test efficacy of a drug candidate lead. Preferably, the drug candidate lead is tested in assay systems with high relevance to the underlying physiological and pathophysiological processes involved in the pathogenesis and pathophysiology of the disease or condition of interest to the project. Likewise, the drug candidate leads are tested for toxic effects, preferably testing for genetic effects (influence on the integrity and arrangement of DNA), metabolic effects (influence on cellular metabolic processes) and cytotoxic effects (influence on cell integrity and organelle integrity). There is a high likelihood that drug candidate leads, that do not show appropriate efficacy or that display toxicity

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will not be used beyond this step in the discovery project because it is expected that such compounds are less suitable as actual drugs to be used in an animal.

In one embodiment drug candidate leads chosen by the discovery project are tested *in vitro* for efficacy (as defined herein), in assay systems with high degree of relevance to the underlying physiological and pathophysiological processes involved in inflammatory diseases, and for toxicity (as defined herein), preferably testing for genetic, metabolic and cytotoxic effects, whereafter the drug candidate leads that display the best efficacy and the least, or no, indications of toxicity are chosen to be the candidates that will enter testing in animals.

In another embodiment drug candidate leads chosen by the discovery project are tested *in vitro* for efficacy (as defined herein), in assay systems with high degree of relevance to the underlying physiological and pathophysiological processes involved in inflammatory airway diseases, and for toxicity (as defined herein), preferably testing for genetic, metabolic and cytotoxic effects, whereafter the drug candidate leads that display the best efficacy and the least, or no, indications of toxicity are chosen to be the candidates that will enter testing in animals.

In another embodiment drug candidate leads chosen by the discovery project are tested *in vitro* for efficacy (as defined herein), in assay systems with high degree of relevance to the underlying physiological and pathophysiological processes involved in inflammatory joint diseases, and for toxicity (as defined herein), preferably testing for genetic, metabolic and cytotoxic effects, whereafter the drug candidate leads that display the best efficacy and the least, or no, indications of toxicity are chosen to be the candidates that will enter testing in animals.

In another embodiment drug candidate leads chosen by the discovery project are tested *in vitro* for efficacy (as defined herein), in assay systems with high degree of relevance to the underlying physiological and pathophysiological processes involved in inflammatory bowel diseases, and for toxicity (as defined herein), preferably testing for genetic, metabolic and cytotoxic effects, whereafter the drug candidate leads that display the best efficacy and the least, or no, indications of toxicity are chosen to be the candidates that will enter testing in animals.

In another embodiment drug candidate leads chosen by the discovery project are tested *in vitro* for efficacy (as defined herein), in assay systems with high degree of relevance to the underlying physiological and pathophysiological processes involved

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in autoimmune diseases, and for toxicity (as defined herein), preferably testing for genetic, metabolic and cytotoxic effects, whereafter the drug candidate leads that display the best efficacy and the least, or no, indications of toxicity are chosen to be the candidates that will enter testing in animals.

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In another embodiment drug candidate leads chosen by the discovery project are tested *in vitro* for efficacy (as defined herein), in assay systems with high degree of relevance to the underlying physiological and pathophysiological processes involved in depression, and for toxicity (as defined herein), preferably testing for genetic, metabolic and cytotoxic effects, whereafter the drug candidate leads that display the best efficacy and the least, or no, indications of toxicity are chosen to be the candidates that will enter testing in animals.

Drug candidate leads are tested for toxic and unwanted effects *in vivo* in animals such as mice and rats. The drug candidate leads are also tested for efficacy in animals that have a disease or condition with high degree of relevance to the disease or condition of interest to the project. The drug candidate leads may also be tested for efficacy in animals which have been treated in a way that make them experience a disease or condition with high degree of relevance to the disease or condition of interest to the project. Drug candidate leads that display efficacy in one or more of such animal tests and that does not display any apparent toxicity at a dosage level, preferably 2 –10 times higher than the level that gives satisfactory efficacy are chosen to be the final drug candidates that should be considered for further animal testing and initial testing in humans. These compounds are hereafter referred to as "discovery project leads".

In one embodiment drug candidate leads chosen by the discovery project are tested for efficacy (as defined herein), in healthy animals and animals with a condition with high degree of relevance to the underlying physiological and pathophysiological processes involved in inflammatory diseases, and for toxicity (as defined herein) and unwanted side effects, whereafter the drug candidate leads that display the best efficacy and the least, or no, indications of toxicity or unwanted side effects are chosen to be the candidates, called discovery project leads, that will enter further testing in animals and testing in humans.

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In one embodiment drug candidate leads chosen by the discovery project are tested for efficacy (as defined herein), in healthy animals and animals with a condition with high degree of relevance to the underlying physiological and pathophysiological processes involved in inflammatory airway diseases, and for toxicity (as defined herein) and unwanted side effects, whereafter the drug candidate leads that display the best efficacy and the least, or no, indications of toxicity or unwanted side effects are chosen to be the candidates, called discovery project leads, that will enter further testing in animals and testing in humans.

In one embodiment drug candidate leads chosen by the discovery project are tested for efficacy (as defined herein), in healthy animals and animals with a condition with high degree of relevance to the underlying physiological and pathophysiological processes involved in inflammatory joint diseases, and for toxicity (as defined herein) and unwanted side effects, whereafter the drug candidate leads that display the best efficacy and the least, or no, indications of toxicity or unwanted side effects are chosen to be the candidates, called discovery project leads, that will enter further testing in animals and testing in humans.

In one embodiment drug candidate leads chosen by the discovery project are tested for efficacy (as defined herein), in healthy animals and animals with a condition with high degree of relevance to the underlying physiological and pathophysiological processes involved in inflammatory bowel diseases, and for toxicity (as defined herein) and unwanted side effects, whereafter the drug candidate leads that display the best efficacy and the least, or no, indications of toxicity or unwanted side effects are chosen to be the candidates, called discovery project leads, that will enter further testing in animals and testing in humans.

In one embodiment drug candidate leads chosen by the discovery project are tested for efficacy (as defined herein), in healthy animals and animals with a condition with high degree of relevance to the underlying physiological and pathophysiological processes involved in autoimmune diseases, and for toxicity (as defined herein) and unwanted side effects, whereafter the drug candidate leads that display the best efficacy and the least, or no, indications of toxicity or unwanted side effects are chosen to be the candidates, called discovery project leads, that will enter further testing in animals and testing in humans.

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In one embodiment drug candidate leads chosen by the discovery project are tested for efficacy (as defined herein), in healthy animals and animals with a condition with high degree of relevance to the underlying physiological and pathophysiological processes involved in depression, and for toxicity (as defined herein) and unwanted side effects, whereafter the drug candidate leads that display the best efficacy and the least, or no, indications of toxicity or unwanted side effects are chosen to be the candidates, called discovery project leads, that will enter further testing in animals and testing in humans.

The administration route of any of the compounds of the invention may be of any suitable route which leads to a concentration in the blood corresponding to a therapeutic concentration by the oral route, the parenteral route, the cutaneous route, the nasal route, the rectal route, the vaginal route and the ocular route. It should be clear to a person skilled in the art that the administration route is dependant on the compound in question, particularly, the choice of administration route depends on the physico-chemical properties of the compound together with the age and weight of the patient and on the particular disease and the severity of the same.

The compounds of the invention may be contained in any appropriate amount in a pharmaceutical composition, and are generally contained in an amount of about 1-95% by weight of the total weight of the composition. The composition may be in form of, e.g., tablets, capsules, pills, powders, granulates, suspensions, emulsions, solutions, gels including hydrogels, pastes, ointments, creams, plasters, drenches, delivery devices, suppositories, enemas, injectables, implants, sprays, aerosols and in other suitable form. The pharmaceutical compositions may be formulated according to conventional pharmaceutical practice, see, e.g., "Remington's Pharmaceutical Sciences" and "Encyclopedia of Pharmaceutical Technology".

Pharmaceutical compositions according to the present invention may be formulated to release the active compound substantially immediately upon administration or at any substantially predetermined time or time period after administration. The latter type of compositions are generally known as controlled release formulations. Controlled release formulations may also be denoted "sustained release", "prolonged release", "programmed release", "time release", "rate-controlled" and/or "targeted release" formulations.

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In the present context every pharmaceutical composition is an actual drug delivery system, since upon administration it presents the active drug substance to the body of the organism.

The compounds of the invention are preferably administered in an amount of about 0.1-30 mg per kg body weight per day, such as about 0.5-15 mg per kg body weight per day. The compound in question may be administered orally in the form of tablets, cap-sules, elixirs or syrups, or rectally in the form of suppositories. Parenteral administration of the compounds of the invention, is suitably performed in the form of saline solutions of the compounds or with the compound incorporated into liposomes. In cases where the compound in itself is not sufficiently soluble to be dissolved, an acid addition salt of a basic compound can be used, or a solubilizer such as ethanol can be applied.

Oral administration. For compositions adapted for oral administration for systemic use, the dosage is normally 1 mg to 1 g per dose administered 1-4 times daily for 1 week, 12 months or even lifelong depending on the disease to be treated.

Rectal administration. For compositions adapted for rectal a somewhat higher amount of compound is usually preferred, i.e. from approximately 1 mg to 100 mg per kg body weight per day.

Parenteral administration. For parenteral administration a dose of about 0.1 mg to about 50 mg per kg body weight per day is convenient. For intravenous administration a dose of about 0.1 mg to about 20 mg per kg body weight per day. For intraarticular administration a dose of about 0.1 mg to about 20 mg per kg body weight per day is usually preferable. For parenteral administration in general, a solution in an aqueous medium of 0.5-2% or more of the active ingredients may be employed.

<u>Cutaneous administration</u>. For topical administration on the skin a dose of about 1 mg to about 5 g administered 1-10 times daily is usually preferable.

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EXAMPLES

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Probes for detection of IKK redistribution. These are specific IKK subunit variants fused to a GFP. As examples, the following three subunits have been chosen: IKK α (GenBank Acc.no. AF009225) , IKK β (GenBank Acc. No. AF031416) and IKK γ (GenBank Acc. No. AF074382).

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Inspection of the scientific literature indicates that IKK β dissociates transiently from the IKAP complex during activation, and so becomes the first choice for a probe to detect redistribution.

To construct the IKKβ-GFP fusion. IKKβ sequences are amplified using PCR according to standard protocols with the specific primers listed below. The PCR product is digested with restriction enzymes Hind3 and Acc651, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Hind3 and Acc651. This produces an IKKβ-EGFP fusion under the control of a CMV promoter (SEQ.ID.NOs.1 and 2).

The top primer includes specific sequences following the ATG and a cloning site (EcoR1). The bottom primer includes specific C-terminal sequences minus the stop codon, an Acc65I cloning site, and two extra nucleotides to preserve the reading frame in EGFP-N1.

20 IKKβ-top (SEQ. ID NO. 3): 5'-GTAAGCTTACATGAGCTGGTCACCTTCCCTG-3'

IKKβ-bottom (SEQ. ID NO. 4): 5'-GTGGTACCCATGAGGCCTGCTCCAG-3'

The resulting plasmids are transfected into a suitable cell line. The subcellular distribution of the probes is examined carefully by fluorescence microscopy, both under resting conditions, and upon activation, e.g. with TNFalpha.

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REFERENCES

- Baeuerle, P.A., Baltimore, D. (1988) I-kappaB: a specific inhibitor of the NF-kappaB transcription factor. Science 242:540-546.
- Baeuerle, P.A., Baltimore, D. (1989) A 65-kappaD subunit of NF-kappaB is required for inhibition of NF-kappaB by I-kappaB. Genes and Dev. 3:1689-1698.
 - Baldwin, A.S. Jr (1996) The NF-kappaB and I-kappaB proteins: new discoveries and insights. Annu. Rev. Immunol. 14:649-683.
 - Beg, A.A., Finco, T.S., Nantermet, P.V., Baldwin, A.S. (1993) Tumor necrosis factor and interleukin-1 lead to phosphorylation and loss of Ikappa-Bα - a mechanism of NF-kappaB activation. Mol. Cell. Biol. 13:3301-3310.
- Bird, T.A., Schooley, K., Dower, S.K., Hagen, H., Virca, G.D., (1997) Activation of nucear transcription factor NF-kappaB by interleukin-1 is accompanied by casein kinase II-mediated phosphorylation of the p65 subunit. J. Biol. Chem. 272:32606-32612.
- Blackwell, T.S., Christman, J.W. (1997) The role of nuclear factor-kappa B in cytokine gene regulation. Am. J. Resp. Cell and Mol. Biol. 17:3-9.
- Bourcier, T., Sukhove, G., Libby P. (1997) The nuclear factor kappa-B signalling pathway participates in dysregulation of vascular smooth muscle cells in vitro and in human atherosclerosis. J.Biol. Chem. 272:15817-15824
 - Bours, V., Dejardin, E., Bonizzi, G., Merville, M.P., Piette, J. (1998). The NF-kappaB transcription factor role in oncogenesis and in response to anticancer therapeutics. Medecine Sciences 14:566-571
 - Brattsand, R., Linden, M. (1996) Cytokine modulation by glucocorticoids: mechanisms and actions in cellular studies. Alimentary Pharmacol. Therapeutics 10:81-90

Chen, ZJ; Parent, L; Maniatis, T. (1996) Site-specific phosphorylation of i-kappa-b-alpha by a novel ubiquitination-dependent protein-kinase activity. Cell 84: 853-862.

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- Cogswell, P.C., Schienman, R.I., Baldwin, A.S. (1993) Promoter of the human NF-kappaB p50/p105 gene regulation by NF-kappaB subunits and by c-rel. J. Immunol. 150:2794-2804.
- Cohen, L.; Henzel, W.J.; Baeuerle, P.A. (1998) IKAP is a scaffold protein of the IkB kinase complex. Nature 395: 292-296.
 - Ferrari, D., Wesselborg, S., Bauer, M.K.A., Schulze-Osthoff, K., (1997) Extracellular ATP activates transcription factor NF-kappaB through the P2Z purinoreceptor by selectively targeting NF-kappaB p65 (RelA). J. Cell Biol. 139:1635-1643
 - Ghosh, S., et al., (1990) Cloning of the p50 DNA-binding subunit of NF-kappaB: homology to rel and dorsal. Cell:62:1007-1018.
- 20 Giri, D.K., Aggarwal, B.B. (1998) Constitutive activation of NF-kappaB causes resistance to apoptosis in human cutaneous t-cell lymphoma HuT-78 cells autocrine role of tumor-necrosis factor and reactive oxygen species. J. Biol. Chem. 273:14008-14014
- 25 Grumont, R.J., Gerondakis, S. (1994) The subunit composition of NF-kappaB complexes changes during B-cell development. Cell Growth and Diffn. 5:1321-1331.
- Hallahan, D., Clark, E.T., Kuchibholta, J., Gewertz, B.L., Collins, T. (1995) E-selectin gene induction by ionizing-radiation is independent of cytokine induction. Biochem. Biophys. Res. Comm. 217:784-795.

- Hattori, Y., Akimoto, K., Murakami, Y., Kasai, K. (1997) Pyrrolidine dithiocarbamate inhibits cytokine-induced VCAM-1 gene expression in rat cardiac myocytes. Mol. Cell. Biochem. 177177-181.
- Hayashi, T., Sekine, T., Okamoto, T. (1993) Identification of a new serine knase that activates NF-kappaB by direct phosphorylation. J. Biol. Chem. 268:26790-26795.
- Hiramoto, M., et al., (1998) Nuclear targeted suppression of NF-kappaB activity by the quinone derivative E3330. J. Immunol. 160:810.819.
 - Jourd'heuil, D., Morise, Z., Conner, E.M., Grisham, M.B. (1997) Oxidants, transcription factors and intestinal inflammation. J. Clin. Gastroenterol. 25:S61-S72.
 - Lezoual'ch, F., Behl, C. (1998) Transcription factor NF-kappaB: Friend or foe of neurons? Mol. Psychiatry 3:15-20
- Makarov, S.S., Johnston, W.N., Olsen, J.C., Watson, J.M., Mondal, K., Rinehart, C.,
 Haskill, J.S. (1997) NF-kappaB as a target for anti-inflammmatory gene therapy
 suppression of inflammatory responses in monocytic and stromal cells by
 stable gene-transfer of I-kappaBα cDNA. Gene Therapy 4:846-852
- Mukaida, N. Mahe, Y., Matsushima, K., (1990) Co-operative interaction of NF-kappaB and cis-regulatory enhancer binding protein-like factor binding elements in activation the interleukin-8 gene by pro-inflammatory cytokines. J. Biol. Chem. 265:21128-21133.
- Naumann, M., Wulczyn, F.G., Scheidereit, C., (1993) The NF-kappaB precursor p105 and the proto-oncogene product bcl-3 are I-kappaB molecules and control nuclear translocation of NF-kappaB. EMBO J. 12:213-222

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- Neumann, M., Marienfeld, R., Serfling, E. (1997) Rel/NF-kappaB transcription factors and cancer oncogenesis by dysregulated transcription. Int. J. Oncology 11:1335-1347
- Nolan, G.P., Fujita, T., Bhatia, K., Huppi, C., Liou, H.C., Scott, M.L., Baltimore, D., (1993) The bcl-3 proto-oncogene encodes a nuclear I-kappaB-like molecule that preferentially interacts with NF-kappaB p50 and p52 ina phosphorylation-dpendent manner. Mol. Cell. Biol., 13:3557-3566
- O'Neill, L.A., Kaltschmidt, C. (1997) NF-kappaB: a crucial transcription factor for glial and neuronal cell function. Trends Neurosci. 20:252-258.
 - Rothwarf, D.M., Zandi, E., Natoli, G., Karin, M. (1998) IKK-g is an essential regulatory subunit of the IkB kinase complex. Nature 395: 297-300.
 - Schulze-Osthoff, K., Ferrari, D., Riehemann, K., Wesselborg, S. (1997) Regulation of NF-kappaB activation by MAP kinase cascades. Immunobiol. 198:35-49.
- Sen, R., Baltimore, D., 1986. Inducibility of kappa immunoglobulin enhancer-binding protein NF-kappaB by a post-translational mechanism. Cell 47:921-928.
 - Smith, R.S., Smith, T.J., Blieden, T.M., Phipps, R.P. (1997) Fibroblasts as sentinel cells. Synthesis of chemokines and regulation of inflammation. Am. J. Path. 151:317-322.
 - Watanabe, N., Iwamura, T., Shinoda, T., Fujita, T. (1997) Regulation of NF-kappaB 1 proteins by the candidate oncoprotein bcl3 generation of NF-kappaB homodimers from the cytoplasmic pool of p50-p105 and nuclear translocation. EMBO J. 16:3609-3620.
 - Wulczyn, F.G., Krappmann, D., Scheidereit, C., (1996) the NF-kappaB B/Rel and I-kappaB gene families: mediators of immune response and inflammation. J. Mol. Medicine 74:749-769.

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Zhong, H.H., Voll, R.E., Ghosh, S., (1998) Phosphorylation of NF-kappaB p65 by PKA stimulates transcriptional activity by promoting a novel bivalent interaction with the coactivator CBP/p300. Molecular Cell 5:661-671.

CLAIMS

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- 1. A method for preventing or treating, in an animal in need thereof, an adverse condition which may be reduced or abolished by modulating the activity of one or more I-kappaB kinases, the method comprising modulating the specific effectiveness of the I-kappaB kinase by modulating their spatial distribution within cells of the animal.
- 2. A method according to claim 1, wherein the I-kappaB kinase is selected from the group consisting of I-kappaB kinase α , I-kappaB kinase β , I-kappaB kinase γ and NIK.
- 3. A method according to claim 1 or 2, wherein the I-kappaB kinase is I-kappaB
 15 kinase β.
 - 4. A method according to any of claims 1-3, wherein the animal is a mammal.
 - 5. A method according to claim 4, wherein the mammal is a human being.
 - 6. A method according to any of claims 1-5, wherein the modulation of the specific effectiveness of the I-kappaB kinase is a dislocation from a native location within the cell.
- 7. A method according to any of claims 1-5, wherein the modulation of the specific effectiveness of the I-kappaB kinase involves a disruption of the targeting of the I-kappaB kinase to a native location within the cell.
- 8. A method according to any of claims 1-5, wherein the modulation of the specific effectiveness of the I-kappaB kinase involves interference with the redistribution of the I-kappaB kinase, the redistribution being associated with an increase or a decrease in the specific effectiveness of the I-kappaB kinase.

- 9. A method according to any of claims 1-8, wherein the adverse condition is an inflammatory diseases such as chronic inflammation.
- 10. A method according to claim 9, wherein the adverse condition is chronic
 5 inflammatory airway diseases such as asthma and chronic bronchial hyperreactivity of non-asthma etiology.
 - 11. A method according to claim 9, wherein the adverse condition is chronic inflammatory joint diseases such as rheumatoid arthritis and pelvospondylitis.
 - 12. A method according to claim 9, wherein the adverse condition is chronic inflammatory bowel diseases such as ulcerative colitis and Crohn's disease.
- 13. A method according to any of claims 1-9, wherein the adverse condition is
 autoimmune diseases with chronic inflammation such as rheumatoid arthritis, diabetes
 mellitus type I, systemic lupus erythematosus, myasthenia gravis, Hashimoto's
 thyreoiditis, Graves' disease and immune thrombocytopenic purpura.
- 14. A method according to any of claims 1-8, wherein the adverse condition involves a disregulation of the immune system such as acute respiratory distress syndrome (ARDS) and septic shock.
 - 15. A method according to any of claims 1-8, wherein the adverse condition is allergy.
- 16. A method according to any of the preceding claims, wherein the modulation of the specific effectiveness of the I-kappaB kinase is performed by exposing cells, in the animal in which dislocation, disruption of targeting, or interference with redistribution of a I-kappaB kinase may take place, to the influence of a substance which modulates the spatial distribution of the I-kappaB kinase in the cells.
 - 17. A method according to claim 16, wherein the substance is one which, in a quantitative fluorescence redistribution assay designed to monitor dislocation of I-

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kappaB kinase, causes dislocation of at least 10% of otherwise natively located I-kappaB kinase within the cell at a concentration of the substance of 100 micromolar.

- 18. A method according to claim 17, wherein at least 50% of otherwise natively
 located I-kappaB kinase is dislocated within the cell at a concentration of the substance of 100 micromolar.
 - 19. A method according to claim 17, wherein at least 70% of otherwise natively located I-kappaB kinase is dislocated within the cell at a concentration of the substance of 100 micromolar.
 - 20. A method according to claim 17, wherein at least 90% of otherwise natively located I-kappaB kinase is dislocated within the cell at a concentration of the substance of 100 micromolar.
 - 21. A method according to claim 16, wherein the substance is one which, in a quantitative fluorescence redistribution assay, designed to monitor targeting of I-kappaB kinase, reduces targeting of the I-kappaB kinase to its native location within the cell by at least 10% at a concentration of the substance of 100 micromolar.
 - 22. A method according to claim 21, wherein the substance reduces targeting of the I-kappaB kinase to its native location within the cell by at least 50% at a concentration of the substance of 100 micromolar.
- 23. A method according to claim 21, wherein the substance reduces targeting of the I-kappaB kinase to its native location within the cell by at least 70% at a concentration of the substance of 100 micromolar.
- 24. A method according to claim 21, wherein the substance reduces targeting of the I-30 kappaB kinase to its native location within the cell by at least 90% at a concentration of the substance of 100 micromolar.

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25. A method according to claim 16, wherein the substance is one which, in a quantitative fluorescence redistribution assay, designed to monitor changes in redistribution caused by an influence, causes a reduction in the induced redistribution by at least 10% of the normal maximum redistribution at a concentration of the substance of 100 micromolar.

- 26. A method according to claim 25, wherein the substance causes a reduction in the induced redistribution of the I-kappaB kinase by at least 50% of the normal maximum redistribution at a concentration of the substance of 100 micromolar.
- 27. A method according to claim 25, wherein the substance causes a reduction in the induced redistribution of the I-kappaB kinase by at least 70% of the normal maximum redistribution at a concentration of the substance of 100 micromolar.
- 15 28. A method according to claim 25, wherein the substance causes a reduction in the induced redistribution of the I-kappaB kinase by at least 90% of the normal maximum redistribution at a concentration of the substance of 100 micromolar.
- 29. A method according to any of claims 16-28, wherein the substance is an organic compound having a molecular weight of at the most 1200 Da.
 - 30. A method according to any of claims 16-28, wherein the substance is an organic compound having a molecular weight of at the most 900 Da.
- 31. A method according to any of claims 16-28, wherein the substance is an organic compound having a molecular weight of at the most 600 Da.
 - 32. A method according to any of claims 16-28, wherein the substance is an organic compound having a molecular weight of at the most 300 Da.
 - 33. A method according to any of claims 16-32, wherein the substance is a peptide.

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34. A method according to any of claim 16-32, wherein the substance is a carbon-containing non-peptide.

- 35. A method according to any of claims 16-32, wherein the organic compound is a compound having one or more chemical domains capable of interacting with one or more functional groups of the targeting sequence of the native anchoring site of the I-kappaB kinase.
- 36. A method according to claim 35, wherein the organic compound is a compound having at least two chemical domains capable of interacting with at least two functional groups of the targeting sequence of the native anchoring site for the I-kappaB kinase.
- 37. A method according to claim 35, wherein the organic compound is a compound having at least three chemical domains capable of interacting with at least three functional groups of the targeting sequence of the native anchoring site for the I-kappaB kinase.
- 38. A method according to any of claims 16-34, wherein the organic compound is a compound having one or more chemical domains capable of interacting with one or more functional groups of the targeting sequence of the I-kappaB kinase.
- 39. A method according to claim 38, wherein the organic compound is a compound having at least two chemical domains capable of interacting with at least two
 25 functional groups of the targeting sequence of the I-kappaB kinase.
 - 40. A method according to claim 38, wherein the organic compound is a compound having at least three chemical domains capable of interacting with at least three functional groups of the targeting sequence of the I-kappaB kinase.
 - 41. A method according to any of claims 16-40, wherein the organic compound is a weak acid in that it is a neutral molecule that can reversible dissociate into an anion (a negatively charged molecule) and a proton (a hydrogen ion).

42. A method according to claims 16-40, wherein the organic compound is a weak base in that it is a neutral molecule that can form a cation (a positively charged molecule) by combining with a proton (a hydrogen ion).

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43. A method according to any of claims 35-42, wherein the functional groups of the targeting sequences include functional groups selected from the group consisting of: methyl-, isopropyl-, isobutyl-, hydroxyl-, thiol-, benzyl-, benzyloyl-, methylindolyl-, methylimidazolyl-, amine-, imine-, carboxyl- and acetamide-groups as parts of amino acids in the targeting sequences.

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44. A method according to any of claims 16-43, wherein the exposure of the animal to the influence of a substance is performed by administering an effective amount of the substance to the animal.

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45. A method according to claim 44, wherein the exposure of the animal to the influence of the substance is performed by administering an effective amount of the substance via the intravenous route of administration to the animal.

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46. A method according to claim 44, wherein the exposure of the animal to the influence of the substance is performed by administering an effective amount of the substance via the oral route of administration to the animal.

47. A method according to claim 44, wherein the exposure of the animal to the influence of the substance is performed by administering an effective amount of the substance via the pulmonary route of administration to the animal.

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48. A method according to claim 44, wherein the exposure of the animal to the influence of the substance is performed by administering an effective amount of the substance via the rectal route of administration to the animal.

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49. A method according to claim 44, wherein the exposure of the animal to the influence of the substance is performed by administering an effective amount of the substance via the transdermal route of administration to the animal.

- 50. A method according to any of claims 17-49, wherein the quantitative fluorescence 5 redistribution assay consists of cells selected from the group of bronchial smooth muscle cells and immortal cell lines derived from such cells, smooth muscle cells and immortal cell lines derived from such cells, neutrophil or eosinophil granulocytes and immortal cell lines derived from such cells, T-lymphocytes and immortal cell lines derived from such cells, monocytes and immortal cell lines derived from such cells, 10 mast cells and immortal cell lines derived from such cells, lung microvascular endothelial cells and immortal cell lines derived from such cells, alveolar epithelial cells and immortal cell lines derived from such cells, and alveolar macrophages and immortal cell lines derived from such cells, transfected with a nucleotide construct encoding a fluorescent probe comprising as the biologically active polypeptide either 15 I-kappaB kinase α , I-kappaB kinase β , I-kappaB kinase γ or NIK, or an I-kappaB kinase splice variant cloned from bronchial smooth muscle cells, lung microvascular endothelial cells, alveolar epithelial cells, neutrophil or eosinophil granulocytes, Th1 lymphocytes, Th2 lymphocytes, B-lymphocytes, monocytes, mast cells, or alveolar macrophages, transfected in such a way, that the construct is expressed by the cells. 20
 - 51. A method according to claim 50, wherein the quantitative fluorescence redistribution assay is a primary screening assay used in a discovery project
- 52. A method according to any of claim 50 or 51, wherein the cells are derived from an animal.
 - 53. A method according to claim 52, wherein the cells are derived from a mammal such as a human.
 - 54. A method according to any of claims 50-53, wherein the fluorescent probe redistributes after the cells have been subjected to a physiologically important and relevant influence that is relevant to the intercellular signalling pathway wherein the I-

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kappaB kinase is an integral part, so that both the normal pattern of spatial distribution and possible redistribution of the fluorescent probe can be established.

- 55. A method according to claim 54 wherein the intracellular signalling pathway comprises a cellular response that modulates the generation of free transcription factors of the NF-kappaB family which are able to redistribute to the nucleus.
 - 56. A method according to any of claims 54 or 55, wherein the fluorescent probe is modified in a systematic way, still keeping the GFP coding sequence intact, so that the new fluorescent probes are fusion polypeptides where parts of the suspected targeting sequences of the I-kappaB kinase are altered.
 - 57. A method according to claim 56, wherein the modification of the suspected targeting sequence of the I-kappaB kinase is a deletion.
 - 58. A method according to any of claims 56 or 57, wherein the spatial distribution of the fluorescent probe is compared to the spatial distribution of the unmodified fluorescent probe deducing the targeting sequence.
- 59. A method according to any of claims 16-58, wherein the substance interacts with the targeting sequence or part thereof in a manner that dislocates, disrupts targeting, or interferes with redistribution of the fluorescent probe as measured in quantitative fluorescence redistribution assay.

ABSTRACT

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This application describes a method by which to identify novel chemical entities that may modulate the specific effectiveness of the I-kappaB kinases (IKKs). The preferred mode of action is dislocation, disruption of targeting or interference with redistribution of specific isoforms of IKKs from their anchoring sites within cells, thereby modulating their specific effectiveness, not their enzymatic capacity. The chemical entities may be useful in preventing or treating, in an animal, preferably a human, in need thereof, an adverse condition which may be reduced or abolished by modulating the specific effectiveness of one or more IKKs. Examples of such adverse conditions are inflammatory and autoimmune diseases.

SEQUENCE LISTING

- (1) GENERAL INFORMATION
- (i) APPLICANT: NovoNordisk, BioImage
- (ii) TITLE OF THE INVENTION: A method for preventing or treating adverse conditions which may be reduced or abolished by modulating the effectiveness of one or more IkappaB kinases.
- (iii) NUMBER OF SEQUENCES: 4
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: NovoNordisk, BioImage
 - (B) STREET: Morkhøjbygade 28
 - (C) CITY: Søborg
 - (D) STATE: DK
 - (E) COUNTRY: DENMARK
 - (F) ZIP: 2860
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Diskette
 - (B) COMPUTER: IBM Compatible
 - (C) OPERATING SYSTEM: DOS
 - (D) SOFTWARE: FastSEQ for Windows Version 2.0
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: , PV&P R
 - (B) REGISTRATION NUMBER:
 - (C) REFERENCE/DOCKET NUMBER:
 - (2) INFORMATION FOR SEQ ID NO:1:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3024 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...3021
 - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

ATG AGC TGG TCA CCT TCC CTG ACA ACG CAG ACA TGT GGG GCC TGG GAA

Met Ser Trp Ser Pro Ser Leu Thr Thr Gln Thr Cys Gly Ala Trp Glu

1 5 10 15

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					GGG . Gly							Val				96
					GGT Gly											144
					AAC Asn											192
ATG Met 65	AGA Arg	AGG Arg	CTG Leu	ACC Thr	CAC His 70	CCC Pro	AAT Asn	GTG Val	GTG Val	GCT Ala 75	GCC Ala	CGA Arg	GAT Asp	GTC Val	CCT Pro 80	240
GAG Glu	GGG Gly	ATG Met	CAG Gln	AAC Asn 85	TTG Leu	GCG Ala	CCC	AAT Asn	GAC Asp 90	CTG Leu	CCC Pro	CTG Leu	CTG Leu	GCC Ala 95	ATG Met	288
GAG Glu	TAC Tyr	TGC Cys	CAA Gln 100	GGA Gly	GGA Gly	GAT Asp	CTC Leu	CGG Arg 105	AAG Lys	TAC Tyr	CTG Leu	AAC Asn	CAG Gln 110	TTT Phe	GAG Glu	336
AAC Asn	TGC Cys	TGT Cys 115	GGT Gly	CTG Leu	CGG Arg	GAA Glu	GGT Gly 120	Ala	ATC	CTC Leu	ACC Thr	TTG Leu 125	CTG Leu	AGT Ser	GAC Asp	384
ATT Ile	GCC Ala 130	Ser	GCG Ala	CTT	AGA Arg	TAC Tyr 135	Leu	CAT His	GAA Glu	AAC Asn	AGA Arg 140	Ile	ATC	CAT	CGG Arg	432
GAT Asp 145	Leu	AAG Lys	CCA Pro	GAA Glu	AAC Asn 150	ATC	GTC	CTG Leu	CAC Glr	G CAP Glr 155	Gly	GAA Glu	CAG Gln	AGG Arg	Leu 160	480
ATA Ile	CAC His	AAA Lys	Ile	ATT	asp	CTA Leu	GG/	ТАТ УТУГ	Ala	AAC	GAC Glu	CTC	GAT Asp	CAC Glr 175	GGC Gly	528
AGT Ser	CTI Lev	TGC Cys	ACA Thr	Ser	TTC	GTC Val	GG(G ACC y Thi 185	Le	G CAC	э тас э Туз	CTC	G GCC 1 Ala 190	a Pro	A GAG o Glu	576
CT <i>A</i> Let	A CTO	GAC Glu 195	ı Glr	G CAC	AAG n Lys	TAC	AC Th	r Val	G ACC	C GTG	C GAG l Ası	TAC TYT 205	r Tr	G AGO	TTC r Phe	624
GG(Gly	C ACC y Thi 210	Le	G GCC	TT a Phe	r GAC e Glu	TG0 1 Cys 21!	s Il	C ACC e Th:	G GG r Gl	C TT y Ph	C CG e Ar 22	g Pr	C TTY	c CT	C CCC u Pro	672
AA	TG(G CAS	G CC	GTY	G CAC	G TG	G CA	T TC.	A AA	A GT	G CG	G CA	G AA	g ag	T GAG	720

Asn 225	Trp	Gln	Pro		Gln 230	Trp	His	Ser		Val 235	Arg	Gln	Lys	Ser	Glu 240	
GTG Val	GAC Asp	ATT Ile	GTT Val	GTT Val 245	AGC Ser	GAA Glu	GAC Asp	TTG Leu	AAT Asn 250	GGA Gly	ACG Thr	GTG Val	AAG Lys	TTT Phe 255	TCA Ser	768
								CTT Leu 265								816
								CTG Leu								864
GGC	ACG Thr 290	GAT Asp	CCC Pro	ACG Thr	TAT Tyr	GGG Gly 295	CCC Pro	AAT Asn	GGC Gly	TGC Cys	TTC Phe 300	AAG Lys	GCC Ala	CTG Leu	GAT Asp	912
GAC Asp 305	Ile	TTA Leu	AAC Asn	TTA Leu	AAG Lys 310	CTG Leu	GTT Val	CAT His	ATC Ile	TTG Leu 315	AAC Asn	ATG Met	GTC Val	ACG Thr	GGC Gly 320	960
ACC Thr	: ATC	CAC	ACC Thr	TAC Tyr 325	Pro	GTG Val	ACA Thr	GAG Glu	GAT Asp 330	GAG Glu	AGT Ser	CTG	CAG Gln	AGC Ser	TTG Leu	1008
				Gln					Ile					Glr	G GAG n Glu	1056
CTC	G CTC	CAC Glr 355	Glu	GCG Ala	GGC Gly	CTC Lev	GC0 Ala 360	a Leu	ATC	CCC Pro	GAT ASI	7 AAC 5 Lys 365	Pro	GCC Ala	C ACT	1104
CA(TG1 n Cys	i Ile	TCA e Ser	A GAC	: GGC	AAC Lys	Le	AAA Asn	GAG	GG(Gly	CAC His	s Thi	A TTO	G GA	C ATG p Met	1152
GA' As ₁ 38	p Lei	r GT ı Val	T TTT	r CTO	TTT Phe	e Ası	AAC Asi	C AGT	r AAA	ATC 110 399	e Th	C TA' r Ty:	T GAG	G AC	T CAG r Gln 400	1200
AT Il	C TC(e Se:	C CC.	A CG(c Arg	G CCC g Pro 40	Glr	CC'	r GA	A AG? u Sei	r GTC r Val	l Se	C TG r Cy	T AT	C CT	T CA u Gl 41	A GAG n Glu 5	1248
CC Pr	C AA	G AG s Ar	G AA' g Asi 42	n Le	GCC Ala	TTV	C TT e Ph	C CAG e Gli 42!	n Lei	G AG u Ar	G AA g Ly	G GT	G TG 1 Tr 43	p Gl	C CAG	1296
GT Va	C TG 1 Tr	G CA p Hi 43	s Se	C AT	C CAG	G AC	C CT r Le 44	u Ly	G GA s Gl	A GA u As	T TG p Cy	C AA 's As 44	n Ar	G CI	CAG eu Gln	1344

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														AGC Ser		1392
	450		J			455					460					
														CTC		1440
Leu	Ser	Lys	Met	Lys		Ser	Met	Ala	Ser		Ser	Gln	Gln	Leu		
465					470					475					480	
														GAG		1488
Ala	Lys	Leu	ASP	485	Pne	Lys	THE	ser	490	GIII	116	ASP	Dea	Glu 495	Dys	
														CTG		1536
Tyr	Ser	Glu	Gln 500	Thr	Glu	Phe	Gly	Ile 505	Thr	Ser	Asp	Lys	Leu 510	Leu	Leu	
														GAG		1584
Ala	Trp	Arg 515	Glu	Met	Glu	Gln	Ala 520	Val	Glu	Leu	Cys	Gly 525	Arg	Glu	Asn	
GAA	GTG	AAA	CTC	CTG	GTA	GAA	CGG	ATG	ATG	GCT	CTG	CAG	ACC	GAC	TTA	1632
Glu	Val	Lys	Leu	Leu	Val		Arg	Met	Met	Ala		Gln	Thr	Asp	Ile	
	530					535					540					
															CTG	1680
		Leu	Gln	Arg	Ser 550	Pro	Met	Gly	Arg	Lys 555	Gln	Gly	GIY	Thr	Leu 560	
545																
															GAA	1728
Asp	Asp	Leu	Glu	565		Ala	Arg	GIU	570		ALG	Arg	Dec	575	Glu	
															CGG	1776
Lys	Pro	Arg	Asp 580		Arg	Thr	Glu	Gly 585		Ser	Gln	Glu	590		. Arg	
CTG	CTC	CTI	, CYC	GCA	TTA	CAG	AGC	TTC	GAG	AAG	AAA	GTG	G CG	A GTC	ATC	1824
Leu	Lev	Lev 595		n Ala		Gln						Val 605		y Val	lle	
TAT	. ACC	G CAC	CTC	AGT	: AAA	ACI	GTC	GT	TGC	: AAC	CAC	AAC	GC(G CTC	G GAA	1872
Туг	Thi 610		ı Lei	ı Ser	Lys	Thr 615		. Val	Cys	Lys	620		s Ala	a Lei	ı Glu	
CTY	יייי ב		- 110	GTY	GAA	GAC	GTC	GT	ago	TTA	YTA A	raa e	r gag	G GA'	r gag	1920
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625	5				630)				635	ò				640	
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Lys	s Th	r Vai	l Vai	1 Arg 645		ı Glı	ı Glu	ı Ly:	650		ı Ly:	s Glı	ı Le	u Trj 65	p Asn 5	
CT	CTY	G AA	g at	T GC	r TGI	DA 7	C AAC	G GTV	c cgʻ	r GG'	r cc	T GT	C AG	T GG	A AGC	2016

Leu	Leu	Lys	Ile 660	Ala	Cys	Ser	Lys	Val 665	Arg	Gly	Pro	o Val	67(G1	у S	er	
CCG Pro	GAT Asp	AGC Ser 675	ATG Met	AAT Asn	GCC Ala	TCT Ser	CGA Arg 680	CTT Leu	AGC Ser	CAC Glr	G CC'	r GG(5 Gl) 689	y Gli	n Le	rg A eu M	.TG let	2064
TCT Ser	CAG Gln 690	Pro	TCC Ser	ACG Thr	GCC Ala	TCC Ser 695	AAC Asn	AGC Ser	TTA Leu	CC'	r GA o Gl 70	u Pr	A GCO	C A	AG <i>P</i> ys I	AG Lys	2112
AGT Ser 705	Glu	GAA	. CTG Leu	GTG Val	GCT Ala 710	GAA Glu	GCA Ala	CAT	AAC	CT Le 71	u Cy	C AC	C CT	G C u L	eu (GAA Glu 720	2160
AAT Asn	GCC	TA C	A CAC	GAC Asp 725	ACT Thr	GTG Val	AGG Arg	GAA Glu	CA/ Glr 730	ı As	.C C.A p Gl	AG AG in S∈	or TT er Ph	ne T	CG (hr .	GCC Ala	2208
CTA Lev	A GAG	TG(S AG0 Sei 740	Tr	TTA	CAG Gln	ACC Thr	GAA Glu 749	ı Gl	A GA u Gl	A GA Lu G	AG CA	is Se	GC T er C	rgc Ys	CTG Leu	2256
GAC Glu	G CA 1 Gl	G GC n Al 75	a Se	A TGC r Tri	GTA Val	CCC Pro	CGC Arg	g Ala	C CG a Ar	G GA	AT CO	ro P	CG G' ro V	TC (GCC Ala	ACC Thr	2304
ATY Me	G GT t Va 77	l Se	C AA r Ly	G GGG S Gl	C GAC y Glu	G GAG L Glu 77!	ı Le	G TT u Ph	C AC	C G(ir G)	ly V	TG G al V 80	TG C	cc . ro	ATC Ile	CTG Leu	2352
GT Va 78	1 G]	G CT u Le	'G GA eu As	c GG p Gl	C GAG y Ası 79	p Va	A AA l As	C GG n Gl	C CA y Hi	s L	AG T ys F 95	TC A he S	GC G	TG al	TCC Ser	GGC Gly 800	2400
GA G1	.G G(u G)	GC GA	.u G1	GA Y As 80		C AC a Th	C TA r Ty	c GG r Gl	уL	AG C Ys L 10	TG A	ACC C	CTG A Leu I	AAG Lys	TTC Phe 815	ITE	2448
C.Z.	C A	CC AC	nr G	€C AA Ly Ly 00	G CT 's Le	G CC	C G1 O Vē	al Pi	CC TY CO T:	3G C rp F	ecc i	ACC (Leu '	GTG Val 830	ACC Thr	ACC Thr	2496
C7 Le	rG A eu T	hr T	AC G yr G 35	GC G1 ly Va	rg CA al Gl	G TC	s Pl	rc AG ne Se 40	GC C er A	GC :	rac (Tyr	Pro .	GAC Asp 845	CAC His	ATC	AAG Lys	2544
C: G	ln H	AC G lis A	AC T sp P	TC T' he Pl	IC AA	ys Se	cc G er A 55	CC A	TG C et F	cc (GAA Glu	GGC Gly 860	TAC Tyr	GTC Val	CA(G GAG n Glu	2592
A	GC A rg 1 65	CC A	TC T	TC T	he L	AG G ys A 70	AC G sp A	AC G sp G	GC A	Asn	TAC Tyr 875	AAG Lys	ACC Thr	CGC	GC Al	C GAG a Glu 880	2640

GTG Val	AAG Lys	TTC Phe	GAG Glu	GGC Gly 885	GAC Asp	ACC Thr	CTG Leu	GTG Val	AAC Asn 890	CGC Arg	ATC Ile	GAG Glu	CTG Leu	AAG Lys 895	GGC Gly	2688
ATC Ile	GAC Asp	TTC Phe	AAG Lys 900	GAG Glu	GAC Asp	GGC Gly	AAC Asn	ATC Ile 905	CTG Leu	GGG Gly	CAC His	AAG Lys	CTG Leu 910	GAG Glu	TAC Tyr	2736
AAC Asn	TAC Tyr	AAC Asn 915	AGC Ser	CAC His	AAC Asn	GTC Val	ТАТ Туг 920	ATC Ile	ATG Met	GCC Ala	GAC Asp	AAG Lys 925	CAG Gln	AAG Lys	AAC Asn	2784
GGC Gly	ATC Ile 930	AAG Lys	GTG Val	AAC Asn	TTC Phe	AAG Lys 935	ATC Ile	CGC Arg	CAC His	AAC Asn	ATC Ile 940	Glu	GAC Asp	GGC Gly	AGC Ser	2832
GTG Val 945	Gln	CTC	GCC Ala	GAC Asp	CAC His 950	TAC Tyr	CAG Gln	CAG Gln	AAC Asn	ACC Thr 955	Pro	ATC	GGC Gly	GAC Asp	GGC Gly 960	2880
CCC Pro	GTG Val	CTC Leu	CTG	CCC Pro 965	Asp	AAC Asn	CAC His	TAC Tyr	CTG Lev 970	. Ser	Thr	CAC Glr	TCC Ser	GCC Ala 975	CTG Leu	2928
AGC Sei	AAA Lys	GAC	980	Asr	GAG	AAC Lys	G CGC	GAT Asp 989	o His	ATC Met	GTY Val	CTC	CTO Let 990	ı Glu	TTC Phe	2976
GT(Va	G ACC	GCC Ala 991	a Ala	GGC Gly	ATC	C ACT	r CTO	u Gl	C ATO	G GAC	GAG	G CTO u Leo 100	u Ty	C AAG	G TAA	3024

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1007 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

 Met
 Ser
 Trp
 Ser
 Leu
 Thr
 Thr
 Gln
 Thr
 Cys
 Gly
 Ala
 Trp
 Glu

 Met
 Lys
 Glu
 Arg
 Leu
 Gly
 Thr
 Gly
 Phe
 Gly
 Asn
 Val
 Ile
 Arg
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 Asn
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65	Arg				His 70	Pro				75					80
Glu	Gly	Met	Gln	Asn 85	Leu .	Ala	Pro	Asn	Asp 90	Leu	Pro	Leu	Leu	Ala 95	Met
Glu	Tyr	Cys	Gln 100	Gly	Gly	Asp	Leu	Arg 105	Lys	Tyr	Leu	Asn	Gln 110	Phe	Glu
Asn	Cys	Cys 115		Leu	Arg	Glu	Gly 120	Ala	Ile	Leu	Thr	Leu 125	Leu	Ser	Asp
	130			Leu		135					140				
145				Glu	150					155					160
				Ile 165					170					175	
			180	Ser				185					190		
		195					200					205			Phe
	210					215					220)			Pro
225					230					235	5				240
				245					250)				255	
			260)				265)				270)	Arg
		275	5				280					285	5		n Arg
	290)				295	•				300	0			r Gly
305	5				310	1				31	5				320 r Leu
				325	5				3.3	0				33	5 n Glu
			340	C				34	5				35	O	a Thr
		35	5				360)				36	5		p Met
	37	0				379	5				38	0			ır Gln
38	5				390)				39	5				400 n Glu
				40	5				41	.0				41	.5 .y Gln
			42	0				42	5				43	30	eu Gln
		43	5				44	0				4.4	15		er Cys
	45	0				45	5				46	50			eu Lys
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490 485 Tyr Ser Glu Gln Thr Glu Phe Gly Ile Thr Ser Asp Lys Leu Leu 505 Ala Trp Arg Glu Met Glu Gln Ala Val Glu Leu Cys Gly Arg Glu Asn 520 Glu Val Lys Leu Leu Val Glu Arg Met Met Ala Leu Gln Thr Asp Ile 535 540 Val Asp Leu Gln Arg Ser Pro Met Gly Arg Lys Gln Gly Gly Thr Leu 555 550 Asp Asp Leu Glu Glu Gln Ala Arg Glu Leu Tyr Arg Arg Leu Arg Glu 570 Lys Pro Arg Asp Gln Arg Thr Glu Gly Asp Ser Gln Glu Met Val Arg 585 580 Leu Leu Cln Ala Ile Gln Ser Phe Glu Lys Lys Val Arg Val Ile 600 Tyr Thr Gln Leu Ser Lys Thr Val Val Cys Lys Gln Lys Ala Leu Glu 620 615 Leu Leu Pro Lys Val Glu Glu Val Val Ser Leu Met Asn Glu Asp Glu 635 630 Lys Thr Val Val Arg Leu Gln Glu Lys Arg Gln Lys Glu Leu Trp Asn 645 650 Leu Leu Lys Ile Ala Cys Ser Lys Val Arg Gly Pro Val Ser Gly Ser 665 Pro Asp Ser Met Asn Ala Ser Arg Leu Ser Gln Pro Gly Gln Leu Met 685 680 Ser Gln Pro Ser Thr Ala Ser Asn Ser Leu Pro Glu Pro Ala Lys Lys 695 Ser Glu Glu Leu Val Ala Glu Ala His Asn Leu Cys Thr Leu Leu Glu 715 710 Asn Ala Ile Gln Asp Thr Val Arg Glu Gln Asp Gln Ser Phe Thr Ala 725 730 Leu Asp Trp Ser Trp Leu Gln Thr Glu Glu Glu Glu His Ser Cys Leu 745 Glu Gln Ala Ser Trp Val Pro Arg Ala Arg Asp Pro Pro Val Ala Thr 760 Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 775 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 795 790 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 810 805 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 825 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 845 840 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 855 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 875 870 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 890 885 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 900 905 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn

 915
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 Gly
 11e
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 Val
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 Arg
 His
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 Asn
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 Thr
 Pro
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(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 31 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GTAAGCTTAC ATGAGCTGGT CACCTTCCCT G

- (2) INFORMATION FOR SEQ ID NO:4:
- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

GTGGTACCCA TGAGGCCTGC TCCAG

AN IMPROVED METHOD for extracting quantitative information relating to an influence on a cellular response.

SUMMARY OF THE INVENTION

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The present invention relates to an improved method and tools for extracting quantitative information relating to an influence on a cellular response, in particular an influence caused by contacting or incubating the cell with a substance influencing a cellular response, wherein the cellular response is manifested in redistribution of at least one component in the cell. In particular, the invention relates to an improved method for extracting the quantitative information relating to an influence on an intracellular pathway involving redistribution of at least one component associated with the pathway. The method of the invention may be used as a very efficient procedure for testing or discovering the influence of a substance on a physiological process, for example in connection with screening for new drugs, testing of substances for toxicity, identifying drug targets for known or novel drugs. In particular, the present invention relates to an 15 improved method for parallelisation of the testing procedure so that a large number of substances can be tested simultaneously using commercially available instrumentation. The invention also describes several ways of contacting the cells with a substance influencing a cellular response and modifications made to the actual cells before, during or after contacting the cells with these substances as to improve the applicability and use of 20 the method for extracting quantitative information relating to influence on an intracellular pathway in a highly parallel fashion. Other valuable uses of the method and technology of the invention will be apparent to the skilled person on the basis of the following disclosure. In a particular embodiment of the invention, the present invention relates to a method of detecting intracellular translocation or redistribution of biologically active polypeptides, 25 preferably an enzyme, affecting intracellular processes, and a DNA construct and a cell for use in the method.

Two appendices are included herein, and are considered part of the application. Appendix I, "METHOD AND APPARATUS FOR HIGH DENSITY FORMAT SCREENING FOR BIOACTIVE MOLECULES", is a pending patent application. Appendix II, "CHANGES

IN INTRACELLULAR cAMP VISUALIZED USING A cAMP-DEPENDENT PROTEIN KINASE-GREEN FLUORESCENT PROTEIN HYBRID", is a manuscript intended for publication.

5 BACKGROUND OF THE INVENTION

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Intracellular pathways are tightly regulated by a cascade of components that undergo modulation in a temporally and spatially characteristic manner. Several disease states can be attributed to altered activity of individual signalling components (i.e. protein kinases, protein phosphatases, transcription factors). These components therefore render themselves as attractive targets for therapeutic intervention.

Protein kinases and phosphatases are well described components of several intracellular signalling pathways. The catalytic activity of protein kinases and phosphatases are assumed to play a role in virtually all regulatable cellular processes. Although the involvement of protein kinases in cellular signalling and regulation have been subjected to extensive studies, detailed knowledge on e.g. the exact timing and spatial characteristics of signalling events is often difficult to obtain due to lack of a convenient technology.

Novel ways of monitoring specific modulation of intracellular pathways in intact, living cells is assumed to provide new opportunities in drug discovery, functional genomics, toxicology, patient monitoring etc.

The spatial orchestration of protein kinase activity is likely to be essential for the high degree of specificity of individual protein kinases. The phosphorylation mediated by protein kinases is balanced by phosphatase activity. Also within the family of phosphatases translocation has been observed, e.g. translocation of PTP2C to membrane ruffles [(Cossette et al. 1996)], and likewise is likely to be indicative of phosphatase activity.

25 Protein kinases often show a specific intracellular distribution before, during and after activation. Monitoring the translocation processes and/or redistribution of individual protein kinases or subunits thereof is thus likely to be indicative of their functional activity. A connection between translocation and catalytic activation has been shown for

protein kinases like the diacyl glycerol (DAG)-dependent protein kinase C (PKC), the cAMP-dependent protein kinase (PKA) [(DeBernardi *et al.* 1996)] and the mitogenactivated-protein kinase Erk-1 [(Sano *et al.* 1995)].

Commonly used methods of detection of intracellular localisation/activity of protein kinases and phosphatases are immunoprecipitation, Western blotting and immunocytochemical detection.

Taking the family of diacyl glycerol (DAG)-dependent protein kinase Cs (PKCs) as an example, it has been shown that individual PKC isoforms that are distributed among different tissues and cells have different activator requirements and undergo differential translocation in response to activation. Catalytically inactive DAG-dependent PKCs are generally distributed throughout the cytoplasm, whereas they upon activation translocate to become associated with different cellular components, e.g. plasma membrane [(Farese, 1992),(Fulop Jr. et al.1995)] nucleus [(Khalil et al.1992)], cytoskeleton [(Blobe et al. 1996)]. The translocation phenomenon being indicative of PKC activation has been monitored using different approaches: a) immunocytochemistry where the localisation of individual isoforms can be detected after permeabilisation and fixation of the cells [(Khalil et al. 1992)]; and b) tagging all DAG-dependent PKC isoforms with a fluorescently labelled phorbol myristate acetate (PMA) [(Godson et al. 1996)]; and c) chemical tagging of PKC \$1 with the fluorophore Cy3 [(Bastiaens & Jovin 1996)] and d) genetic tagging of PKC α ([Schmidt et al. 1997]) and of PKC γ and PKC δ [(Sakai et al. 1996)]. The first method does not provide dynamic information whereas the latter methods will. Tagging PKC with fluorescently labelled phorbol myristate acetate cannot distinguish between different DAG-dependent isoforms of PKC but will label and show movement of all isoforms. Chemical and genetic labelling of specific DAG-dependent PKCs confirmed that they in an isoform specific manner upon activation move to cell periphery or nucleus.

In an alternative method, protein kinase A activity has been measured in living cells by chemical labelling one of the kinase's subunit [(Adams *et al.*1991)]. The basis of the methodology is that the regulatory and catalytic subunit of purified protein kinase A is labelled with fluorescein and rhodamine, respectively. At low cAMP levels protein kinase A is assembled in a heterotetrameric form which enables fluorescence resonance energy

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transfer between the two fluorescent dyes. Activation of protein kinase A leads to dissociation of the complex, thereby eliminating the energy transfer. A disadvantage of this technology is that the labelled protein kinase A has to be microinjected into the cells of interest. This highly invasive technique is cumbersome and not applicable to large scale screening of biologically active substances. A further disadvantage of this technique as compared to the presented invention is that the labelled protein kinase A cannot be inserted into organisms/animals as a transgene.

Recently it was discovered that Green Fluorescent Protein (GFP) expressed in many different cell types, including mammalian cells, became highly fluorescent [(Chalfie et al.1994)]. WO95/07463 describes a cell capable of expressing GFP and a method for detecting a protein of interest in a cell based on introducing into a cell a DNA molecule having DNA sequence encoding the protein of interest linked to DNA sequence encoding a GFP such that the protein produced by the DNA molecule will have the protein of interest fused to the GFP, then culturing the cells in conditions permitting expression of the fused protein and detecting the location of the fluorescence in the cell, thereby localizing the protein of interest in the cell. However, examples of such fused proteins are not provided, and the use of fusion proteins with GFP for detection or quantitation of translocation or redistribution of biologically active polypeptides affecting intracellular processes upon activation, such as proteins involved in signalling pathways, e.g. protein kinases or phosphatases, has not been suggested. WO 95/07463 further describes cells useful for the detection of molecules, such as hormones or heavy metals, in a biological sample, by operatively linking a regulatory element of the gene which is affected by the molecule of interest to a GFP, the presence of the molecules will affect the regulatory element which in turn will affect the expression of the GFP. In this way the gene encoding GFP is used as a reporter gene in a cell which is constructed for monitoring the presence of a specific molecular identity.

Green Fluorescent Protein has been used in an assay for the detection of translocation of the glucocorticoid receptor (GR) [(Carey, KL et al.1996)]. A GR-S65TGFP fusion has been used to study the mechanisms involved in translocation of the glucocorticoid receptor (GR) in response to the agonist dexamethasone from the cytosol, where it is present in the absence of a ligand, through the nuclear pore to the nucleus where it remains after ligand

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binding. The use of a GR-GFP fusion enables real-time imaging and quantitation of nuclear/cytoplasmic ratios of the fluorescence signal. A similar genetic construct has been used to follow and quantify dexamethasone induced translocation of GR to the nucleus in HeLa cells [(Guiliano, K.A et al. 1997)] in a system called Array ScanTM (WO 97/45730) designed for automated drug screening. Recently, several other investigators have demonstrated that tagging a specific protein (or part of a protein) involved in an intracellular signalling pathway with GFP provides a new means to measure and quantify the influence of substances on this pathway. The concept has been shown to work both for cytoplasmic to nuclear translocation of the androgen receptor [(Georget V et al. 1997)] and transcription factors such as NF-ATc [(Beals CR et al. 1997)] in analogy with what has already been described for GR above. Another relevant example is a β -arrestin – GFP construct that was shown to report on activation of G-protein coupled receptors by translocating from the cytosol to the plasma membrane [(Barak LS et al. 1997)]. Finally, it has also been demonstrated that attaching GFP to a smaller part of a protein like the pleckstrin homology domain of phospholipase C & 1 [(Stauffer TP et al. 1998)] and a cysteine-rich domain of PKC y [(Oancea E et al. 1998)] can be used to report on an influence from a substance by quantifying their redistribution within the cells during activation of the specific signalling pathway to which they belong.

Many currently used screening programmes designed to find compounds that affect protein kinase activity are based on measurements of kinase phosphorylation of artificial or natural substrates, receptor binding and/or reporter gene expression. The interest in fluorescence measurements as the basis for future high-throughput drug screening has however increased dramatically over the last few years [(Silverman L *et al.* 1998)]. Of particular interest to the present invention is a scanning laser imager for rapid screening of fluorescence changes in living cells [(Schroeder K & Neagle B 1996)] currently offered commercially by Molecular Devices, Inc. as the FLIPRTM.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides an important new dimension in the investigation of cellular systems involving redistribution in that the invention provides quantification of the

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redistribution responses or events caused by an influence, typically contact with a chemical substance or mixture of chemical substances, but also changes in the physical environment. The quantification makes it possible to set up meaningful relationships, expressed numerically, or as curves or graphs, between the influences (or the degree of influences) on cellular systems and the redistribution response. This is highly advantageous because, as has been found, the quantification can be achieved in both a fast and reproducible manner, and - what is perhaps even more important - the systems which become quantifiable utilising the method of the invention are systems from which enormous amounts of new information and insight can be derived.

The present screening assays have the distinct advantage over other screening assays, e.g., receptor binding assays, enzymatic assays, and reporter gene assays, in providing a system in which biologically active substances with completely novel modes of action, e.g. inhibition or promotion of redistribution/translocation of a biologically active polypeptide as a way of regulating its action rather than inhibition/activation of enzymatic activity, can be identified in a way that insures very high selectivity to the particular isoform of the biologically active polypeptide and further development of compound selectivity versus other isoforms of the same biologically active polypeptide or other components of the same signalling pathway.

In its broadest aspect, the invention relates to an improved method, with higher throughput compared to previous methods, for extracting quantitative information relating to an influence on a cellular response, the method comprising recording variation, caused by the influence on mechanically intact living cells, in spatially distributed light emitted from a luminophore, the luminophore being present in the cells and being capable of being redistributed in a manner which is related with the degree of the influence, and/or of being modulated by a component which is capable of being redistributed in a manner which is related to the degree of the influence, the association resulting in a modulation of the luminescence characteristics of the luminophore, detecting and recording the spatially distributed light from the luminophore, and processing the recorded variation in the spatially distributed light to provide quantitative information correlating the spatial distribution or change in the spatial distribution to the degree of the influence. In one aspect of the present invention the mechanically intact living cell is permeabilised at some

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time after the influence has begun but during or before the actual experimental recording. In another aspect, the present invention relates to an improved method for extracting quantitative information relating to an influence on a cellular response, the method comprising recording variation, caused by the influence on permeabilised living cells, in spatially distributed light emitted from a luminophore, the luminophore being present in the cells and being capable of being redistributed in a manner which is related with the degree of the influence, and/or of being modulated by a component which is capable of being redistributed in a manner which is related to the degree of the influence, the association resulting in a modulation of the luminescence characteristics of the luminophore, detecting and recording the spatially distributed light from the luminophore, and processing the recorded variation in the spatially distributed light to provide quantitative information correlating the spatial distribution or change in the spatial distribution to the degree of the influence. In a preferred embodiment of the invention the luminophore, which is present in the cells, is capable of being redistributed by modulation of an intracellular pathway, in a manner which is related to the redistribution of at least one component of the intracellular pathway. In another preferred embodiment of the invention, the luminophore is a fluorophore.

In the invention the cell and/or cells are mechanically intact and alive throughout the experiment. In another embodiment of the invention, the cells are fixed at a point in time after the application of the influence at which the response has been predetermined to be significant, and the recording is made at an arbitrary later time. In another embodiment the cell and/or cells are mechanically intact and alive throughout the experiment but are mechanically or chemically disrupted or permeabilised as the initial step of experimental analysis. In another aspect of the invention the cells have their plasma membrane permanently and stably permeabilised before the initiation of the experiment in such a way that the plasma membrane stays permeable during the experiment. This allows the components of intracellular pathways to be contacted by substances that are not normally permeating the cell plasma membrane such as peptides, proteins and hydrophilic organic compounds.

The mechanically intact or permeabilised living cells could be selected from the group consisting of fungal cells, such as yeast cells; invertebrate cells including insect cells; and

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vertebrate cells, such as mammalian cells. These cells are incubated at a temperature of 30°C or above, preferably at a temperature of from 32°C to 39°C, more preferably at a temperature of from 35°C to 38°C, and most preferably at a temperature of about 37°C during the time period over which the influence is observed. In one aspect of the invention the mechanically intact or permeabilised living cell is part of a matrix of identical or non-identical cells. In one embodiment of the invention the cells comprise a group or groups of cells contained within a spatial limitation or spatial limitations. In one embodiment, the cells comprise multiple groups of cells that are qualitatively the same but subjected to different influences. In another embodiment, the cells comprise multiple groups of cells that are qualitatively different but subjected to the same influence.

In one embodiment of the invention the spatial limitations are domains defined on a substrate on which the cells are present. The spatial limitations may be arranged in one or more arrays on a common carrier. The spatial limitations may be wells in a plate of microtiter type, such that 96, 384, 864 and 1536 wells are situated on the common carrier. In another embodiment the spatial limitations are wells in a plate of a format different from the microtiter type. In one embodiment of the invention the domains are established by the presence of the cells on the substrate in a pattern that defines the domains. In another aspect of the invention, the domains are instead established by the spatial pattern or array of the influence or influences as it/they are applied to or contacted by the cells. This aspect is thoroughly described in Appendix I. Briefly, in this aspect of the invention the mechanically intact or permeabilised living cells are part of a continuous or discontinuous sheet of cells cultured on an optically clear flat surface optimised or not for cell culture. The optically clear and flat surface may be a porous membrane that may allow cellular processes to grow through the membrane pores and may allow directed capillary flow of fluid through the pores.

A cell used in the present invention should contain a nucleic acid construct encoding a fusion polypeptide as defined herein and be capable of expressing the sequence encoded by the construct. The cell is a eukaryotic cell selected from the group consisting of fungal cells, such as yeast cells; invertebrate cells including insect cells; vertebrate cells such as mammalian cells. The preferred cells are mammalian cells.

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In another aspect of the invention the cells could be from an organism carrying in at least one of its component cells a nucleic acid sequence encoding a fusion polypeptide as defined herein and be capable of expressing said nucleic acid sequence. The organism is selected from the group consisting of unicellular and multicellular organisms, such as a mammal.

The luminophore is the component that allows the redistribution to be visualised and/or recorded by emitting light in a spatial distribution related to the degree of influence. The term redistribution is intended to cover all aspects of a change in spatial location, such as a translocation of the luminophore or other components. In one embodiment of the invention, the luminophore is capable of being redistributed in a manner that is physiologically relevant to the degree of the influence. It should be understood that redistribution. In another embodiment, the luminophore is capable of associating with a component that is capable of being redistributed in a manner that is physiologically relevant to the degree of the influence. In another embodiment, a correlation between the redistribution of the luminophore and the degree of the influence could be determined experimentally. In a preferred aspect of the invention, the luminophore is capable of being redistributed in substantially the same manner as the at least one component of an intracellular pathway. In another embodiment of the invention, the luminophore is capable of being quenched upon spatial association with a component that is redistributed by modulation of the pathway, the quenching being measured as a change in the intensity of the luminescence. In another embodiment of the invention, the luminophore is stationary but may have a certain spatial distribution, and interacts with at least one component that is capable of being redistributed in a manner which is physiologically relevant to the degree of the influence, in such a way that one or more luminescence characteristics of the luminophore is/are modulated as the component moves closer to, or farther from, the luminophore.

The luminophore could be a fluorophore. In a preferred embodiment of the invention, the luminophore is a polypeptide encoded by and expressed from a nucleotide sequence harboured in the cells. The luminophore could be a hybrid polypeptide comprising a fusion of at least a portion of each of two polypeptides one of which comprises a luminescent polypeptide and the other one of which comprises a biologically active polypeptide, as

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defined herein.

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The luminescent polypeptide could be a GFP as defined herein or could be selected from the group consisting of green fluorescent proteins having the F64L mutation as defined herein such as F64L-GFP, F64L-Y66H-GFP, F64L-S65T-GFP, and EGFP. The GFP could be N- or C-terminally tagged, optionally via a peptide linker, to the biologically active polypeptide or a part or a subunit thereof. The fluorescent probe could be a component of an intracellular signalling pathway. The probe is coded for by a nucleic acid construct.

The pathway of investigation in the present invention could be an intracellular signalling pathway.

In a preferred embodiment of the invention, the influence could be contact between the group or groups of mechanically intact or permeabilised living cells and a chemical substance, and/or incubation of the group or groups of mechanically intact or permeabilised living cells with a chemical substance in solution. In one aspect of the invention that is thoroughly described in Appendix I, the chemical substances are attached to an underlying matrix. In this aspect, the chemical substances may also be produced and secreted from, or attached to the plasma membrane surfaces of, a sheet of genetically engineered cells. In this aspect of the invention the chemical substances may also have been separated two-dimensionally in a non-denaturing gel using electrophoresis and the gel is directly put in close proximity or direct contact with the mechanically intact or permeabilised living cells so that the chemical substances can contact the cells through diffusion or convection.

The influence will modulate the intracellular processes. In one aspect the modulation could be an activation of the intracellular processes. In another aspect the modulation could be a deactivation of the intracellular processes. In yet another aspect, the influence could inhibit or promote the redistribution without directly affecting the metabolic activity of the component of the intracellular processes.

In one embodiment the invention is used to establish a dose-response relationship for one or many chemical substances. In one embodiment the invention is used as a basis for a screening program, where the effect of unknown influences such as a compound library,

can be compared to influence of known reference compounds under standardised conditions.

In addition to the intensity, there are several parameters of fluorescence or luminescence that can be modulated by the effect of the influence on the underlying cellular phenomena, and can therefore be used in the invention. Some examples are resonance energy transfer, fluorescence lifetime, polarisation, and wavelength shift. Each of these methods requires a particular kind of filter in the emission light path to select the component of the light desired and reject other components. The recording of property of light could be in the form of an ordered array of values such as a CCD array or a vacuum tube device such as a vidicon. In addition, the translational mobility, or freedom of movement, of the luminophore attached to the protein of interest can be an important property affected by the influence on the underlying cellular phenomena, and can therefore be used in he invention.

In one embodiment of the invention, the spatially distributed light emitted by a luminophore is detected by a change in the resonance energy transfer between the luminophore and another luminescent entity capable of delivering energy to the luminophore, each of which has been selected or engineered to become part of, bound to or associated with particular components of the intracellular pathway. In this embodiment, either the luminophore or the luminescent entity capable of delivering energy to the luminophore undergoes redistribution in response to an influence. The resonance energy transfer would be measured as a change in the intensity of emission from the luminophore, preferably sensed by a single channel photodetector that responds only to the average intensity of the luminophore in a non-spatially resolved fashion.

In one embodiment of the invention, the spatially distributed light emitted by a luminophore includes the case of uniform spatial distribution of the light.

In one aspect of the invention, the luminophore is a fluorophore which redistributes through a non-homogenous excitation light field, resulting in a change in the intensity of the light emitted from the luminophore as a result of the change in the amount of excitation light intensity at different points in the field.

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In one embodiment of the invention, the recording of the spatially distributed light could be made at a single point in time after the application of the influence. In another embodiment, the recording could be made at two points in time, one point being before, and the other point being after the application of the influence. The result or variation is determined from the change in fluorescence compared to the fluorescence measured prior to the influence or modulation. In another embodiment of the invention, the recording could be performed at a series of points in time, in which the application of the influence occurs at some time after the first time point in the series of recordings, the recording being performed, e.g., with a predetermined time spacing of from 0.1 seconds to 1 hour, preferably from 1 to 60 seconds, more preferably from 1 to 30 seconds, in particular from 1 to 10 seconds, over a time span of from 1 second to 12 hours, such as from 10 seconds to 12 hours, e.g., from 10 seconds to one hour, such as from 60 seconds to 30 minutes or 20 minutes. The result or variation is determined from the change in fluorescence over time. The result or variation could also be determined as a change in the spatial distribution of the fluorescence over time.

In one embodiment the recording comprises a time series of total luminescence of the cells of one or several of the spatial limitations. In one embodiment the signal from all of the spatial limitations, one at a time, is measured by a recording being made in the individual spatial limitations by means of an apparatus to sequentially position each one of the limitations in the field of view of the detector and repeating the positioning and measurement process until all of the spatial limitations have been measured. The detector may be a photomultiplier tube. In a preferred embodiment of the present invention more than one spatial limitation is measured simultaneously. This may be done by means of a one- or two-dimensional array detector, whereby the multiple spatial limitations are imaged onto the array detector such that discrete subsets of the detecting units (pixels) in the array detector measure the signal from one and only one of the multiple spatial limitations, the signal from any one spatial limitation being the combined signal from those pixels that receive the image from one of the spatial limitations. This array detector may be a linear diode array, a video camera (according to any present or future standards and definitions of image acquisition and transmission) or a charge transfer device such as a charge-coupled device (CCD). In one embodiment the recording of signal requires

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illumination of the multiple spatial limitations to excite the luminophores so that they emit light. In one embodiment all of the spatial limitations are simultaneously illuminated during the measurement. In another embodiment the spatial limitations are singly illuminated only during the time in which they are being measured. In a preferred embodiment the illumination is provided by a laser that is scanned in a raster fashion over some or all of the spatial limitations being measured. The scanning may take place at a rate that is substantially faster than the measurement process such that the illumination appears to the measurement process to be continuous in time and spatially uniform over the region being measured.

The recording of spatially distributed luminescence emitted from the luminophore is performed by an apparatus for measuring the distribution of fluorescence in the cells, and thereby any change in the distribution of fluorescence in the cells, which includes at a minimum the following component parts: (a) a light source, (b) a method for selecting the wavelength(s) of light from the source which will excite the luminescence of the luminophore, (c) a device which can rapidly block or pass the excitation light into the rest of the system, (d) a series of optical elements for conveying the excitation light to the specimen, collecting the emitted fluorescence in a spatially resolved fashion, and forming an image from this fluorescence emission (or another type of intensity map relevant to the method of detection and measurement), (e) a bench or stand which holds the container of the cells being measured in a predetermined geometry with respect to the series of optical elements, (f) a detector to record the spatially resolved fluorescence in the form of an image, (g) a computer or electronic system and associated software to acquire and store the recorded images, and to compute the degree of redistribution from the recorded images.

In a preferred embodiment of the invention the apparatus system is automated. In one embodiment the components in d and e mentioned above comprise a fluorescence microscope. In one embodiment the component in f mentioned above is a CCD camera. In one embodiment the component in f mentioned above is an array of photomultiplier tubes/devices.

In one embodiment the image is formed and recorded by an optical scanning system.

In one embodiment the optical scanning system is used to illuminate the bottom of a plate

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of microtiter type so that a time-resolved recording of changes in luminescence or fluorescence can be made from all spatial limitations simultaneously.

In a preferred embodiment the actual luminescence or fluorescence measurements are made in a FLIPRTM instrument, commercially available from Molecular Devices, Inc.

In one embodiment of the invention the actual fluorescence measurements are made in a standard type of fluorometer for plates of microtiter type (fluorescence plate reader).

In one embodiment a liquid addition system is used to add a known or unknown compound to any or all of the cells in the cell holder at a time determined in advance. Preferably, the liquid addition system is under the control of the computer or electronic system. Such an automated system can be used for a screening program due to its ability to generate results from a larger number of test compounds than a human operator could generate using the apparatus in a manual fashion.

The methods whereby the detector layer of cells are physically contacted by the compounds can also be of another conceptual type where the compounds are delivered to the cells through a porous membrane by convection/diffusion or by directly contacting compounds attached to an inorganic or organic support (such as glass, plastic or the plasma membrane of intact living cells) with the cells. These methods are thoroughly described in Appendix I, but are also outlined in the following paragraphs.

In one aspect of the present invention where the detector layer of cells is a continuous or discontinuous sheet of cells without any separation into test units or wells. The compounds are printed onto a nonabsorbent sheet of porous material as a solution in solvent and allowed to dry. This printed sheet of compounds then defines the test pattern for the experiment as it is brought down in close proximity to or in direct contact with the underlying detector layer of cells. The compounds, now dissolved by the fluid layer on the cells, is brought in contact with the cells through the pores of the membrane by convection. The porous membrane onto which the compounds are printed is optically clear and preferably composed as stated in Appendix I. In another embodiment of this aspect of the present invention the detector layer of cells is a continuous or discontinuous sheet of cells, without any separation into test units or wells, growing on a porous and optically clear

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membrane preferably of the types mentioned above. The porous membrane may allow the cells to send cellular processes through the pores of the membrane. The compounds are printed onto an optically clear substratum such as glass, plastic or quartz as solutions in solvent and allowed to dry. At the time of the experiment the cell sheet on the membrane, surrounded by a thin film of fluid, is layered ontop of the printed compound pattern. The compounds then dissolve and contact the cells via diffusion and convection. The compounds may be made using combinatorial chemistry techniques, and may be peptides. The compounds may be covalently attached to the optically clear substratum or porous membrane. The compounds may also be proteins, polypeptides or peptides secreted by or attached to the plasma membrane of genetically modified cells growing as a continuous or discontinuous sheet on a flat optically clear surface or an optically clear porous membrane.

The recording of the variation or result with respect to light emitted from the luminophore is performed by recording the spatially distributed light as one or more digital images, and the processing of the recorded variation to reduce it to one or more numbers representative of the degree of redistribution comprises a digital image processing procedure or combination of digital image processing procedures. The quantitative information which is indicative of the degree of the cellular response to the influence or the result of the influence on the intracellular pathway is extracted from the recording or recordings according to a predetermined calibration based on responses or results, recorded in the same manner, to known degrees of a relevant specific influence. This calibration procedure is developed according to principles described below (Developing an Image-based Assay Technique). Specific descriptions of the procedures for particular assays are given in the examples.

While the stepwise procedure necessary to reduce the image or images to the value representative of the response caused by the influence is particular to each assay, the individual steps are generally well-known methods of image processing. Some examples of the individual steps are point operations such as subtraction, ratioing, and thresholding, digital filtering methods such as smoothing, sharpening, and edge detection, spatial frequency methods such as Fourier filtering, image cross-correlation and image autocorrelation, object finding and classification (blob analysis), and colour space manipulations for visualisation. In addition to the algorithmic procedures, heuristic

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methods such as neural networks may also be used. In a preferred embodiment of the invention, a dose-response relationship is established based on quantification of the responses caused by a particular influence, representative of the underlying intracellular signalling process, using the methods described above and in examples 1-22 and 25. The dose-response relationship for the particular influence is then compared to the dose-response relationship obtained by performing the same assay in an instrument which allows parallel monitoring of all wells in a microtiter plate such as a FLIPRTM or an ordinary fluorescence plate reader for microtiter plates. If a good correlation between the dose-response relationships obtained from the two different measurement systems is obtained, it can be said that the parallel measurement mode has been validated (see examples 23 and 24). This implies that it can be used as the primary basis for a screening assay with the potential benefit of screening a significantly higher number of substances per unit of time for their influence on the response.

Imaging plate readers integrate the signal from each well into a single value per time point. Thus the data resulting from a single "run" of the instrument is a set of time series of single values, one for each well, with the injection of the test compound taking place at a known point in the time series. The primary advantage of this type of instrumentation is that it greatly increases the number of samples that can be processed in a given amount of time (the throughput). This is of great advantage when using the assay in a screening program for new pharmaceutical lead compounds.

The first step in the data analysis is to normalise the results from each well so that they can be compared with each other or with previously analysed known compounds. This always begins with correcting the signal by subtracting the instrument bias from all data points on a well-by-well basis. From this point, either of two techniques can be followed depending on the design of the assay:

Procedure 1: The average of the signal prior to the addition of the test compound is subtracted from all data points on a well-by-well basis.

Procedure 2: The data are corrected for any known background by subtracting the background value from all data points on a well-by-well basis. The resulting background-corrected data are normalised by dividing each data set by the average of the data values

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prior to the injection of the test compound on a well-by-well basis.

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The corrected or normalised time series data sets are then further reduced by a technique that converts the time series to a single value. There are at least three such approaches:

- 1. For transient responses, the maximum deviation from the baseline is determined. This is also known as the "peak height" technique.
 - 2. Alternatively, the signal is integrated over time between pre-defined limits. If the data were treated according to Procedure 2 above, then the offset is subtracted such that the integral of a non-response is zero within the limit of measurement error. This is also known as the "peak area" technique.
- 3. If the response is a cumulative one, e.g., an exponential change to a new level, the result is taken as the either the difference or the ratio between the signal after a predetermined time and the signal prior to the addition of the test compound.

All of the above procedures reduce the data for a given well to one or more single values. For screening purposes, these values will be searched for those that are greater than a certain statistically determined cut-off value. For characterisation, the values represent a quantitative response, and are further treated in sets by techniques such as dose-response curve fitting.

In another embodiment of the invention, the measurement of redistribution is accomplished indirectly by taking advantage of the fact that in order for redistribution to occur, the probe will experience some change in its freedom, or restriction, of movement within the intracellular milieu. The degree of translocation will correlate with the amount of freely mobile luminophore in the cytoplasm. At a point in time after the test compound has begun to have any influence it may have, the amount or fraction of restricted luminophore can be measured by disrupting or permeabilising the plasma membrane of the cells and allowing the freely mobile luminophore to diffuse away. If the detection volume of the detector is limited to the region immediately surrounding the cells, and the overall volume into which the freely mobile luminophore can diffuse is much larger, then the freely mobile luminophore essentially disappears from the detector's view and its signal is

not recorded.

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In one aspect of the invention, the above mentioned measurement of redistribution is made on cells with permanently permeabilised plasma membranes immersed in a solution mimicking the cytoplasmic environment. In this way the influence of compounds that can normally not enter the cytoplasm of cells can be tested.

The nucleic acid constructs used in the present invention encode in their nucleic acid sequences fusion polypeptides comprising a biologically active polypeptide that is a component of an intracellular signalling pathway, or a part thereof, and a GFP, preferably an F64L mutant of GFP, N- or C-terminally fused, optionally via a peptide linker, to the biologically active polypeptide or part thereof.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a protein kinase or a phosphatase.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a transcription factor or a part thereof which changes cellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a protein, or a part thereof, which is associated with the cytoskeletal network and which changes cellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a protein kinase or a part thereof which changes cellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a serine/threonine protein kinase or a part thereof capable of changing intracellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a tyrosine protein kinase or a part thereof capable of changing intracellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a phospholipid-dependent serine/threonine protein kinase or a part thereof capable of changing intracellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a cAMP-dependent protein kinase or a part thereof capable of changing cellular localisation upon activation. In a preferred embodiment the biologically active polypeptide encoded by the nucleic acid construct is a PKAc-F64L-S65T-GFP fusion.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a cGMP-dependent protein kinase or a part thereof capable of changing cellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a calmodulin-dependent serine/threonine protein kinase or a part thereof capable of changing cellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a mitogen-activated serine/threonine protein kinase or a part thereof capable of changing cellular localisation upon activation. In preferred embodiments the biologically active polypeptide encoded by the nucleic acid constructs are an ERK1-F64L-S65T-GFP fusion or an EGFP-ERK1 fusion.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a cyclin-dependent serine/threonine protein kinase or a part thereof capable of changing cellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a protein phosphatase or a part thereof capable of changing cellular localisation upon activation.

In one preferred embodiment of the invention the nucleic acid constructs may be DNA constructs.

In one embodiment the biologically active polypeptide encoded by the nucleic acid

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construct. In one embodiment the gene encoding GFP in the nucleic acid construct is derived from Aequorea victoria. In a preferred embodiment the gene encoding GFP in the nucleic acid construct is EGFP or a GFP variant selected from F64L-GFP, F64L-Y66H-GFP and F64L-S65T-GFP.

In preferred embodiments of the invention the DNA constructs which can be identified by any of the DNA sequences shown in SEQ ID NO: 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, 78, 108, 110, 112, 114, 116, 118, 120, 122, 124, 126, 128, 130, 132, 134, 136, 138, 140, 142, 144, 146, 148, 150, and 152 or are variants of these sequences capable of encoding the same fusion polypeptide or a fusion polypeptide which is biologically equivalent thereto, e.g. an isoform, or a splice variant or a homologue from another species.

The present invention describes a method that may be used to establish a screening program for the identification of biologically active substances that directly or indirectly affects intracellular signalling pathways and because of this property are potentially useful as medicaments. Based on measurements in living cells of the redistribution of spatially resolved luminescence from luminophores which undergo a change in distribution upon activation or deactivation of an intracellular signalling pathway the result of the individual measurement of each substance being screened indicates its potential biological activity.

In one embodiment of the invention the screening program is used for the identification of a biologically toxic substance as defined herein that exerts its toxic effect by interfering with an intracellular signalling pathway. Based on measurements in living cells of the redistribution of spatially resolved luminescence from luminophores which undergo a change in distribution upon activation or deactivation of an intracellular signalling pathway the result of the individual measurement of each substance being screened indicates its potential biologically toxic activity. In one embodiment of a screening program a compound that modulates a component of an intracellular pathway as defined herein, can be found and the therapeutic amount of the compound estimated by a method according to the method of the invention. In a preferred embodiment the present invention leads to the discovery of a new way of treating a condition or disease related to the intracellular function of a biologically active polypeptide comprising administration to a

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patient suffering from said condition or disease of an effective amount of a compound which has been discovered by any method according to the invention. In another preferred embodiment of the invention a method is established for identification of a new drug target or several new drug targets among the group of biologically active polypeptides which are components of intracellular signalling pathways.

In another embodiment of the invention an individual treatment regimen is established for the selective treatment of a selected patient suffering from an ailment where the available medicaments used for treatment of the ailment are tested on a relevant primary cell or cells obtained from said patient from one or several tissues, using a method comprising transfecting the cell or cells with at least one DNA sequence encoding a fluorescent probe according to the invention, transferring the transfected cell or cells back the said patient, or culturing the cell or cells under conditions permitting the expression of said probes and exposing it to an array of the available medicaments, then comparing changes in fluorescence patterns or redistribution patterns of the fluorescent probes in the intact living cells to detect the cellular response to the specific medicaments (obtaining a cellular action profile), then selecting one or more medicament or medicaments based on the desired activity and acceptable level of side effects and administering an effective amount of these medicaments to the selected patient.

The present invention describes a method that may be used to establish a screening program for back-tracking signal transduction pathways as defined herein. In one embodiment the screening program is used to establish more precisely at which level one or several compounds affect a specific signal transduction pathway by successively or in parallel testing the influence of the compound or compounds on the redistribution of spatially resolved luminescence from several of the luminophores which undergo a change in distribution upon activation or deactivation of the intracellular signalling pathway under study.

In general, a probe, i.e. a "GeneX"-GFP fusion or a GFP-"GeneX" fusion, is constructed using PCR with "GeneX"-specific primers followed by a cloning step to fuse "GeneX" in frame with GFP. The fusion may contain a short vector derived sequence between "GeneX" and GFP (e.g. part of a multiple cloning site region in the plasmid) resulting in a

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peptide linker between "GeneX" and GFP in the resulting fusion protein.

Some of the steps involved in the development of a probe include the following:

- Identify the sequence of the gene. This is most readily done by searching a depository of genetic information, e.g. the GenBank Sequence Database, which is widely available and routinely used by molecular biologists. In the specific examples below the GenBank Accession number of the gene in question is provided.
 - Design the gene-specific primers. Inspection of the sequence of the gene allows design of gene-specific primers to be used in a PCR reaction. Typically, the top-strand primer encompasses the ATG start codon of the gene and the following ca. 20 nucleotides, while the bottom-strand primer encompasses the stop codon and the ca. 20 preceding nucleotides, if the gene is to be fused behind GFP, i.e. a GFP-"GeneX" fusion. If the gene is to be fused in front of GFP, i.e. a "GeneX"-GFP fusion, a stop codon must be avoided. Optionally, the full-length sequence of GeneX may not be used in the fusion, but merely the part that localizes and redistributes like GeneX in response to a signal. In addition to gene-specific sequences, the primers contain at least one recognition sequence for a restriction enzyme, to allow subsequent cloning of the PCR product. The sites are chosen so that they are unique in the PCR product and compatible with sites in the cloning vector. Furthermore, it may be necessary to include an exact number of nucleotides between the restriction enzyme site and the gene-specific sequence in order to establish the correct reading frame of the fusion gene and/or a translation initiation consensus sequence. Lastly, the primers always contain a few nucleotides in front of the restriction enzyme site to allow efficient digestion with the enzyme.
- Identify a source of the gene to be amplified. In order for a PCR reaction to produce a product with gene-specific primers, the gene-sequence must initially be present in the reaction, e.g. in the form of cDNA. Information in GenBank or the scientific literature will usually indicate in which tissue(s) the gene is expressed, and cDNA libraries from a great variety of tissues or cell types from various species are commercially available, e.g. from Clontech (Palo Alto), Stratagene (La Jolla) and Invitrogen (San Diego).
 Many genes are also available in cloned form from The American Type Tissue

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Collection (Virginia).

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Optimise the PCR reaction. Several factors are known to influence the efficiency and specificity of a PCR reaction, including the annealing temperature of the primers, the concentration of ions, notably Mg²⁺ and K⁺, present in the reaction, as well as pH of the reaction. If the result of a PCR reaction is deemed unsatisfactory, it might be because the parameters mentioned above are not optimal. Various annealing temperatures should be tested, e.g. in a PCR machine with a built-in temperature gradient, available from e.g. Stratagene (La Jolla), and/or various buffer compositions should be tried, e.g. the OptiPrime buffer system from Stratagene (La Jolla).

Clone the PCR product. The vector into which the amplified gene product will be cloned and fused with GFP will already have been taken into consideration when the primers were designed. When choosing a vector, one should at least consider in which cell types the probe subsequently will be expressed, so that the promoter controlling expression of the probe is compatible with the cells. Most expression vectors also contain one or more selective markers, e.g. conferring resistance to a drug, which is a useful feature when one wants to make stable transfectants. The selective marker should also be compatible with the cells to be used.

The actual cloning of the PCR product should present no difficulty as it typically will be a one-step cloning of a fragment digested with two different restriction enzymes into a vector digested with the same two enzymes. If the cloning proves to be problematic, it may be because the restriction enzymes did not work well with the PCR fragment. In this case one could add longer extensions to the end of the primers to overcome a possible difficulty of digestion close to a fragment end, or one could introduce an intermediate cloning step not based on restriction enzyme digestion. Several companies offer systems for this approach, e.g. Invitrogen (San Diego) and Clontech (Palo Alto).

Once the gene has been cloned and, in the process, fused with the GFP gene, the resulting product, usually a plasmid, should be carefully checked to make sure it is as expected. The most exact test would be to obtain the nucleotide sequence of the fusion-gene.

Once a DNA construct for a probe has been generated, its functionality and usefulness may

be evaluated by transfecting it into cells capable of expressing the probe. The fluorescence of the cell is inspected soon after, typically the next day. At this point, two features of cellular fluorescence are noted: the intensity and the sub-cellular localisation.

The intensity should usually be at least as strong as that of unfused GFP in the cells. If it is not, the sequence or quality of the probe-DNA might be faulty, and should be carefully checked.

The sub-cellular localisation is an indication of whether the probe is likely to perform well. If it localises as expected for the gene in question, e.g. is excluded from the nucleus, it can immediately go on to a functional test. If the probe is not localised soon after the transfection procedure, it may be because of overexpression at this point in time, as the cell typically will have taken up very many copies of the plasmid, and localisation will occur in time, e.g. within a few weeks, as plasmid copy number and expression level decreases. If localisation does not occur after prolonged time, it may be because the fusion to GFP has destroyed a localisation function, e.g. masked a protein sequence essential for interaction with its normal cellular anchor-protein. In this case the opposite fusion might work, e.g. if GeneX-GFP does not work, GFP-GeneX might, as two different parts of GeneX will be affected by the proximity to GFP. If this does not work, the proximity of GFP at either end might be a problem, and it could be attempted to increase the distance by incorporating a longer linker between GeneX and GFP in the DNA construct.

If there is no prior knowledge of localisation, and no localisation is observed, it may be because the probe should not be localised at this point, because such is the nature of the protein fused to GFP. It should then be subjected to a functional test.

In a functional test, the cells expressing the probe are treated with at least one compound known to perturb, usually by activating, the signalling pathway on which the probe is expected to report by redistributing itself within the cell. If the redistribution is as expected, e.g. if prior knowledge tell that it should translocate from location X to location Y, it has passed the first critical test. In this case it can go on to further characterisation and quantification of the response.

If it does not perform as expected, it may be because the cell lacks at least one component

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of the signalling pathway, e.g. a cell surface receptor, or there is species incompatibility, e.g. if the probe is modelled on sequence information of a human gene product, and the cell is of hamster origin. In both instances one should identify other cell types for the testing process where these potential problems would not apply.

If there is no prior knowledge about the pattern of redistribution, the analysis of the redistribution will have to be done in greater depth to identify what the essential and indicative features are, and when this is clear, it can go on to further characterisation and quantification of the response. If no feature of redistribution can be identified, the problem might be as mentioned above, and the probe should be retested under more optimal cellular conditions.

If the probe does not perform under optimal cellular conditions, then it's back to the drawing board.

The process of developing an image-based redistribution assay begins with either the unplanned experimental observation that a redistribution phenomenon can be visualised, or the design of a probe specifically to follow a redistribution phenomenon already known to occur. In either event, the first and best exploratory technique is for a trained scientist or technician to observe the phenomenon. Even with the rapid advances in computing technology, the human eye-brain combination is still the most powerful pattern recognition system known, and requires no advance knowledge of the system in order to detect potentially interesting and useful patterns in raw data. This is especially if those data are presented in the form of images, which are the natural "data type" for human visual processing. Because human visual processing operates most effectively in a relatively narrow frequency range, i.e., we cannot see either very fast or very slow changes in our visual field, it may be necessary to record the data and play it back with either time dilation or time compression.

Some luminescence phenomena cannot be seen directly by the human eye. Examples include polarisation and fluorescence lifetime. However, with suitable filters or detectors, these signals can be recorded as images or sequences of images and displayed to the human in the fashion just described. In this way, patterns can be detected and the same methods can be applied.

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Once the redistribution has been determined to be a reproducible phenomenon, one or more data sets are generated for the purpose of developing a procedure for extracting the quantitative information from the data. In parallel, the biological and optical conditions are determined which will give the best quality raw data for the assay. This can become an iterative process; it may be necessary to develop a quantitative procedure in order to assess the effect on the assay of manipulating the assay conditions.

The data sets are examined by a person or persons with knowledge of the biological phenomenon and skill in the application of image processing techniques. The goal of this exercise is to determine or at least propose a method that will reduce the image or sequence of images constituting the record of a "response" to a value corresponding to the degree of the response. Using either interactive image processing software or an image processing toolbox and a programming language, the method is encoded as a procedure or algorithm that takes the image or images as input and generates the degree of response (in any units) as its output. Some of the criteria for evaluating the validity of a particular procedure are:

- Does the degree of the response vary in a biologically significant fashion, i.e., does
 it show the known or putative dependence on the concentration of the stimulating
 agent or condition?
- Is the degree of response reproducible, i.e., does the same concentration or level of stimulating agent or condition give the same response with an acceptable variance?
- Is the dynamic range of the response sufficient for the purpose of the assay? If not, can a change in the procedure or one of its parameters improve the dynamic range?
- Does the procedure exhibit any clear "pathologies", i.e., does it give ridiculous values for the response if there are commonly occurring imperfections in the imaging process? Can these pathologies be eliminated, controlled, or accounted for?
- Can the procedure deal with the normal variation in the number and/or size of cells in an image?

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In some cases the method may be obvious; in others, a number of possible procedures may suggest themselves. Even if one method appears clearly superior to others, optimisation of parameters may be required. The various procedures are applied to the data set and the criteria suggested above are determined, or the single procedure is applied repeatedly with adjustment of the parameter or parameters until the most satisfactory combination of signal, noise, range, etc. are arrived at. This is equivalent to the calibration of any type of single-channel sensor.

The number of ways of extracting a single value from an image are extremely large, and thus an intelligent approach must be taken to the initial step of reducing this number to a small, finite number of possible procedures. This is not to say that the procedure arrived at is necessarily the best procedure - but a global search for the best procedure is simply out of the question due to the sheer number of possibilities involved.

Image-based assays are no different than other assay techniques in that their usefulness is characterised by parameters such as the specificity for the desired component of the sample, the dynamic range, the variance, the sensitivity, the concentration range over which the assay will work, and other such parameters. While it is not necessary to characterise each and every one of these before using the assay, they represent the only way to compare one assay with another.

The final step is then to see whether there exists a possibility to increase the throughput of the assay to improve its utility as the basis of a screening program. In order to do this, a dose-response relationship is established based on quantification of the responses caused by a particular influence, representative of the underlying intracellular signalling process, using the methods described above and in examples 1-22 and 25. The dose-response relationship for the particular influence is then compared to the dose-response relationship obtained by performing the same assay in an instrument which allows parallel monitoring of all wells in a microtiter plate such as a FLIPRTM or an ordinary imaging or fluorescence plate reader for microtiter plates. If a good correlation between the dose-response relationships obtained from the two different measurement systems is obtained, it can be said that the parallel measurement mode has been validated (see examples 23 and 24). This implies that it can be used as the primary basis for a screening program with the potential

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benefit of screening a significantly higher number of substances for their influence on the response per unit of time.

The process of developing an image-based assay is best illustrated by example. The development of such an assay for GLUT4 translocation is hereby described. GLUT4 is a member of the class of glucose transporter molecules that are important in cellular glucose uptake. It is known to translocate to the plasma membrane under some conditions of stimulation of glucose uptake. The ability to visualise the glucose uptake response non-invasively, without actually measuring glucose uptake, would be a very useful assay for anyone looking for, for example, treatments for type II diabetes.

A CHO cell line which stably expressed the human insulin receptor was used as the basis for a new cell line which stably expressed a fusion between GLUT4 and GFP. This cell line was expected to show translocation of GLUT4 to the plasma membrane as visualised by the movement of the GFP. The translocation could definitely be seen in the form of the appearance of local increases in the fluorescence in regions of the plasma membrane which had a characteristic shape or pattern. This is shown in Figure 12.

These objects became known as "snircles", and the phenomenon of their appearance as "snircling". In order to quantify their appearance, a method had to be found to isolate them as objects in the image field, and then enumerate them, measure their area, or determine some parameter about them which correlated in a dose-dependent fashion with the concentration of insulin to which the cells had been exposed. In order to separate the snircles, a binarization procedure was applied in which one copy of the image smoothed with a relatively severe gaussian kernel (sigma = 2.5) was subtracted from another copy to which only a relatively light gaussian smooth had been applied (sigma=0.5). The resultant image was rescaled to its min/max range, and an automatic threshold was applied to divide the image into two levels. The thresholded image contains a background of one value all found object with another value. The found objects were first filtered through a filter to remove objects far too large and far too small to be snircles. The remaining objects, which represent snircles and other artifacts from the image with approximately the same size and intensity characteristics as snircles, are passed into a classification procedure which has been previously trained with many images of snircles to recognize snircles and exclude the

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other artifacts. The result of this procedure is a binary image that shows only the found snircles to the degree to which the classification procedure can accurately identify them. The total area of the snircles is then summed and this value is the quantitative measure of the degree of snircling for that image.

Another approach to the problem of quantifying GLUT 4 translocation has been performed and validated using the same type of experimental protocol but a different image processing approach. In this case the objects of interest in the cells are not the appearance of snircles at the plasma membrane but the disappearance of GLUT4-GFP fluorescence from its intracellular site. With this method the bright area, consisting of GLUT4-GFP, centrally located in each cell is identified by a thresholding procedure. This demarcates a certain area for the centrally located GLUT4-GFP. In the next step the total fluorescence intensity in this area is quantified on each image in the image series, i.e. over time. The response for each cell is defined as the difference in fluorescence intensity in the centrally located GLUT4-GFP area before and a fixed point in time after application of the influence. The dose-response relationship for insulin using the above described quantitation procedure is shown in Figure 13. It can be seen that the ED50 value for insulin to reduce central GLUT4-GFP fluorescence is 0.3 nM.

In the present specification and claims, the term "an influence" covers any influence to which the cellular response comprises a redistribution. Thus, e.g., heating, cooling, high pressure, low pressure, humidifying, or drying are influences on the cellular response on which the resulting redistribution can be quantified, but as mentioned above, perhaps the most important influences are the influences of contacting or incubating the cells with substances which are known or suspected to exert an influence on the cellular response involving a redistribution contribution. In another embodiment of the invention the influence could be substances from a compound drug library.

In the present context, the term "green fluorescent protein" is intended to indicate a protein which, when expressed by a cell, emits fluorescence upon exposure to light of the correct excitation wavelength (cf. [(Chalfie, M. et al. (1994) Science 263, 802-805)]). In the following, GFP in which one or more amino acids have been substituted, inserted or deleted is most often termed "modified GFP". "GFP" as used herein includes wild-type

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GFP derived from the jelly fish *Aequorea victoria* and modifications of GFP, such as the blue fluorescent variant of GFP disclosed by Heim *et al.* (1994). Proc.Natl.Acad.Sci. 91:26, pp 12501-12504, and other modifications that change the spectral properties of the GFP fluorescence, or modifications that exhibit increased fluorescence when expressed in cells at a temperature above about 30°C described in PCT/DK96/00051, published as WO 97/11094 on 27 March 1997 and hereby incorporated by reference, and which comprises a fluorescent protein derived from *Aequorea* Green Fluorescent Protein (GFP) or any functional analogue thereof, wherein the amino acid in position 1 upstream from the chromophore has been mutated to provide an increase of fluorescence intensity when the fluorescent protein of the invention is expressed in cells. Preferred GFP variants are F64L-GFP, F64L-Y66H-GFP and F64L-S65T-GFP. An especially preferred variant of GFP for use in all the aspects of this invention is EGFP (DNA encoding EGFP which is a F64L-S65T variant with codons optimized for expression in mammalian cells is available from Clontech, Palo Alto, plasmids containing the EGFP DNA sequence, cf. GenBank Acc. Nos. U55762, U55763).

The term "intracellular signalling pathway" and "signal transduction pathway" are intended to indicate the co-ordinated intracellular processes whereby a living cell transduce an external or internal signal into cellular responses. Said signal transduction will involve an enzymatic reaction said enzymes include but are not limited to protein kinases, GTPases, ATPases, protein phosphatases, phospholipases and cyclic nucleotide phosphodiesterases. The cellular responses include but are not limited to gene transcription, secretion, proliferation, mechanical activity, metabolic activity, cell death.

The term "second messenger" is used to indicate a low molecular weight component involved in the early events of intracellular signal transduction pathways.

The term "luminophore" is used to indicate a chemical substance that has the property of emitting light either inherently or upon stimulation with chemical or physical means. This includes but is not limited to fluorescence, bioluminescence, phosphorescence, and chemiluminescence.

The term "mechanically intact living cell" is used to indicate a cell which is considered living according to standard criteria for that particular type of cell such as maintenance of normal membrane potential, energy metabolism, proliferative capability, and has not

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experienced any physically invasive treatment designed to introduce external substances into the cell such as microinjection.

In the present context, the term "permeabilised living cell" is used to indicate cells where a pore forming agent such as Streptolysin O or Staphylococcus Aureus α-toxin has been applied and thereby incorporated into the plasma membrane in the cells. This creates proteinaceous pores with a defined pore size in the plasma membranes of the exposed cells. Pores could also be made by electroporation, i.e. exposing the cells to high voltage discharges, a procedure that creates small holes in the plasma membrane by coagulating integral membrane proteins. Treatment with a mild detergent such as saponin may accomplish the same thing. Common to all these treatments are that pores are formed only in the plasma membrane without affecting the integrity of cytoplasmic structural elements and organelles. The term living in this context means that the permeabilised cells bathed in a solution mimicking the intracellular milieu still have functional organelles, such as actively respiring mitochondria and endoplasmic reticulum that can take up and release calcium ions, and functional structural elements. The benefit of this method is that substances that normally can not traverse the plasma membrane, but most likely exert their influence intracellularly, can be introduced and their influence studied without cumbersome microinjection of the substances into single cells. Using this method the response to an influence can be recorded from many cells simultaneously.

In the present context, the term "permeabilisation" is intended to indicate the selective disruption of the plasma membrane barrier so that soluble substances freely mobile in the cytosol are lost from the cells. The permeabilisation can be achieved as described above under "permeabilised living cells" or by using other chemical detergents such as Triton X-100 or digitonin in carefully titrated amounts.

The term "physiologically relevant", when applied to an experimentally determined redistribution of an intracellular component, as measured by a change in the luminescence properties or distribution, is used to indicate that said redistribution can be explained in terms of the underlying biological phenomenon which gives rise to the redistribution.

The terms "image processing" and "image analysis" are used to describe a large family of digital data analysis techniques or combination of such techniques which reduce ordered

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arrays of numbers (images) to quantitative information describing those ordered arrays of numbers. When said ordered arrays of numbers represent measured values from a physical process, the quantitative information derived is therefore a measure of the physical process.

The term "fluorescent probe" is used to indicate a fluorescent fusion polypeptide comprising a GFP or any functional part thereof which is N- or C-terminally fused to a biologically active polypeptide as defined herein, optionally via a peptide linker consisting of one or more amino acid residues, where the size of the linker peptide in itself is not critical as long as the desired functionality of the fluorescent probe is maintained. A fluorescent probe according to the invention is expressed in a cell and basically mimics the physiological behaviour of the biologically active polypeptide moiety of the fusion polypeptide.

The term "mammalian cell" is intended to indicate any living cell of mammalian origin. The cell may be an established cell line, many of which are available from The American Type Culture Collection (ATCC, Virginia, USA) or a primary cell with a limited life span derived from a mammalian tissue, including tissues derived from a transgenic animal, or a newly established immortal cell line derived from a mammalian tissue including transgenic tissues, or a hybrid cell or cell line derived by fusing different cell types of mammalian origin e.g. hybridoma cell lines. The cells may optionally express one or more non-native gene products, e.g. receptors, enzymes, enzyme substrates, prior to or in addition to the fluorescent probe. Preferred cell lines include but are not limited to those of fibroblast origin, e.g. BHK, CHO, BALB, or of endothelial origin, e.g. HUVEC, BAE (bovine artery endothelial), CPAE (cow pulmonary artery endothelial), HLMVEC (human lung microvascular endothelial cells) or of pancreatic origin, e.g. RIN, INS-1, MIN6, bTC3, aTC6, bTC6, HIT, or of hematopoietic origin, e.g.primary isolated human monocytes, macrophages, neutrophils, basophils, eosinophils and lyphocyte populations, AML-193, HL-60, RBL-1, adipocyte origin, e.g. 3T3-L1, neuronal/neuroendocrine origin, e.g. AtT20, PC12, GH3, muscle origin, e.g. SKMC, A10, C2C12, renal origin, e.g. HEK 293, LLC-PK1.

The term "hybrid polypeptide" is intended to indicate a polypeptide which is a fusion of at

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least a portion of each of two proteins, in this case at least a portion of the green fluorescent protein, and at least a portion of a catalytic and/or regulatory domain of a protein kinase. Furthermore a hybrid polypeptide is intended to indicate a fusion polypeptide comprising a GFP or at least a portion of the green fluorescent protein that contains a functional fluorophore, and at least a portion of a biologically active polypeptide as defined herein provided that said fusion is not the PKCα-GFP, PKCγ-GFP, and PKCε-GFP disclosed by Schmidt *et al.* and Sakai *et al.*, respectively. Thus, GFP may be N- or C-terminally tagged to a biologically active polypeptide, optionally via a linker portion or linker peptide consisting of a sequence of one or more amino acids. The hybrid polypeptide or fusion polypeptide may act as a fluorescent probe in intact living cells carrying a DNA sequence encoding the hybrid polypeptide under conditions permitting expression of said hybrid polypeptide.

The term "kinase" is intended to indicate an enzyme that is capable of phosphorylating a cellular component.

The term "protein kinase" is intended to indicate an enzyme that is capable of phosphorylating serine and/or threonine and/or tyrosine in peptides and/or proteins.

The term "phosphatase" is intended to indicate an enzyme that is capable of dephosphorylating phosphoserine and/or phosphothreonine and/or phosphotyrosine in peptides and/or proteins.

The term "cyclic nucleotide phosphodiesterase" is intended to indicate an enzyme that is capable of inactivating the second messengers cAMP and cGMP by hydrolysis of their 3'-ester bond.

In the present context, the term "biologically active polypeptide" is intended to indicate a polypeptide affecting intracellular processes upon activation, such as an enzyme which is active in intracellular processes or a portion thereof comprising a desired amino acid sequence which has a biological function or exerts a biological effect in a cellular system. In the polypeptide one or several amino acids may have been deleted, inserted or replaced to alter its biological function, e.g. by rendering a catalytic site inactive. Preferably, the biologically active polypeptide is selected from the group consisting of proteins taking part

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in an intracellular signalling pathway, such as enzymes involved in the intracellular phosphorylation and dephosphorylation processes including kinases, protein kinases and phosphorylases as defined herein, but also proteins making up the cytoskeleton play important roles in intracellular signal transduction and are therefore included in the meaning of "biologically active polypeptide" herein. More preferably, the biologically active polypeptide is a protein which according to its state as activated or non-activated changes localisation within the cell, preferably as an intermediary component in a signal transduction pathway. Included in this preferred group of biologically active polypeptides are cAMP dependent protein kinase A.

The term "a substance having biological activity" is intended to indicate any sample that has a biological function or exerts a biological effect in a cellular system. The sample may be a sample of a biological material such as a sample of a body fluid including blood, plasma, saliva, milk, urine, or a microbial or plant extract, an environmental sample containing pollutants including heavy metals or toxins, or it may be a sample containing a compound or mixture of compounds prepared by organic synthesis or genetic techniques.

The phrase "any change in fluorescence" means any change in absorption properties, such as wavelength and intensity, or any change in spectral properties of the emitted light, such as a change of wavelength, fluorescence lifetime, intensity or polarisation, or any change in the intracellular localisation of the fluorophore. It may thus be localised to a specific cellular component (e.g. organelle, membrane, cytoskeleton, molecular structure) or it may be evenly distributed throughout the cell or parts of the cell.

The term "organism" as used herein indicates any unicellular or multicellular organism preferably originating from the animal kingdom including protozoans, but also organisms that are members of the plant kingdoms, such as algae, fungi, bryophytes, and vascular plants are included in this definition.

The term "nucleic acid" is intended to indicate any type of poly- or oligonucleic acid sequence, such as a DNA sequence, a cDNA sequence, or an RNA sequence.

The term "biologically equivalent" as it relates to proteins is intended to mean that a first protein is equivalent to a second protein if the cellular functions of the two proteins may

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substitute for each other, e.g. if the two proteins are closely related isoforms encoded by different genes, if they are splicing variants, or allelic variants derived from the same gene, if they perform identical cellular functions in different cell types, or in different species. The term "biologically equivalent" as it relates to DNA is intended to mean that a first DNA sequence encoding a polypeptide is equivalent to a second DNA sequence encoding a polypeptide if the functional proteins encoded by the two genes are biologically equivalent.

The phrase "back-tracking of a signal transduction pathway" is intended to indicate a process for defining more precisely at what level a signal transduction pathway is affected, either by the influence of chemical compounds or a disease state in an organism. Consider a specific signal transduction pathway represented by the bioactive polypeptides A - B - C - D, with signal transduction from A towards D. When investigating all components of this signal transduction pathway compounds or disease states that influence the activity or redistribution of only D can be considered to act on C or downstream of C whereas compounds or disease states that influence the activity or redistribution of C and D, but not of A and B can be considered to act downstream of B.

The term "fixed cells" is used to mean cells treated with a cytological fixative such as glutaraldehyde or formaldehyde, treatments that serve to chemically cross-link and stabilise soluble and insoluble proteins within the structure of the cell. Once in this state, such proteins cannot be lost from the structure of the now-dead cell.

In the present context a "screening assay" is intended to mean any measurement protocol, including materials, cells, instruments, chemicals, reagents, detection units, calibration and quantification procedures used to measure a response from mechanically intact or permeabilised living cells relevant to influences on an intracellular pathway.

The term "dose-response relationship" and "screening programme" is in the present context intended to mean a clear correlation between the quantified response of cells in a screening assay to application of an influence, such as a compound, and the concentration of the applied influence. The response to the influence may be both an up-regulation and a down-regulation of the quantified parameter used in the screening assay.

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In the present context, the term "physiology" is intended to mean the normal function of biological and biochemical processes inside cells, between cells and in the whole organism or animal.

5 BRIEF DESCRIPTION OF THE DRAWINGS

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Figure 1. CHO cells expressing the PKAc-F64L-S65T-GFP hybrid protein have been treated in HAM's F12 medium with 50 μ M forskolin at 37°C. The images of the GFP fluorescence in these cells have been taken at different time intervals after treatment, which were: a) 40 seconds b) 60 seconds c) 70 seconds d) 80 seconds. The fluorescence changes from a punctate to a more even distribution within the (non-nuclear) cytoplasm.

Figure 2. Time-lapse analysis of forskolin induced PKAc-F64L-S65T-GFP redistribution. CHO cells, expressing the PKAc-F64L-S65T-GFP fusion protein were analysed by time-lapse fluorescence microscopy. Fluorescence micrographs were acquired at regular intervals from 2 min before to 8 min after the addition of agonist. The cells were challenged with 1 μ M forskolin immediately after the upper left image was acquired (t=0). Frames were collected at the following times: i) 0, ii) 1, iii) 2, iv) 3, v) 4 and vi) 5 minutes. Scale bar 10 μ m.

Figure 3. Time-lapse analyses of PKAc-F64L-S65T-GFP redistribution in response to various agonists. The effects of 1 μM forskolin (A), 50 μM forskolin (B), 1mM dbcAMP (C) and 100 μM IBMX (D) (additions indicated by open arrows) on the localisation of the PKAc-F64L-S65T-GFP fusion protein were analysed by time-lapse fluorescence microscopy of CHO/PKAc-F64L-S65T-GFP cells. The effect of addition of 10 μM forskolin (open arrow), followed shortly by repeated washing with buffer (solid arrow), on the localisation of the PKAc-F64L-S65T-GFP fusion protein was analysed in the same cells (E). In a parallel experiment, the effect of adding 10 μM forskolin and 100 μM IBMX (open arrow) followed by repeated washing with buffer containing 100 μM IBMX

(solid arrow) was analysed (F). Removing forskolin caused PKAc-F64L-S65T-GFP fusion protein to return to the cytoplasmic aggregates while this is prevented by the continued presence of IBMX (F). The effect of 100 nM glucagon (Fig 3G, open arrow) on the localisation of the PKAc-F64L-S65T-GFP fusion protein is also shown for BHK/GR, PKAc-F64L-S65T-GFP cells. The effect of 10 μ M norepinephrine (H), solid arrow, on the localisation of the PKAc-F64L-S65T-GFP fusion protein was analysed similarly, in transiently transfected CHO, PKAc-F64L-S65T-GFP cells, pretreated with 10 μ M forskolin, open arrow, to increase [cAMP]. N.B. in Fig 3H the x-axis counts the image numbers, with 12 seconds between images. The raw data of each experiment consisted of 60 fluorescence micrographs acquired at regular intervals including several images acquired before the addition of buffer or agonist. The charts (A-G) each show a quantification of the response seen through all the 60 images, performed as described in analysis method 2. The change in total area of the highly fluorescent aggregates, relative to the initial area of fluorescent aggregates is plotted as the ordinate in all graphs in Figure 3, versus time for each experiment. Scale bar 10 μ m.

Figure 4. Dose-response curve (two experiments) for forskolin-induced redistribution of the PKAc-F64L-S65T-GFP fusion.

Figure 5. Time from initiation of a response to half maximal (t_{1/2max}) and maximal (t_{max}) PKAc-F64L-S65T-GFP redistribution. The data was extracted from curves such as that shown in "Figure 2." All t_{1/2max} and t_{max} values are given as mean±SD and are based on a total of 26-30 cells from 2-3 independent experiments for each forskolin concentration. Since the observed redistribution is sustained over time, the t_{max} values were taken as the earliest time point at which complete redistribution is reached. Note that the values do not relate to the degree of redistribution.

Figure 6. Parallel dose-response analyses of forskolin induced cAMP elevation and PKAc-

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F64L-S65T-GFP redistribution. The effects of buffer or 5 increasing concentrations of forskolin on the localisation of the PKAc-F64L-S65T-GFP fusion protein in CHO/PKAc-F64L-S65T-GFP cells, grown in a 96 well plate, were analysed as described above. Computing the ratio of the SD's of fluorescence micrographs taken of the same field of cells, prior to and 30 min after the addition of forskolin, gave a reproducible measure of PKAc-F64L-S65T-GFP redistribution. The graph shows the individual 48 measurements and a trace of their mean±s.e.m at each forskolin concentration. For comparison, the effects of buffer or 8 increasing concentrations of forskolin on [cAMP], was analysed by a scintillation proximity assay of cells grown under the same conditions. The graph shows a trace of the mean ± s.e.m of 4 experiments expressed in arbitrary units.

Figure 7. BHK cells stably transfected with the human muscarinic (hM1) receptor and the PKC α -F64L-S65T-GFP fusion. Carbachol (100 μ M added at 1.0 second) induced a transient redistribution of PKC α -F64L-S65T-GFP from the cytoplasm to the plasma membrane. Images were taken at the following times: a) 1 second before carbachol addition, b) 8.8 seconds after addition and c) 52.8 seconds after addition.

Figure 8. BHK cells stably transfected with the hM1 receptor and PKC α -F64L-S65T-GFP fusion were treated with carbachol (1 μ M, 10 μ M, 100 μ M). In single cells intracellular [Ca²+] was monitored simultaneously with the redistribution of PKC α -F64L-S65T-GFP. Dashed line indicates the addition times of carbachol. The top panel shows changes in the intracellular Ca²+ concentration of individual cells with time for each treatment. The middle panel shows changes in the average cytoplasmic GFP fluorescence for individual cells against time for each treatment. The bottom panel shows changes in the fluorescence of the periphery of single cells, within regions that specifically include the circumferential edge of a cell as seen in normal projection, the best regions for monitoring changes in the fluorescence intensity of the plasma membrane.

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Figure 9.

- a) The hERK1-F64L-S65T-GFP fusion expressed in HEK293 cells treated with 100 μM of the MEK1 inhibitor PD98059 in HAM F-12 (without serum) for 30 minutes at 37 °C. The nuclei empty of fluorescence during this treatment.
- 5 b) The same cells as in (a) following treatment with 10 % foetal calf serum for 15 minutes at 37 °C.
 - c) Time profiles for the redistribution of GFP fluorescence in HEK293 cells following treatment with various concentrations of EGF in Hepes buffer (HAM F-12 replaced with Hepes buffer directly before the experiment). Redistribution of fluorescence is expressed as the change in the ratio value between areas in nucleus and cytoplasm of single cells. Each time profile is the mean for the changes seen in six single cells.
 - d) Bar chart for the end-point measurements, 600 seconds after start of EGF treatments, of fluorescence change (nucleus:cytoplasm) following various concentrations of EGF.

15 Figure 10.

- a) The SMAD2-EGFP fusion expressed in HEK293 cells starved of serum overnight in HAM F-12. HAM F-12 was then replaced with Hepes buffer pH 7.2 immediately before the experiment. Scale bar is $10~\mu m$.
- b) HEK 293 cells expressing the SMAD2-EGFP fusion were treated with various concentration of TGF-beta as indicated, and the redistribution of fluorescence monitored against time. The time profile plots represent increases in fluorescence within the nucleus, normalised to starting values in each cell measured. Each trace is the time profile for a single cell nucleus.
- c) A bar chart representing the end-point change in fluorescence within nuclei (after 850 seconds of treatment) for different concentrations of TGF-beta. Each bar is the value for a single nucleus in each treatment.

Figure 11. The VASP-F64L-S65T-GFP fusion in CHO cells stably transfected with the human insulin receptor. The cells were starved for two hours in HAM F-12 without serum, then treated with 10% foetal calf serum. The image shows the resulting redistribution of fluorescence after 15 minutes of treatment. GFP fluorescence becomes localised in structures identified as focal adhesions along the length of actin stress fibres.

Figure 12. Time lapse recording GLUT4-GFP redistribution in CHO-HIR cells. Time indicates minutes after the addition of 100 nM insulin.

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Figure 13. Dose-response relationships for the influence of insulin on the disappearance of total fluorescence from the centrally located area of GLUT4-GFP. Data points indicate mean±SE.

Figure 14. Dose-response relationship for the translocation of PKCα-GFP in BHKhM1 cells stimulated with the muscarinine agonist carbamylcholine using a FLIPR™ to do the actual experiments.

Figure 15. Dose-response relationship for the translocation of PKAc-GFP in CHO/PKAc-20 F64L-S65T-GFP cells stimulated with forskolin using a FLIPR™ to do the actual experiments.

Figure 16. Dose-response relationship for the disappearance of fluorescence from permeabilised CHO/PKAc-F64L-S65T-GFP when previously exposed to different doses of forskolin.

EXAMPLES

EXAMPLE 1

Construction, testing and implementation of an assay for cAMP based on PKA activation in real time within living cells.

Useful for monitoring the activity of signalling pathways that lead to altered concentrations of cAMP, e.g. activation of G-protein coupled receptors which couple to G-proteins of the G_s or G_t class.

The catalytic subunit of the murine cAMP dependent protein kinase (PKAc) was fused Cterminally to a F64L-S65T derivative of GFP. The resulting fusion (PKAc-F64L-S65T-GFP) was used for monitoring *in vivo* the translocation and thereby the activation of PKA.

To construct the PKAc-F64L-S65T-GFP fusion, convenient restriction endonuclease sites were introduced into the cDNAs encoding murine PKAc (Gen Bank Accession number: M12303) and F64L-S65T-GFP (sequence disclosed in WO 97/11094) by polymerase chain reaction (PCR). The PCR reactions were performed according to standard protocols with the following primers:

5'PKAc:

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 $\label{top:top:condition} TTggACACAAgCTTTggACACCCTCAggATATgggCAACgCCgCCgCCgCCAAg~(SEQ~ID~NO:3),$

20 3'PKAc:

gTCATCTTCTCgAgTCTTTCAggCgCgCCCAAACTCAgTAAACTCCTTgCCACAC (SEO ID NO:4) ,

5'GFP: TTggACACAAgCTTTggACACggCgCCCATgAgTAAAggAgAAGAACTTTTC (SEQ ID NO:1),

25 3'GFP: gTCATCTTCTCgAgTCTTACTCCTgAggTTTgTATAgTTCATCCATgCCATgT (SEQ ID NO:2).

The PKAc amplification product was then digested with HindIII+AscI and the F64L-S65T-GFP product with AscI+XhoI. The two digested PCR products were subsequently ligated with a HindIII+XhoI digested plasmid (pZeoSV® mammalian expression vector, Invitrogen, San Diego, CA, USA). The resulting fusion construct (SEQ ID NO:68 & 69) was under control of the SV40 promoter.

Transfection and cell culture conditions:

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Chinese hamster ovary cells (CHO), were transfected with the plasmid containing the PKAc-F64L-S65T-GFP fusion using the calcium phosphate precipitate method in HEPES-buffered saline (Sambrook *et al.*, 1989). Stable transfectants were selected using 1000 μg Zeocin/ml (Invitrogen) in the growth medium (DMEM with 1000 mg glucose/l, 10 % fetal bovine serum (FBS), 100 μg penicillin-streptomycin mixture ml⁻¹, 2 mM L-glutamine purchased from Life Technologies Inc., Gaithersburg, MD, USA). Untransfected CHO cells were used as the control. To assess the effect of glucagon on fusion protein translocation, the PKAc-F64L-S65T-GFP fusion was stably expressed in baby hamster kidney cells overexpressing the human glucagon receptor (BHK/GR cells). Untransfected BHK/GR cells were used as the control. Expression of GR was maintained with 500 μg G418/ml (*Neo* marker) and PKAc-F64L-S65T-GFP was maintained with 500 μg Zeocin/ml (*Sh ble* marker). CHO cells were also simultaneously co-transfected with vectors containing the PKAc-F64L-S65T-GFP fusion and the human α2a adrenoceptor (hARa2a).

For fluorescence microscopy, cells were allowed to adhere to Lab-Tek chambered coverglasses (Nalge Nunc Int., Naperville, IL, USA) for at least 24 hours and cultured to about 80% confluence. Prior to experiments, the cells were cultured over night without selection pressure in HAM F-12 medium with glutamax (Life Technologies), 100 µg penicillin-streptomycin mixture ml⁻¹ and 0.3 % FBS. This medium has low autofluorescence enabling fluorescence microscopy of cells straight from the incubator.

Monitoring activity of PKA activity in real time:

Image aquisition of live cells were gathered using a Zeiss Axiovert 135M fluorescence microscope fitted with a Fluar 40X, NA: 1.3 oil immersion objective and coupled to a Photometrics CH250 charged coupled device (CCD) camera. The cells were illuminated with a 100 W HBO arc lamp. In the light path was a 470±20 nm excitation filter, a 510 nm dichroic mirror and a 515±15 nm emission filter for minimal image background. The cells were maintained at 37°C with a custom built stage heater.

Images were processed and analysed in the following manner:

Method 1: Stepwise procedure for quantitation of translocation of PKA:

- 1. The image was corrected for dark current by performing a pixel-by-pixel subtraction of a dark image (an image taken under the same conditions as the actual image, except the camera shutter is not allowed to open).
- 2. The image was corrected for non-uniformity of the illumination by performing a pixel-by-pixel ratio with a flat field correction image (an image taken under the same conditions as the actual image of a uniformly fluorescent specimen).
- 3. The image histogram, i.e., the frequency of occurrence of each intensity value in the image, was calculated.
 - 4. A smoothed, second derivative of the histogram was calculated and the second zero is determined. This zero corresponds to the inflection point of the histogram on the high side of the main peak representing the bulk of the image pixel values.
- 5. The value determined in step 4 was subtracted from the image. All negative values were discarded.
 - 6. The variance (square of the standard deviation) of the remaining pixel values was determined. This value represents the "response" for that image.
 - 7. Scintillation proximity assay (SPA) for independent quantitation of cAMP.

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Method 2: Alternative method for quantitation of PKA redistribution:

- 1. The fluorescent aggregates are segmented from each image using an automatically found threshold based on the maximisation of the information measure between the object and background. The *a priori* entropy of the image histogram is used as the information measure.
- 5 2. The area of each image occupied by the aggregates is calculated by counting pixels in the segmented areas.
 - 3. The value obtained in step 2 for each image in a series, or treatment pair, is normalised to the value found for the first (unstimulated) image collected. A value of zero (0) indicates no redistribution of fluorescence from the starting condition. A value of one (1) by this method equals full redistribution.

Cells were cultured in HAM F-12 medium as described above, but in 96-well plates. The medium was exchanged with Ca²⁺-HEPES buffer including 100 µM IBMX and the cells were stimulated with different concentrations of forskolin for 10 min. Reactions were stopped with addition of NaOH to 0.14 M and the amount of cAMP produced was measured with the cAMP-SPA kit, RPA538 (Amersham) as described by the manufacturer.

Manipulating intracellular levels of cAMP to test the PKAc-F64L-S65T-GFP fusion.

The following compounds were used to vary cAMP levels: Forskolin, an activator of adenylate cyclase; dbcAMP, a membrane permeable cAMP analog which is not degraded by phosphodiesterase; IBMX, an inhibitor of phosphodiesterase.

CHO cells stably expressing the PKAc-F64L-S65T-GFP, showed a dramatic translocation of the fusion protein from a punctate distribution to an even distribution throughout the cytoplasm following stimulation with 1 μ M forskolin (n=3), 10 μ M forskolin (n=4) and 50 μ M forskolin (n=4) (Fig 1), or dbcAMP at 1mM (n=6).

Fig. 2 shows the progression of response in time following treatment with 1 μM forskolin.

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Fig. 3 gives a comparison of the average temporal profiles of fusion protein redistribution and a measure of the extent of each response to the three forskolin concentrations (Fig. 3A, E, B), and to 1 mM dbcAMP (fig 3C) which caused a similar but slower response, and to addition of $100 \, \mu M$ IBMX (n=4, Fig. 3D) which also caused a slow response, even in the absence of adenylate cyclase stimulation. Addition of buffer (n=2) had no effect (data not shown).

As a control for the behaviour of the fusion protein, F64L-S65T-GFP alone was expressed in CHO cells and these were also given 50 μ M forskolin (n=5); the uniform diffuse distribution characteristic of GFP in these cells was unaffected by such treatment (data not shown).

The forskolin-induced translocation of PKAc-F64L-S65T-GFP showed a dose-response relationship (Fig 4 and 6), see quantitative procedures above.

Reversibility of PKAc-F64L-S65T-GFP translocation.

The release of the PKAc probe from its cytoplasmic anchoring hotspots was reversible. Washing the cells repeatedly (5-8 times) with buffer after 10μM forskolin treatment completely restored the punctate pattern within 2-5 min (n=2, Fig. 3E). In fact the fusion protein returned to a pattern of fluorescent cytoplasmic aggregates virtually indistinguishable from that observed before forskolin stimulation.

To test whether the return of fusion protein to the cytoplasmic aggregates reflected a decreased [cAMP], cells were treated with a combination of 10 µM forskolin and 100 µM IBMX (n=2) then washed repeatedly (5-8 times) with buffer containing 100 µM IBMX (Fig. 3F). In these experiments, the fusion protein did not return to its prestimulatory localisation after removal of forskolin.

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Testing the PKA-F64L-S65T-GFP probe with physiologically relevant agents.

To test the probe's response to receptor activation of adenylate cyclase, BHK cells stably transfected with the glucagon receptor and the PKA-F64L-S65T-GFP probe were exposed to glucagon stimulation. The glucagon receptor is coupled to a G_s protein which activates adenylate cyclase, thereby increasing the cAMP level. In these cells, addition of 100 nM glucagon (n=2) caused the release of the PKA-F64L-S65T-GFP probe from the cytoplasmic aggregates and a resulting translocation of the fusion protein to a more even cytoplasmic distribution within 2-3 min (Fig. 3G). Similar but less pronounced effects were seen at lower glucagon concentrations (n=2, data not shown). Addition of buffer (n=2) had no effect over time (data not shown).

Transiently transfected CHO cells expressing hARα2a and the PKA-F64L-S65T-GFP probe were treated with 10 μM forskolin for 7.5 minutes, then, in the continued presence of forskolin, exposed to 10 μM norepinephrine to stimulate the exogenous adrenoreceptors, which couple to a G₁ protein, which inhibit adenylate cyclase. This treatment led to reappearance of fluorescence in the cytoplasmic aggregates indicative of a decrease in [cAMP], (Fig. 3H).

Fusion protein translocation correlated with [cAMP],

As described above, the time it took for a response to come to completion was dependent on the forskolin dose (Fig. 5) In addition the degree of responses was also dose-dependent. To test the PKA-F64L-S65T-GFP fusion protein translocation in a semi high through-put system, CHO cells stably transfected with the PKA-F64L-S65T-GFP fusion was stimulated with buffer and 5 increasing doses of forskolin (n=8). Using the image analysis algorithm described above (Method 1), a dose-response relationship was observed in the range from 0.01-50 μ M forskolin (Fig. 6). A half-maximal stimulation was observed at about 2 μ M forskolin. In parallel, cells were stimulated with buffer and 8 increasing concentrations of forskolin (n=4) in the range 0.01-50 μ M. The amount of cAMP produced was measured in an SPA assay. A steep increase was observed between 1 and 5 μ M forskolin coincident with the steepest part of the curve for fusion protein translocation (also Fig. 6).

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EXAMPLE 2

Quantitation of redistribution in real-time within living cells.

Probe for detection of PKC activity in real time within living cells:

5 Construction of PKC-GFP fusion:

The probe was constructed by ligating two restriction enzyme treated polymerase chain reaction (PCR) amplification products of the cDNA for murine PKCα (GenBank Accession number: M25811) and F64L-S65T-GFP (sequence disclosed in WO 97/11094) respectively. Taq® polymerase and the following oligonucleotide primers were used for PCR;

5'mPKCa:

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TTggACACAAgCTTTggACACCCTCAggATATggCTgACgTTTACCCggCCAACg (SEQ ID NO:5),

3'mPKCa:

gTCATCTTCTCgAgTCTTTCAggCgCgCCCTACTgCACTTTgCAAgATTgggTgC (SEQ ID NO:6),

5'F64L-S65T-GFP:

TTggACACAAgCTTTggACACggCgCCCATgAgTAAAggAgAAACTTTTC (SEQ ID NO:1).

20 3'F64L-S65T-GFP:

gTCATCTTCTCgAgTCTTACTCCTgAggTTTgTATAgTTCATCCATgCCATgT (SEQ ID NO:2).

The hybrid DNA strand was inserted into the pZeoSV® mammalian expression vector as a HindlII-XhoI casette as described in example 1.

BHK cells expressing the human M1 receptor under the control of the inducible metallothionine promoter and maintained with the dihydrofolate reductase marker were

transfected with the PKCα-F64L-S65T-GFP probe using the calcium phosphate precipitate method in HEPES buffered saline (HBS [pH 7.10]). Stable transfectants were selected using 1000 µg Zeocin®/ml in the growth medium (DMEM with 1000 mg glucose/l, 10 % foetal bovine serum (FBS), 100 µg penicillin-streptomycin mixture ml-1, 2 mM l-glutamine). The hM1 receptor and PKCα-F64L-S65T-GFP fusion protein were maintained with 500 nM methotrexate and 500 µg Zeocin®/ml respectively. 24 hours prior to any experiment, the cells were transferred to HAM F-12 medium with glutamax, 100 µg penicillin-streptomycin mixture ml-1 and 0.3 % FBS. This medium relieves selection pressure, gives a low induction of signal transduction pathways and has a low autofluorescence at the relevant wavelength enabling fluorescence microscopy of cells straight from the incubator.

Method 1: Monitoring the PKC α activity in real time:

Digital images of live cells were gathered using a Zeiss Axiovert 135M fluorescence microscope fitted with a 40X, NA: 1.3 oil immersion objective and coupled to a Photometrics CH250 charged coupled device (CCD) camera. The cells were illuminated with a 100 W arc lamp. In the light path was a 470±20 nm excitation filter, a 510 nm dichroic mirror and a 515±15 nm emission filter for minimal image background. The cells were kept and monitored to be at 37°C with a custom built stage heater.

Images were analyzed using the IPLab software package for Macintosh.

Upon stimulation of the M1-BHK cells, stably expressing the PKC α -F64L-S65T-GFP fusion, with carbachol we observed a dose-dependent transient translocation from the cytoplasm to the plasma membrane (Fig. 7a,b,c). Simultaneous measurement of the cytosolic free calcium concentration shows that the carbachol-induced calcium mobilisation precedes the translocation (Fig. 8).

Stepwise procedure for quantitation of translocation of PKC α :

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- 1. The image was corrected for dark current by performing a pixel-by-pixel subtraction of a dark image (an image taken under the same conditions as the actual image, except the camera shutter is not allowed to open).
- The image was corrected for non-uniformity of the illumination by performing a
 pixel-by-pixel ratio with a flat field correction image (an image taken under the same conditions as the actual image of a uniformly fluorescent specimen).
 - 3. A copy of the image was made in which the edges are identified. The edges in the image are found by a standard edge-detection procedure convolving the image with a kernel which removes any large-scale unchanging components (i.e., background) and accentuates any small-scale changes (i.e., sharp edges). This image was then converted to a binary image by threshholding. Objects in the binary image which are too small to represent the edges of cells were discarded. A dilation of the binary image was performed to close any gaps in the image edges. Any edge objects in the image which were in contact with the borders of the image are discarded. This binary image represents the edge mask.
 - 4. Another copy of image was made via the procedure in step 3. This copy was further processed to detect objects which enclose "holes" and setting all pixels inside the holes to the binary value of the edge, i.e., one. This image represents the whole cell mask.
- 5. The original image was masked with the edge mask from step 3 and the sum total of all pixel values is determined.
 - 6. The original image was masked with the whole cell mask from step 4 and the sum total of all pixel values was determined.
- 7. The value from step 5 was divided by the value from step 6 to give the final result, the fraction of fluorescence intensity in the cells which was localized in the edges.

EXAMPLE 3

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Probes for detection of mitogen activated protein kinase Erk1 redistribution.

Useful for monitoring signalling pathways involving MAPK, e.g. to identify compounds which modulate the activity of the pathway in living cells.

Erk1, a serine/threonine protein kinase, is a component of a signalling pathway that is activated by e.g. many growth factors.

Probes for detection of ERK-1 activity in real time within living cells:

The extracellular signal regulated kinase (ERK-1, a mitogen activated protein kinase, MAPK) is fused N- or C-terminally to a derivative of GFP. The resulting fusions expressed in different mammalian cells are used for monitoring *in vivo* the nuclear translocation, and thereby the activation, of ERK1 in response to stimuli that activate the MAPK pathway.

a) Construction of murine ERK1 - F64L-S65T-GFP fusion:

Convenient restriction endonuclease sites are introduced into the cDNAs encoding murine ERK1 (GenBank Accession number: Z14249) and F64L-S65T-GFP (sequence disclosed in WO 97/11094) by polymerase chain reaction (PCR). The PCR reactions are performed according to standard protocols with the following primers:

5'ERK1:

20 3'ERK1:

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5'F64L-S65T-GFP:

TTggACACAAgCTTTggACACggCgCgCCATgAgTAAAggAgAAGAACTTTTC (SEQ ID NO:1)

3'F64L-S65T-GFP:

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gTCATCTTCTCgAgTCTTACTCCTgAggTTTgTATAgTTCATCCATgCCATgT (SEQ ID NO:2)

To generate the mERK1-F64L-S65T-GFP (SEQ ID NO:56 & 57) fusion the ERK1 amplification product is digested with HindIII+AscI and the F64L-S65T-GFP product with AscI+XhoI. To generate the F64L-S65T-GFP-mERK1 fusion the ERK1 amplification product is then digested with HindIII+Bsu36I and the F64L-S65T-GFP product with Bsu36I+XhoI. The two pairs of digested PCR products are subsequently ligated with a HindIII+XhoI digested plasmid (pZeoSV® mammalian expression vector, Invitrogen, San Diego, CA, USA). The resulting fusion constructs are under control of the SV40 promoter.

b) The human Erk1 gene (GenBank Accession number: X60188) was amplified using PCR according to standard protocols with primers Erk1-top (SEQ ID NO:9) and Erk1-bottom/+stop (SEQ ID NO:10). The PCR product was digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with EcoR1 and BamH1. This produces an EGFP-Erk1 fusion (SEQ ID NO:38 &39) under the control of a CMV promoter.

The plamid containing the EGFP-Erk1 fusion was transfected into HEK293 cells employing the FUGENE transfection reagent (Boehringer Mannheim). Prior to experiments the cells were grown to 80%-90% confluency 8 well chambers in DMEM with 10% FCS. The cells were washed in plain HAM F-12 medium (without FCS), and then incubated for 30-60 minutes in plain HAM F-12 (without FCS) with 100 micromolar PD98059, an inhibitor of MEK1, a kinase which activates Erk1; this step effectively empties the nucleus of EGFP-Erk1. Just before starting the experiment, the HAM F-12 was replaced with Hepes buffer following a wash with Hepes buffer. This removes the PD98059 inhibitor; if blocking of MEK1 is still wanted (e.g. in control experiments), the inhibitor is included in the Hepes buffer.

The experimental setup of the microscope was as described in example 1.

60 images were collected with 10 seconds between each, and with the test compound added after image number 10.

Addition of EGF (1-100 nM) caused within minutes a redistribution of EGFP-Erk1 from the cytoplasm into the nucleus (Fig. 9a,b).

The response was quantitated as described below and a dose-dependent relationship between EGF concentration and nuclear translocation of EGFP-Erk1 was found (Fig. 9c,d). Redistribution of GFP fluorescence is expressed in this example as the change in the ratio value between areas in nuclear versus cytoplasmic compartments of the cell. Each time profile is the average of nuclear to cytoplasmic ratios from six cells in each treatment.

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EXAMPLE 4

Probes for detection of Erk2 redistribution.

Useful for monitoring signalling pathways involving MAPK, e.g. to identify compounds which modulate the activity of the pathway in living cells.

- 15 Erk2, a serine/threonine protein kinase, is closely related to Erk1 but not identical; it is a component of a signalling pathway that is activated by e.g. many growth factors.
 - a) The rat Erk2 gene (GenBank Accession number: M64300) was amplified using PCR according to standard protocols with primers Erk2-top (SEQ ID NO:11) and Erk2-bottom/+stop (SEQ ID NO:13) The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produces an EGFP-Erk2 fusion (SEQ ID NO:40 &41) under the control of a CMV promoter.
 - b) The rat Erk2 gene (GenBank Accession number: M64300) was amplified using PCR according to standard protocols with primers (SEQ ID NO:11) Erk2-top and Erk2-bottom/-stop (SEQ ID NO:12). The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank

Accession number U55762) digested with Xho1 and BamH1. This produces an Erk2-EGFP fusion (SEQ ID NO:58 &59) under the control of a CMV promoter.

The resulting plasmids were transfected into CHO cells and BHK cells. The cells were grown under standard conditions. Prior to experiments, the cells were starved in medium without serum for 48-72 hours. This led to a predominantly cytoplasmic localisation of both probes, especially in BHK cells. 10% fetal calf serum was added to the cells and the fluorescence of the cells was recorded as explained in example 3. Addition of serum caused the probes to redistribute into the nucleus within minutes of addition of serum.

10 EXAMPLE 5

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Probes for detection of Smad2 redistribution.

Useful for monitoring signalling pathways activated by some members of the transforming growth factor-beta family, e.g. to identify compounds which modulate the activity of the pathway in living cells.

- Smad 2, a signal transducer, is a component of a signalling pathway that is induced by some members of the TGFbeta family of cytokines.
 - a) The human Smad2 gene (GenBank Accession number: AF027964) was amplified using PCR according to standard protocols with primers Smad2-top (SEQ ID NO:24) and Smad2-bottom/+stop (SEQ ID NO:26). The PCR product was digested with restriction enzymes EcoR1 and Acc651, and ligated into pEGFP-C1 (Clontech; Palo Alto; GenBank Accession number U55763) digested with EcoR1 and Acc651. This produces an EGFP-Smad2 fusion (SEQ ID NO:50&51) under the control of a CMV promoter.
 - b) The human Smad2 gene (GenBank Accession number: AF027964) was amplified using PCR according to standard protocols with primers Smad2-top (SEQ ID NO:24) and Smad2-bottom/-stop (SEQ ID NO:25). The PCR product was digested with restriction enzymes EcoR1 and Acc651, and ligated into pEGFP-N1 (Clontech, Palo Alto;

GenBank Accession number U55762) digested with EcoR1 and Acc65I. This produces a Smad2-EGFP fusion (SEQ ID NO:74 &75) under the control of a CMV promoter.

The plasmid containing the EGFP-Smad2 fusion was transfected into HEK293 cells, where it showed a cytoplasmic distribution. Prior to experiments the cells were grown in 8 well Nunc chambers in DMEM with 10% FCS to 80% confluence and starved overnight in HAM F-12 medium without FCS.

For experiments, the HAM F-12 medium was replaced with Hepes buffer pH 7.2.

The experimental setup of the microscope was as described in example 1.

90 images were collected with 10 seconds between each, and with the test compound added after image number 5.

After serum starvation of cells, each nucleus contains less GFP fluorescence than the surrounding cytoplasm (Fig. 10a). Addition of TGFbeta caused within minutes a redistribution of EGFP-Smad2 from the cytoplasma into the nucleus (Fig. 10b).

The redistribution of fluorescence within the treated cells was quantified simply as the fractional increase in nuclear fluorescence normalised to the starting value of GFP fluorescence in the nucleus of each unstimulated cell.

EXAMPLE 6

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Probe for detection of VASP redistribution.

Useful for monitoring signalling pathways involving rearrangement of cytoskeletal elements, e.g. to identify compounds which modulate the activity of the pathway in living cells.

VASP, a phosphoprotein, is a component of cytoskeletal structures, which redistributes in response to signals that affect focal adhesions.

The human VASP gene (GenBank Accession number: Z46389) was amplified using PCR according to standard protocols with primers VASP-top (SEQ ID NO:94) and VASP-bottom/+stop (SEQ ID NO:95). The PCR product was digested with restriction enzymes Hind3 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Hind3and BamH1. This produces an EGFP-VASP fusion (SEQ ID NO:124 &125) under the control of a CMV promoter.

The resulting plasmid was transfected into CHO cells expressing the human insulin receptor using the calcium-phosphate transfection method. Prior to experiments, cells were grown in 8 well Nunc chambers and starved overnight in medium without FCS.

Experiments are performed in a microscope setup as described in example 1.

10% FCS was added to the cells and images were collected. The EGFP-VASP fusion was redistributed from a somewhat even distribution near the periphery into more localised structures, identified as focal adhesion points (Fig. 11).

A large number of further GFP fusions have been made or are in the process of being made, as apparent from the following Examples 7-22 which also suggest suitable host cells and substances for activation of the cellular signalling pathways to be monitored and analyzed.

EXAMPLE 7

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20 Probe for detection of actin redistribution.

Useful for monitoring signalling pathways involving rearrangement or formation of actin filaments, e.g. to identify compounds which modulate the activity of pathways leading to cytoskeletal rearrangements in living cells.

Actin is a component of cytoskeletal structures, which redistributes in response to very many cellular signals.

The actin binding domain of the human alpha-actinin gene (GenBank Accession number:

X15804) was amplified using PCR according to standard protocols with primers ABD-top (SEQ ID NO:90) and ABD-bottom/-stop (SEQ ID NO:91). The PCR product was digested with restriction enzymes Hind3 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Hind3 and BamH1. This produced an actin-binding-domain-EGFP fusion (SEQ ID NO:128 &129) under the control of a CMV promoter.

The resulting plasmid was transfected into CHO cells expressing the human insulin receptor. Cells were stimulated with insulin that caused the actin binding domain-EGFP probe to become redistributed into morphologically distinct membrane-associated structures.

EXAMPLE 8

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Probes for detection of p38 redistribution.

Useful for monitoring signalling pathways responding to various cellular stress situations,
e.g. to identify compounds which modulate the activity of the pathway in living cells, or as a counterscreen.

p38, a serine/threonine protein kinase, is a component of a stress-induced signalling pathway which is activated by many types of cellular stress, e.g. TNFalpha, anisomycin, UV and mitomycin C.

- a) The human p38 gene (GenBank Accession number: L35253) was amplified using PCR according to standard protocols with primers p38-top (SEQ ID NO:14) and p38-bottom/+stop (SEQ ID NO: 16). The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produced an EGFP-p38 fusion (SEQ ID NO:46 & 47) under the control of a CMV promoter.
 - b) The human p38 gene (GenBank Accession number: L35253) was amplified using PCR according to standard protocols with primers p38-top (SEQ ID NO:13) and p38-

bottom/-stop (SEQ ID NO:15). The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and BamH1. This produced a p38-EGFP fusion (SEQ ID NO:64 & 65) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. HEK293, in which the EGFP-p38 probe and/or the p38-EGFP probe should change its cellular distribution from predominantly cytoplasmic to nuclear within minutes in response to activation of the signalling pathway with e.g. anisomycin.

EXAMPLE 9

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Probes for detection of Jnk1 redistribution.

Useful for monitoring signalling pathways responding to various cellular stress situations, e.g. to identify compounds which modulate the activity of the pathway in living cells, or as a counterscreen.

- Jnk1, a serine/threonine protein kinase, is a component of a stress-induced signalling pathway different from the p38 described above, though it also is activated by many types of cellular stress, e.g. TNFalpha, anisomycin and UV.
 - a) The human Jnk1 gene (GenBank Accession number: L26318) was amplified using PCR according to standard protocols with primers Jnk-top (SEQ ID NO:17) and Jnk-bottom/+stop (SEQ ID NO:19). The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produced an EGFP-Jnk1 fusion (SEQ ID NO:44 &45) under the control of a CMV promoter.
- b) The human Jnk1 gene (GenBank Accession number: L26318) was amplified using PCR according to standard protocols with primers Jnk-top (SEQ ID NO:17) and Jnk-bottom/-stop (SEQ ID NO:18). The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank

Accession number U55762) digested with Xho1 and BamH1. This produced a Jnk1-EGFP fusion (SEQ ID NO:62 &63) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. HEK293, in which the EGFP-Jnk1 probe and/or the Jnk1-EGFP probe should change its cellular distribution from predominantly cytoplasmic to nuclear in response to activation of the signalling pathway with e.g. anisomycin.

EXAMPLE 10

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Probes for detection of PKG redistribution.

Useful for monitoring signalling pathways involving changes in cyclic GMP levels, e.g. to identify compounds which modulate the activity of the pathway in living cells.

PGK, a cGMP-dependent serine/threonine protein kinase, mediates the guanylyl-cyclase/cGMP signal.

- a) The human PKG gene (GenBank Accession number: Y07512) is amplified using PCR according to standard protocols with primers PKG-top (SEQ ID NO:81) and PKG-bottom/+stop (SEQ ID NO:83). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produces an EGFP-PKG fusion (SEQ ID NO:134 &135) under the control of a CMV promoter.
- b) The human PKG gene (GenBank Accession number: Y07512) is amplified using PCR according to standard protocols with primers PKG-top (SEQ ID NO:81) and PKG-bottom/-stop (SEQ ID NO: 82). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and BamH1. This produces a PKG-EGFP fusion (SEQ ID NO:136 &137) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. A10, in which the EGFP-PKG probe and/or the PKG-EGFP probe should change its cellular distribution

from cytoplasmic to one associated with cytoskeletal elements within minutes in response to treatment with agents which raise nitric oxide (NO) levels.

EXAMPLE 11

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5 Probes for detection of IkappaB kinase redistribution.

Useful for monitoring signalling pathways leading to NFkappaB activation, e.g. to identify compounds which modulate the activity of the pathway in living cells.

IkappaB kinase, a serine/threonine kinase, is a component of a signalling pathway which is activated by a variety of inducers including cytokines, lymphokines, growth factors and stress.

- a) The alpha subunit of the human IkappaB kinase gene (GenBank Accession number: AF009225) is amplified using PCR according to standard protocols with primers IKK-top (SEQ ID NO:96) and IKK-bottom/+stop (SEQ ID NO:98). The PCR product is digested with restriction enzymes EcoR1 and Acc651, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with EcoR1and Acc651. This produces an EGFP-IkappaB-kinase fusion (SEQ ID NO:120 &121) under the control of a CMV promoter.
- b) The alpha subunit of the human IkappaB kinase gene (GenBank Accession number:

 AF009225) is amplified using PCR according to standard protocols with primers IKK
 top (SEQ ID NO:96) and IKK-bottom/-stop (SEQ ID NO:97). The PCR product is

 digested with restriction enzymes EcoR1 and Acc65I, and ligated into pEGFP-N1

 (Clontech, Palo Alto; GenBank Accession number U55762) digested with EcoR1 and

 Acc65I. This produces an IkappaB-kinase-EGFP fusion (SEQ ID NO:122 &123) under

 the control of a CMV promoter.
- The resulting plasmids are transfected into a suitable cell line, e.g. Jurkat, in which the EGFP-lkappaB-kinase probe and/or the lkappaB-kinase-EGFP probe should achieve a more cytoplasmic distribution within seconds following stimulation with e.g. TNFalpha.

EXAMPLE 12

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Probes for detection of CDK2 redistribution.

Useful for monitoring signalling pathways of the cell cycle, e.g. to identify compounds that modulate the activity of the pathway in living cells.

CDK2, a cyclin-dependent serine/threonine kinase, is a component of the signalling system that regulates the cell cycle.

- a) The human CDK2 gene (GenBank Accession number: X61622) is amplified using PCR according to standard protocols with primers CDK2-top (SEQ ID NO:102) and CDK2-top (SEQ ID NO: 104). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produces an EGFP-CDK2 fusion (SEQ ID NO:114 &115) under the control of a CMV promoter.
- b) The human CDK2 gene (GenBank Accession number: X61622) is amplified using PCR according to standard protocols with primers CDK2-top (SEQ ID NO:102) and CDK2-bottom/-stop (SEQ ID NO:103). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and BamH1. This produces a CDK2-EGFP fusion (SEQ ID NO:112 &113) under the control of a CMV promoter.
- The resulting plasmids are transfected into a suitable cell line, e.g. HEK293 in which the EGFP-CDK2 probe and/or the CDK2-EGFP probe should change its cellular distribution from cytoplasmic in contact-inhibited cells, to nuclear location in response to activation with a number of growth factors, e.g. IGF.

25 EXAMPLE 13

Probes for detection of Grk5 redistribution.

Useful for monitoring signalling pathways involving desensitisation of G-protein coupled receptors, e.g. to identify compounds which modulate the activity of the pathway in living cells.

Grk5, a G-protein coupled receptor kinase, is a component of signalling pathways involving membrane bound G-protein coupled receptors.

- a) The human Grk5 gene (GenBank Accession number: L15388) is amplified using PCR according to standard protocols with primers Grk5-top (SEQ ID NO:27) and Grk5-bottom/+stop (SEQ ID NO:29). The PCR product is digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with EcoR1 and BamH1. This produces an EGFP-Grk5 fusion (SEQ ID NO:42 &43) under the control of a CMV promoter.
- b) The human Grk5 gene (GenBank Accession number: L15388) is amplified using PCR according to standard protocols with primers Grk5-top (SEQ ID NO:27) and Grk5-bottom/-stop (SEQ ID NO:28). The PCR product is digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with EcoR1 and BamH1. This produces a Grk5-EGFP fusion (SEQ ID NO:60 &61) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. HEK293 expressing a rat dopamine D1A receptor, in which the EGFP-Grk5 probe and/or the Grk5-EGFP probe should change its cellular distribution from predominantly cytoplasmic to peripheral in response to activation of the signalling pathway with e.g. dopamine.

EXAMPLE 14

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25 Probes for detection of Zap70 redistribution.

Useful for monitoring signalling pathways involving the T cell receptor, e.g. to identify compounds which modulate the activity of the pathway in living cells.

Zap70, a tyrosine kinase, is a component of a signalling pathway which is active in e.g. T-cell differentiation.

- a) The human Zap70 gene (GenBank Accession number: L05148) is amplified using PCR according to standard protocols with primers Zap70-top (SEQ ID NO:105) and Zap70-bottom/+stop (SEQ ID NO:107). The PCR product is digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-C1 (GenBank Accession number U55763) digested with EcoR1 and BamH1. This produces an EGFP-Zap70 fusion (SEQ ID NO:108 & 109) under the control of a CMV promoter.
- b) The human Zap70 gene (GenBank Accession number: L05148) is amplified using PCR according to standard protocols with primers Zap70-top (SEQ ID NO:105) and Zap70-bottom/-stop (SEQ ID NO:106). The PCR product is digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with EcoR1 and BamH1. This produces a Zap70-EGFP fusion (SEQ ID NO:110 &111) under the control of a CMV promoter.
- The resulting plasmids are transfected into a suitable cell line, e.g. Jurkat, in which the EGFP-Zap70 probe and/or the Zap70-EGFP probe should change its cellular distribution from cytoplasmic to membrane-associated within seconds in response to activation of the T cell receptor signalling pathway with e.g. antibodies to CD3epsilon.

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Probes for detection of p85 redistribution.

Useful for monitoring signalling pathways involving PI-3 kinase, e.g. to identify compounds which modulate the activity of the pathway in living cells.

p85alpha is the regulatory subunit of PI3-kinase which is a component of many pathways involving membrane-bound tyrosine kinase receptors and G-protein-coupled receptors.

a) The human p85alpha gene (GenBank Accession number: M61906) was amplified using PCR according to standard protocols with primers p85-top-C (SEQ ID NO:22) and p85-

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bottom/+stop (SEQ ID NO:23). The PCR product was digested with restriction enzymes Bgl2 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Bgl2 and BamH1. This produced an EGFP-p85alpha fusion (SEQ ID NO:48 &49) under the control of a CMV promoter.

b) The human p85alpha gene (GenBank Accession number: M61906) was amplified using PCR according to standard protocols with primers p85-top-N (SEQ ID NO:20) and p85-bottom/-stop (SEQ ID NO:21). The PCR product was digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with EcoR1 and BamH1. This produced a p85alpha-EGFP fusion (SEQ ID NO:66 &67) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. CHO expressing the human insulin receptor, in which the EGFP-p85 probe and/or the p85-EGFP probe may change its cellular distribution from cytoplasmic to membrane-associated within minutes in response to activation of the receptor with insulin.

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EXAMPLE 16

Probes for detection of protein-tyrosine phosphatase redistribution.

Useful for monitoring signalling pathways involving tyrosine kinases, e.g. to identify compounds which modulate the activity of the pathway in living cells.

- 20 Protein-tyrosine phosphataselC, a tyrosine-specific phosphatase, is an inhibitory component in signalling pathways involving e.g. some growth factors.
 - a) The human protein-tyrosine phosphatase 1C gene (GenBank Accession number: X62055) is amplified using PCR according to standard protocols with primers PTP-top (SEQ ID NO:99) and PTP-bottom/+stop (SEQ ID NO:101). The PCR product is digested with restriction enzymes Xho1 and EcoR1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and EcoR1. This produces an EGFP-PTP fusion (SEQ ID NO:116 & 117) under the control

of a CMV promoter.

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b) The human protein-tyrosine phosphatase 1C gene (GenBank Accession number: X62055) is amplified using PCR according to standard protocols with primers PTP-top (SEQ ID NO:99) and PTP-bottom/-stop (SEQ ID NO:100). The PCR product is digested with restriction enzymes Xho1 and EcoR1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and EcoR1. This produces a PTP-EGFP fusion (SEQ ID NO:118 & 119) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. MCF-7 in which the EGFP-PTP probe and/or the PTP-EGFP probe should change its cellular distribution from cytoplasm to the plasma membrane within minutes in response to activation of the growth inhibitory signalling pathway with e.g. somatostatin.

EXAMPLE 17

15 Probes for detection of Smad4 redistribution.

Useful for monitoring signalling pathways involving most members of the transforming growth factor-beta family, e.g. to identify compounds which modulate the activity of the pathway in living cells.

Smad4, a signal transducer, is a common component of signalling pathways induced by various members of the TGFbeta family of cytokines.

- a) The human Smad4 gene (GenBank Accession number: U44378) was amplified using PCR according to standard protocols with primers Smad4-top and Smad4-bottom/+stop (SEQ ID NO:35). The PCR product was digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with EcoR1 and BamH1. This produce an EGFP-Smad4 fusion (SEQ ID NO:52 & 53) under the control of a CMV promoter.
- b) The human Smad4 gene (GenBank Accession number: U44378) was amplified using

PCR according to standard protocols with primers Smad4-top (SEQ ID NO:33) and Smad4-bottom/-stop (SEQ ID NO:34). The PCR product was digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with EcoR1 and BamH1. This produced a Smad4-EGFP fusion (SEQ ID NO:76 & 77) under the control of a CMV promoter.

The resulting plasmids are transfected into a cell line, e.g. HEK293 in which the EGFP-Smad4 probe and/or the Smad4-EGFP probe should change its cellular distribution within minutes from cytoplasmic to nuclear in response to activation of the signalling pathway with e.g. TGFbeta.

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EXAMPLE 18

Probes for detection of Stat5 redistribution.

Useful for monitoring signalling pathways involving the activation of tyrosine kinases of the Jak family, e.g. to identify compounds that modulate the activity of the pathway in living cells.

Stat5, signal transducer and activator of transcription, is a component of signalling pathways that are induced by e.g. many cytokines and growth factors.

- a) The human Stat5 gene (GenBank Accession number: L41142) was amplified using PCR according to standard protocols with primers Stat5-top (SEQ ID NO:30) and Stat5-bottom/+stop (SEQ ID NO:32). The PCR product was digested with restriction enzymes Bgl2 and Acc65I, and ligated into pEGFP-C1 (Clontech; Palo Alto; GenBank Accession number U55763) digested with Bgl2 and Acc65I. This produced an EGFP-Stat5 fusion (SEQ ID NO:54 & 55) under the control of a CMV promoter.
- b) The human Stat5 gene (GenBank Accession number: L41142) was amplified using PCR according to standard protocols with primers Stat5-top (SEQ ID NO:30) and Stat5-bottom/-stop (SEQ ID NO:331). The PCR product was digested with restriction

enzymes Bgl2 and Acc651, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Bgl2 and Acc65I. This produced a Stat5-EGFP fusion (SEQ ID NO:78 & 79) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. MIN6 in which the EGFP-Stat5 probe and/or the Stat5-EGFP probe should change its cellular distribution from cytoplasmic to nuclear within minutes in response to activation signalling pathway with e.g. prolactin.

EXAMPLE 19

10 Probes for detection of NFAT redistribution.

Useful for monitoring signalling pathways involving activation of NFAT, e.g. to identify compounds which modulate the activity of the pathway in living cells.

NFAT, an activator of transcription, is a component of signalling pathways involved in e.g. immune responses.

- a) The human NFAT1 gene (GenBank Accession number: U43342) is amplified using PCR according to standard protocols with primers NFAT-top (SEQ ID NO:84) and NFAT-bottom/+stop (SEQ ID NO:86). The PCR product is digested with restriction enzymes Xho1 and EcoR1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and EcoR1. This produces an EGFP-NFAT fusion (SEQ ID NO:130 & 131) under the control of a CMV promoter.
 - b) The human NFAT gene (GenBank Accession number: U43342) is amplified using PCR according to standard protocols with primers NFAT-top (SEQ ID NO:84) and NFAT-bottom/-stop (SEQ ID NO:85). The PCR product is digested with restriction enzymes Xho1 and EcoR1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and EcoR1. This produces an NFAT-EGFP fusion (SEQ ID NO:132 & 133) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. Jurkat, in which the

EGFP-NFAT probe and/or the NFAT-EGFP probe should change its cellular distribution from cytoplasmic to nuclear within minutes in response to activation of the signalling pathway with e.g. antibodies to CD3epsilon.

EXAMPLE 20

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Probes for detection of NFkappaB redistribution.

Useful for monitoring signalling pathways leading to activation of NFkappaB, e.g. to identify compounds which modulate the activity of the pathway in living cells.

NFkappaB, an activator of transcription, is a component of signalling pathways that are responsive to a varity of inducers including cytokines, lymphokines, and some immunosuppressive agents.

- a) The human NFkappaB p65 subunit gene (GenBank Accession number: M62399) is amplified using PCR according to standard protocols with primers NFkappaB-top (SEQ ID NO:87) and NFkappaB-bottom/+stop (SEQ ID NO:89). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produces an EGFP-NFkappaB fusion (SEQ ID NO:142 & 143) under the control of a CMV promoter.
- b) The human NFkappaB p65 subunit gene (GenBank Accession number: M62399) is
 amplified using PCR according to standard protocols with primers NFkappaB-top (SEQ ID NO:87) and NFkappaB-bottom/-stop (SEQ ID NO:88). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and BamH1. This produces an NFkappaB-EGFP fusion (SEQ ID NO:140 & 141) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. Jurkat, in which the EGFP-NFkappaB probe and/or the NFkappaB-EGFP probe should change its cellular

distribution from cytoplasmic to nuclear in response to activation of the signalling pathway with e.g. TNFalpha.

EXAMPLE 21

5 Probe for detection of RhoA redistribution.

Useful for monitoring signalling pathways involving RhoA, e.g. to identify compounds which modulate the activity of the pathway in living cells.

RhoA, a small GTPase, is a component of many signalling pathways, e.g. LPA induced cytoskeletal rearrangements.

The human RhoA gene (GenBank Accession number: L25080) was amplified using PCR according to standard protocols with primers RhoA-top (SEQ ID NO:92) and RhoA-bottom/+stop (SEQ ID NO:93). The PCR product was digested with restriction enzymes Hind3 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Hind3and BamH1. This produced an EGFP-RhoA fusion (SEQ ID NO:126 &127) under the control of a CMV promoter.

The resulting plasmid is transfected into a suitable cell line, e.g. Swiss3T3, in which the EGFP-RhoA probe should change its cellular distribution from a reasonably homogenous to a peripheral distribution within minutes of activation of the signalling pathway with e.g. LPA.

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EXAMPLE 22

Probes for detection of PKB redistribution.

Useful for monitoring signalling pathways involving PKB e.g. to identify compounds which modulate the activity of the pathway in living cells.

25 PKB, a serine/threonine kinase, is a component in various signalling pathways, many of

which are activated by growth factors.

- a) The human PKB gene (GenBank Accession number: M63167) is amplified using PCR according to standard protocols with primers PKB-top (SEQ ID NO:36) and PKB-bottom/+stop (SEQ ID NO:80). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produces an EGFP-PKB fusion (SEQ ID NO:138 & 139) under the control of a CMV promoter.
- b) The human PKB gene (GenBank Accession number: M63167) was amplified using
 PCR according to standard protocols with primers PKB-top (SEQ ID NO:36) and PKBbottom/-stop (SEQ ID NO:37). The PCR product was digested with restriction
 enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank
 Accession number U55762) digested with Xho1 and BamH1. This produced a PKBEGFP fusion (SEQ ID NO:70 &71) under the control of a CMV promoter.
- The resulting plasmids are transfected into a suitable cell line, e.g. CHO expressing the human insulin receptor, in which the EGFP-PKB probe and/or the PKB-EGFP probe cycles between cytoplasmic and membrane locations during the activation-deactivation process following addition of insulin. The transition should be apparent within minutes.

20 EXAMPLE 23

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Measurement of the real-time redistribution of protein kinase C α isoform-GFP fusion (PKC α -GFP) in response to carbamylcholine stimulation of the muscarinic M1 receptor; 96 parallel redistribution measurements in microtiter plates.

BHK cells were stably expressing a recombinant human muscarinic typ 1 receptor, under the selection with 500 μg/ml Methotrexate, and also a PKCα-GFP construct (KaA 048), under the selection of 500 nM Zeocin. The cells were grown in 96-well plates (Packard ViewPlate, black with transparent bottom), washed and preincubated in a Hank's Buffered

Salt solution (HBSS) without phenol red, with 20 mM HEPES and 5.5 mM glucose.

The plate was measured in a FLIPRTM (Fluorescence Imaging Plate Reader) from Molecular Devices. The 488 nm emission line from an argon ion laser, run at between 0.4 and 0.8 W output, was used to excite fluorescence form the GFP. Emission wavelengths were collected through a 510 to 565 nm band pass filter.

The cells were challenged with three doses of carbamylcholine, an M1 receptor agonist known from previous studies to give a microscopically detectable redistribution of the PKCα-GFP construct [(Almholt *et al.* 1997)]. Measurements were made every 10 seconds for 5 minutes. After data handling including normalisation of baseline fluorescence for the different wells, background subtraction and averaging the 6 wells used for each concentration the data presented in figure 14 were obtained. It can clearly be seen (Fig 14) that carbamylcholine gave a time- and dose-dependent, and transient, decrease in fluorescence very similar to the time- and dose-dependent profile seen in microscopic fluorescence measurements [(see Almholt *et al.* 1997)]. This experiment was repeated twice on the same batch of cells with similar results.

EXAMPLE 24

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Measurement of the real-time redistribution of cyclic-AMP dependent protein kinase catalytic subuit-GFP fusion (C-GFP^{LT}) in response to forskolin stimulation of the adenylate cyclase; 96 parallel redistribution measurements in microtiter plates.

CHO cells were stably transfected with hybrid DNA for the PKA catalytic subunit-F64L+S65T GFP (C-GFP^{LT}) fusion protein, and were typically under continuous selection with 1000 µg/ml zeocin (Invitrogen). The cells were grown without selection for 2 days in 96-well plates (Packard ViewPlate, black with transparent bottom), washed and preincubated in a Hank's Buffered Salt solution (HBSS) without phenol red, with 20 mM HEPES and 5.5 mM glucose.

The plate was measured in a FLIPRTM (Fluorescence Imaging Plate Reader) from Molecular Devices. The 488 nm emission line from an argon ion laser, run at between 0.4

and 0.8 W output, was used to excite fluorescence from the GFP. Emission wavelengths were collected through a 510 to 565 nm band pass filter.

The cells were challenged with three doses of forskolin (Fig 15), an adenylate cyclase agonist known from previous studies to give a microscopically detectable redistribution of the C-GFP^{LT} construct [(Almholt *et al.* 1998)]. Measurements were made every 10 seconds for over 6 minutes from the point of addition of forskolin. After data handling including normalisation of baseline fluorescence for the different wells, background subtraction and averaging the 6 wells used for each concentration the data presented below were obtained. It can clearly be seen in figure 15 that forskolin gave a time- and dose-dependent decrease in fluorescence very similar to the time- and dose-dependent profile seen in microscopic fluorescence measurements [(see Almholt *et al.* 1998)]. This experiment was repeated twice on the same batch of cells with similar results.

EXAMPLE 25

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Measurement of the redistribution response of cyclic-AMP dependent protein kinase catalytic subuit-GFP fusion (C-GFP^{LT}) after forskolin stimulation of the adenylate cyclase; measurement of the change in total fluorescence upon permeabilisation of agonist-treated cells.

CHO cells were stably transfected with hybrid DNA for the PKA catalytic subunit- F64L+S65T GFP (C-GFP^{LT}) fusion protein, and were typically under continuous selection with 1000 μ g/ml zeocin (Invitrogen). For the experiments reported here, cells were grown without selection to 90% confluence in 8-well tissue culture-treated Lab-Tek® chambered coverglass units (chambers, obtained from Nunc, Inc. Illinois, USA). Immediately prior to the experiment growth medium was washed from the cells and replaced with 200 μ l HEPES buffer per well.

For the results reported here, chambers were measured using a cooled CCD camera (KAF1400 chip, Photometrics Ltd., USA) attached to an inverted microscope (Diaphot 300, Nikon, Japan) equipped with a x40 oil-immersion Fluar lens, NA 1.4. Cells were

illuminated with 450-490 nm light from a 50 W HBO lamp, and emitted light collected between 510-560 nm.

The cells were challenged with four doses of forskolin, an adenylate cyclase agonist known from previous studies to give a microscopically detectable redistribution of the C-GFP^{LT} construct [(Almholt *et al.* 1998)]. Images were collected at 10-second intervals for a period of 10 minutes for each treatment. Six minutes after the addition of forskolin or buffer control, Triton-X100 was added to a final concentration of 0.1%. The detergent releases freely mobile C-GFP^{LT} from the cells. The change in fluorescence resulting from this loss was measured after 1 minute of equilibration. After data handling including background subtraction and normalisation to pre-detergent values, the data presented in figure 16 were obtained. Permeabilisation caused decreases in fluorescence, the magnitude of which were dependent on the forskolin treatments. The dose-dependent profile for forskolin activation of the cAMP system as revealed by this method was very similar to that registered by other methods (see Almholt *et al.* 1998). This experiment was repeated twice on the same batch of cells with similar results.

EXAMPLE 26

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Probe for detection of PKCbeta2 redistribution.

Useful for monitoring signalling pathways involving protein kinase C, e.g. for identifying compounds which modulate the activity of the pathway in living cells.

PKCbeta2, a serine/threonine protein kinase, is closely related to PKCalpha but not identical; it is a component of a signalling pathway that is activated by elevation of intracellular calcium concomitant with an increase in diacylglycerol species.

a) The human PKCbeta2 gene (GenBank Accession number: X07109) was amplified using PCR according to standard protocols with primers PKCbeta2-top (SEQ ID NO:162) and PKCbeta2-bottom (SEQ ID NO:163). The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and BamH1. This produces a PKCbeta2-

EGFP fusion (SEQ ID NO:146 & 147) under the control of a CMV promoter.

The resulting plasmids are transfected into BHK cells transfected with a human muscarinic acetylcholine receptor type M1. The cells are grown under standard conditions. The fluorescence of the cells is recorded as explained in example 3. Addition of $1\mu M$ - $100\mu M$ carbachol causes a transient redistribution of fluorescence within the cells whereby it changes from a cytosolic location to the plasma membrane.

EXAMPLE 27

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Probes for detection of PDE4D redistribution.

Useful for monitoring signalling pathways involving Protein Kinase A, e.g. to identify compounds which modulate the activity of the pathway in living cells.

PDE4D3, PDE4D4 and PDE4D5 are closely related splicing variants of PDE4D, a cAMP dependent phosphodiesterase. They are components of signalling pathways which involves cAMP.

The human PDE4D3, PDE4D4 and PDE4D5 genes (GenBank Accession numbers: L20970, L20969 and AF012073) are amplified using PCR according to standard protocols with the common bottom primer PDE4D-bottom (SEQ ID NO:159) and PDE4D3-top (SEQ ID NO:156), PDE4D4-top (SEQ ID NO:157) and PDE4D5-top respectively (SEQ ID NO:158) The PCR products are digested with restriction enzymes Hind3 and EcoR1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Hind3 and EcoR1. This produces a PDE4D3-EGFP fusion (SEQ ID NO:154 & 155), a PDE4D4-EGFP fusion (SEQ ID NO:150 & 151) and a PDE4D5-EGFP fusion (SEQ ID NO:148 & 149), all three under the control of a CMV promoter.

The resulting plasmids are transfected into MVLEC cells. The cells are grown under standard conditions. The fluorescence of the cells is recorded as explained in example 3. Addition of test compounds may cause a redistribution of fluorescence within the cells from an organised cytosolic distribution to a more random one.

EXAMPLE 28

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Probes for detection of PDE5 redistribution.

Useful for monitoring signalling pathways involving Protein Kinase G, e.g. to identify compounds which modulate the activity of the pathway in living cells.

PDE5 is a cGMP specific phosphodiesterase. It is a component of a signalling pathway which is activated by e.g. nitric oxide.

a) The human PDE5 gene (GenBank Accession numbers: AJ004865) is amplified using PCR according to standard protocols with primers PDE5-top (SEQ ID NO:160) and PDE5-bottom (SEQ ID NO:161). The PCR product is digested with restriction enzymes EcoR1 and Acc651, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with EcoR1 and Acc651. This produces a PDE5-EGFP fusion (SEQ ID NO 144 & 145) under the control of a CMV promoter.

The resulting plasmids are transfected into e.g. A10 cells. The cells are grown under standard conditions. The fluorescence of the cells is recorded as explained in example 3. Addition of test compounds may cause a redistribution of fluorescence within the cells from an organized cytosolic distribution to a more random one.

EXAMPLE 29

20 Probe for detection of Ikappa-kinase redistribution.

The human IKKbeta (GenBank Acc. No. AF031416) is amplified using PCR according to standard protocols with primers IKKbeta-top (SEQ ID NO:164) and IKKbeta-bottom (SEQ ID NO:165). The PCR product is digested with restriction enzymes Hind3 and Acc651, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Hind3 and Acc651. This produces a IKKbeta-EGFP fusion (SEQ ID NO 152 & 153) under the control of a CMV promoter.

EXAMPLE 30

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Construction of catalytically inactive Erk1 probes.

A catalytically inactive probe has the advantage that it interferes less with the normal physiology of the cell while retaining its ability to report on activation of a cellular signalling pathway by redistribution.

The Erk1 probes described above in Example 3 were subjected to site specific mutagenesis which specifically replaced the lysine at amino acid residue number 71 in the native Erk1 sequence with arginine. This mutation is known to inactivate the catalytic activity of Erk1. The redistribution patterns of the inactive Erk1 probes were identical to the original Erk1 probes, i.e. they reported on activation of the pathway by redistributing from the cytoplasm into the nucleus. The establishment of stable cell lines expressing the probe was facilitated.

REFERENCES:

- Adams, S.R., Harootunian, A.T., Buechler, Y.J., Taylor, S.S. & Tsien, R.Y. (1991) Nature 349, 694-697.
- Almholt, K., Arkhammar, P.O.G., Thastrup, O., & Tullin, S. (1997) Mol. Biology of the Cell 8: Suppl. S: 72.
 - Almholt, K., Terry, B.R., Skyggebjerg, O., Scudder, K., Tullin, S., Thastrup, O. (1998) [Manuscript], see Appendix II.
- Barak, L.S., Ferguson, S.S.G., Zhang, J. & Caron, M.G. (1997) J. Biol. Chem. **272**:44, 27497-27500.
 - Bastiaens, P.I.H. & Jovin, T.M. (1996) Proc. Natl. Acad. Sci. USA 93, 8407-8412.
 - Beals, C.R., Clipstone, N.A., Ho, S.N. & Crabtree, G.R. (1998) Genes and Development 11:7, 824-834.
 - Blobe, G.C., Stribling, D.S., Fabbro, D., Stabel, S & Hannun, Y.A. (1996) J. Biol. Chem. **271**, 15823-15830.
 - Carey, K.L., Richards, S.A., Lounsbury, K.M. & Macara, I.G. (1996) J. Cell Biol. **133**: 5, 985-996.
 - Chalfie, M., Tu, Y., Euskirchen, G., Ward, W.W. & Prasher, D.C. (1994) Science 263, 802-805.
- 20 Cossette, L.J., Hoglinger, O., Mou, L.J. & Shen, S.H. (1997) Exp. Cell Res. 223, 459-466.
 - DeBernardi, M.A. & Brooker, G. (1996) Proc. Natl. Acad. Sci. USA 93, 4577-4582.
 - Farese, R.V. (1992) Biochem. J. 288, 319-323.
 - Fulop Jr., T., Leblanc, C., Lacombe, G. & Dupuis, G. (1995) FEBS Lett. 375, 69-74.

- Georget, V., Lobaccaro, J.M., Terouanne, B., Mangeat, P., Nicolas, J.C. & Sultan, C. (1997) Mol. Cell. Endocrinol. 129:1, 17-26.
- Godson, C., Masliah, E., Balboa, M.A., Ellisman, M.H. & Insel, P.A. (1996) Biochem. Biophys. Acta 1313, 63-71.
- Guiliano, K.A., DeBiasio, R, Dunlay, R.T., Gough, A., Volosky, J.M., Zock, J., Pavlakis, G.N. & Taylor, D.L. (1997) J. Biomol. Screening 2:4, 249-259.
 - Khalil, R.A., Lajoie, C., Resnick, M.S. & Morgan, K.G. (1992) Am. J. Physiol. **263** (Cell Physiol. 32) C714-C719.
 - Oancea, E., Teruel, M.N., Quest, A.F.G. & Meyer, T. (1998) J. Cell Biol. 140:3, 485-498.
- 10 Sano, M., Kohno, M. & Iwanaga, M. (1995) Brain Res. 688, 213-218.
 - Sakai, N., Sasaki, K., Hasegawa, C., Ohkura, M., Sumioka, K., Shirai, Y. & Saito, N. (1996) Soc. Neuroscience 22, 69P (Abstract).
 - Sakai, N., Sakai, K. Hasegawa, C., Ohkura, M., Sumioka, K., Shirai, Y., & Naoaki, S. (1997) Japanese Journal of Pharmacology 73, 69P (Abstract of a meeting held 22-23 March).
 - Schmidt, D.J., Ikebe, M., Kitamura, K. & Fay, F.S. (1997) FASEB J. 11, 2924 (Abstract).
 - Schroeder, K. & Neagle, B.J. (1996) Biomolecular Screening 1, 75-80.
 - Silverman, L., Campbell, R. & Broach, J.R. (1998) Current Opinion in Chemical Biology 2:3, 397-403.
- 20 Stauffer, T.P., Ahn, S. & Meyer, T. (1998) Current Biol. 8:6, 343-346.

CLAIMS

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- 1. A method for extracting quantitative information relating to an influence on a cellular response, the method comprising recording variation, caused by the influence on mechanically intact or permeabilised living cells, in spatially distributed light emitted from a luminophore, the luminophore being present in the cells and being capable of being redistributed in a manner which is related with the degree of the influence, and/or of being modulated by a component which is capable of being redistributed in a manner which is related to the degree of the influence, resulting in a modulation of the luminescence characteristics of the luminophore, and processing the recorded variation in the luminescence characteristics to provide quantitative information correlating the recorded variation to the degree of the influence on the cellular response.
- 2. A method according to claim 1 for extracting quantitative information relating to an influence on an intracellular pathway involving redistribution of at least one component associated with the pathway, or part thereof, the method comprising recording the result of the influence on mechanically intact or permeabilised living cells, as manifested in spatially distributed light emitted from a luminophore which is present in the cells and which is capable of being redistributed, by modulation of the pathway, in a manner which is related to the redistribution of the at least one component of the intracellular pathway, processing the recorded result to provide quantitative information correlating the change in the measured property of the light to the degree of the influence on the intracellular pathway.
 - 3. A method according to claim 1 or 2, wherein the quantitative information which is indicative of the degree of the cellular response to the influence or the result of the influence on the intracellular pathway is extracted from the recorded variation according to a predetermined calibration based on responses or results, recorded in the same manner, to known degrees of a relevant specific influence.
 - 4. A method according to any of claims 1-3, wherein the influence comprises contact between the mechanically intact or permeabilised living cells and a chemical substance and/or incubation of the mechanically intact or permeabilised living cells with a chemical substance.

- 5. A method according to any of claims 1-4, wherein the influence is a substance whose effect on an intracellular pathway is to be determined.
- 6. A method according to any of claims 1-5, wherein the cells comprise a group of cells contained within a spatial limitation.
- 7. A method according to any of claims 1-5, wherein the cells comprise multiple groups of cells contained within multiple spatial limitations.
 - 8. A method according to any of claims 1-7, wherein the cells comprise multiple groups of cells that are qualitatively the same but are subjected to different influences.
- 9. A method according to any of claims 1-7, wherein the cells comprise multiple groups of cells that are qualitatively different but are subjected to the same influence.
 - 10. A method according to any of claims 1-9, wherein the recording is performed by means of a detector capable of measuring total luminescence in a non-spatially resolved fashion, the recording comprising a time series of measurements of the total luminescence of the cells of one or several of the spatial limitations.
- 11. A method according to claim 10, wherein the signal is measured from individual spatial limitations one at a time, the recording being made in the individual spatial limitation by means of an apparatus to sequentially position each one of the limitations in the field of view of the detector, and repeating the positioning and measuring process until all of the spatial limitations have been measured.
- 12. A method according to claim 11, wherein the detector is a photomultiplier tube (PMT).
 - 13. A method according to any of claims 1-9, wherein more than one of the spatial limitations are measured simultaneously.
- 14. A method according to claim 13, wherein the multiple spatial limitations are measured simultaneously by means of a one- or two-dimensional array detector, whereby the multiple spatial limitations are imaged onto the array detector such that discrete subsets of the detecting units (pixels) in the array detector measure the signal from one and

only one of the multiple spatial limitations, the signal from any one spatial limitation being the combined signal from those pixels that receive the image from one of the spatial limitations.

- 15. A method according to claim 14, wherein the detector is a linear diode array.
- 5 16. A method according to claim 14, wherein the detector is a video camera.
 - 17. A method according to claim 14, wherein the detector is a charge transfer device.
 - 18. A method according to claim 17, wherein the charge transfer device is a charge-coupled device.
 - 19. A method according to any of claims 1-18, wherein the luminophore must be illuminated in order to emit light.
 - 20. A method according to any of claims 13-18, wherein all of the multiple spatial limitations are simultaneously illuminated during the measurement operation.
 - 21. A method according to any of claims 10-18, wherein the individual spatial limitations are singly illuminated only during the time period in which they are being measured.
- 15 22. A method according to any of claims 10-18, wherein the illumination is provided by a laser which is scanned in a raster fashion over some or all of the spatial limitations being measured, the scanning taking place at a rate substantially faster than the measurement process such that the illumination appears to the measurement process to be continuous in time and spatially uniform over the region being measured.
- 23. A method according to any of claims 1-22, wherein the spatial limitations are spatial limitations arranged in one or more arrays on a common carrier.
 - 24. A method according to claim 23, wherein the spatial limitations are wells in a plate of microtiter type.
- 25. A method according to any of claims 1-22 wherein the spatial limitations are domains
 defined on a substrate on which the cells are present.

22131DK1 Appendix A

- 26. A method according to claim 25 wherein the domains are domains established by the presence of the cells on the substrate in a pattern defining the domains.
- 27. A method according to claim 25 wherein the domains are domains established by the spatial pattern of the influence as it is applied to or contacted with the cells.
- 28. A method according to any of claims 1-27, wherein the recording is performed at a series of points in time, in which the application of the influence occurs at some time after the first time point in the series of recordings, the recording being performed, e.g., with a predetermined time spacing of from 0.1 seconds to 1 hour, preferably from 1 to 60 seconds, more preferably from 1 to 30 seconds, in particular from 1 to 10 seconds, over a time span of from 1 second to 12 hours, such as from 10 seconds to 12 hours, e.g., from 10 seconds to one hour, such as from 60 seconds to 30 minutes or 20 minutes.
 - 29. A method according to claim 28, wherein the recording is made at two points in time, one point being before, and the other point being after the application of the influence.
- 30. A method according to any of claims 1-29, wherein the cells are fixed at a point in time after the application of the influence at which the response has been predetermined to be significant, and the recording is made at an arbitrary later time.
 - 31. A method according to any of claims 1-30, wherein the luminophore is a luminophore that is capable of being redistributed in a manner that is physiologically relevant to the degree of the influence.
 - 32. A method according to any of claims 1-30, wherein the luminophore is a luminophore which is capable of associating with a component which is capable of being redistributed in manner which is physiologically relevant to the degree of the influence.
- 33. A method according to any of claims 1-30, wherein the luminophore is a luminophore which is capable of being redistributed in a manner which is experimentally determined to be correlated to the degree of the influence.
 - 34. A method according to any of claims 1-30, wherein the luminophore is a luminophore

which is capable of being redistributed, by modulation of the intracellular pathway, in substantially the same manner as the at least one component of the intracellular pathway.

- 35. A method according to any of claims 1-30, wherein the luminophore is a luminophore which is capable of being quenched upon spatial association with a component which is redistributed by modulation of the pathway, the quenching being measured as a decrease in the intensity of the luminescence.
- 36. A method according to any of claims 1-30, wherein the variation in spatially distributed light emitted by the luminophore is detected by a change in the resonance energy transfer between the luminophore and another luminescent entity capable of delivering energy to the luminophore, each of which has been selected or engineered to become part of, bound to or associated with particular components of the intracellular pathway, and one of which undergoes redistribution in response to the influence, thereby changing the amount of resonance energy transfer, the change in the resonance energy transfer being measured as a change in the intensity of emission from the luminophore.
- 37. A method according to any of claims 1-35, wherein the intensity of the light being recorded is a function of the fluorescence lifetime, polarisation, wavelength shift, or other property which is modulated as a result of the underlying cellular response.
- 38. A method according to any of claims 1-37, wherein the light to be measured passes through a filter which selects the desired component of the light to be measured and rejects other components.
 - 39. A method according to any of claims 2-38, wherein the intracellular pathway is an intracellular signalling pathway.
- 40. A method according to any of claims 1-39, wherein the luminophore is a fluorophore.
 - 41. A method according to any of claims 1-40, wherein the luminophore is a polypeptide encoded by and expressed from a nucleotide sequence harboured in the cells.

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- 42. A method according to any of claims 1-41 for detecting intracellular redistribution of a biologically active polypeptide affecting intracellular processes upon activation, the method comprising
- a) culturing one or more cells containing a nucleotide sequence coding for a hybrid polypeptide comprising a GFP which is N- or C-terminally tagged, optionally through a linker, to a biologically active polypeptide under conditions permitting expression of the nucleotide sequence,
 - b) modulating the activity of the biologically active polypeptide by incubating the cells with a substance having biological activity, and
- 10 c) measuring the fluorescence produced by the incubated cells and determining the result or variation with respect to the fluorescence, such result or variation being indicative of the redistribution of a biologically active polypeptide in said cells.
 - 43. A method according to claim 42, wherein the luminophore is a hybrid polypeptide comprising a fusion of at least a portion of each of two polypeptides one of which comprises a luminescent polypeptide and the other one of which comprises a biologically active polypeptide, as defined herein.
 - 44. A method according to claim 43, wherein the luminescent polypeptide is a GFP as defined herein.
- 45. A method according to claim 44, wherein the GFP is selected from the group consisting of green fluorescent proteins having the F64L mutation as defined herein.
 - 46. A method according to claim 45, wherein the GFP is a GFP variant selected from the group consisting of F64L-GFP, F64L-Y66H-GFP, F64L-S65T-GFP, and EGFP.
 - 47. A method according to claim 42, wherein the nucleotide sequence is a DNA sequence.
 - 48. A method according to claims 42-47, wherein the modulation is activation.
- 49. A method according to claims 42-47, wherein the modulation is deactivation.
 - 50. A method according to any of claims 1-49, wherein the cells are selected from the

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group consisting of fungal cells, such as yeast cells; invertebrate cells including insect cells; and vertebrate cells, such as mammalian cells.

- 51. A method according to claim 50, wherein the mechanically intact or permeabilised living cells are mammalian cells which, during the time period over which the influence is observed, are incubated at a temperature of 30°C or above, preferably at a temperature of from 32°C to 39°C, more preferably at a temperature of from 35°C to 38°C, and most preferably at a temperature of about 37°C.
- 52. A method according to any of claims 1-51, wherein the mechanically intact or permeabilised living cells are part of a matrix of identical or non-identical cells.
- 53. A method according to any of claims 41-52, wherein the nucleotide sequence has been introduced into the cells in the form of a nucleic acid construct coding for a fusion polypeptide comprising a biologically active polypeptide that is a component of an intracellular signalling pathway, or a part thereof, and a GFP.
 - 54. A method according to claim 53, wherein the nucleic acid construct is a nucleic acid construct coding for a fusion polypeptide comprising a biologically active polypeptide that is a component of an intracellular signalling pathway, or a part thereof, and an F64L mutant of GFP.
 - 55. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 53 or 54, wherein the biologically active polypeptide is a protein kinase or a phosphatase.
 - 56. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 53 55, wherein the GFP is N- or C-terminally tagged, optionally via a peptide linker, to the biologically active polypeptide or part thereof.
- 57. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 53, 54 or 56, wherein the biologically active polypeptide is a transcription factor or a part thereof which changes cellular localisation upon activation.

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- 58. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 53, 54 or 56, wherein the biologically active polypeptide is a protein, or a part thereof, which is associated with the cytoskeletal network and which changes cellular localisation upon activation.
- 59. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to any of claims 53-56, wherein the biologically active polypeptide is a protein kinase or a part thereof which changes cellular localisation upon activation.
 - 60. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a serine/threonine protein kinase or a part thereof capable of changing intracellular localisation upon activation.
 - 61. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a tyrosine protein kinase or a part thereof capable of changing intracellular localisation upon activation.
 - 62. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a phospholipid-dependent serine/threonine protein kinase or a part thereof capable of changing intracellular localisation upon activation.
- 20 63. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a cAMP-dependent protein kinase or a part thereof capable of changing cellular localisation upon activation.
- 64. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 63 which codes for a PKAc-F64L-S65T-GFP fusion.
 - 65. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a cGMP-dependent protein kinase or a part thereof capable of changing cellular localisation upon

activation.

- 66. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a calmodulin-dependent serine/threonine protein kinase or a part thereof capable of changing cellular localisation upon activation.
- 67. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a mitogen-activated serine/threonine protein kinase or a part thereof capable of changing cellular localisation upon activation.
- 10 68. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 67, which codes for an ERK1-F64L-S65T-GFP fusion.
 - 69. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 67, which codes for an EGFP-ERK1 fusion.
- 70. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a cyclin-dependent serine/threonine protein kinase or a part thereof capable of changing cellular localisation upon activation.
- 71. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 55 or 56, wherein the biologically active polypeptide is a protein phosphatase or a part thereof capable of changing cellular localisation upon activation.
 - 72. A method according to claim 53 -71, wherein the nucleic acid construct is a nucleic acid construct which is a DNA construct.
- 73. A method according to claim 53 -72, wherein the nucleic acid construct is a nucleic acid construct according to any of claims 53-72 wherein the gene encoding GFP is derived from Aequorea victoria.

- 74. A method according to claim 73, wherein the nucleic acid construct is a nucleic acid construct according to claim 73 in which the gene encoding GFP is the gene encoding EGFP as defined herein.
- 75. A method according to claim 73, wherein the nucleic acid construct is a nucleic acid construct according to claim 73 in which the gene encoding a GFP is a gene encoding a GFP variant selected from F64L-GFP, F64L-Y66H-GFP and F64L-S65T-GFP.
 - 76. A method according to claims 72 and 74, wherein the nucleic acid construct is a DNA construct according to claims 72 and 74 or, where applicable, 75, which is a construct as identified by any of the DNA sequences shown in SEQ ID NO: 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, 78, 108, 110, 112, 114, 116, 118, 120, 122, 124, 126, 128, 130, 132, 134, 136, 138, 140, 142, 144, 146, 148, 150, and 152 or is a variant thereof capable of encoding the same fusion polypeptide or a fusion polypeptide which is biologically equivalent thereto, as defined herein.
 - 77. A method comprising a cell containing a nucleic acid construct according to any of claims 53-76 and capable of expressing the sequence encoded by the construct.
 - 78. A method comprising a cell according to claim 77, which is a eukaryotic cell.
 - 79. A method comprising a cell according to claim 77, which is selected from the group consisting of fungal cells, such as yeast cells; invertebrate cells, including insect cells, and vertebrate cells, such as mammalian cells.
- 80. A method according to any of claims 1-79, as used in a screening program as defined herein.
 - 81. A method according claim 80, wherein the method is a screening program for the identification of a biologically active substance as defined herein that directly or indirectly affects an intracellular signalling pathway and is potentially useful as a medicament, wherein the result of the individual measurement of each substance being screened which indicates its potential biological activity is based on measurement of the redistribution of spatially resolved luminescence in living cells and which undergoes a change in distribution upon activation of an intracellular signalling

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pathway.

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- 82. A method according to claim 80, wherein the method is a screening program for the identification of a biologically toxic substance as defined herein that exerts its toxic effect by interfering with an intracellular signalling pathway, wherein the result of the individual measurement of each substance being screened which indicates its potential biologically toxic activity is based on measurement of the redistribution of said fluorescent probe in living cells and which undergoes a change in distribution upon activation of an intracellular signalling pathway.
- 83. A method according to any of claims 1-82 wherein a fluorescent probe is used in backtracking of signal transduction pathways as defined herein.
- 84. A method according to any of claims 1-83, for treating a condition or disease related to the intracellular function of a protein kinase comprising administering to a patient suffering from said condition or disease an effective amount of a compound which has been discovered by any method.
- 15 85. A compound that modulates a component of an intracellular pathway as defined herein, as determined by any method according to any of claims 1-83.
 - 86. A medical composition comprising a therapeutic amount of a compound identified according to any method according to any of claims 1-83.
- 87. A method of selectively treating a patient suffering from an ailment which responds to medical treatment comprising obtaining a primary cells from said patient, transfecting the cells with at least one DNA sequence encoding a fluorescent probe according to any of the preceding claims, culturing the cells under conditions permitting the expression of said probes and exposing it to an array of medicaments suspected of being capable of alleviating said ailment, then comparing changes in fluorescence patterns or redistribution patterns of the fluorescent probes in the intact living cells to detect the cellular response to the specific medicaments (obtaining a cellular action profile), then selecting a medicament(s) based on desired activity and acceptable level of side effects and administering an effective amount of said medicament(s) to said

patient.

88. A method according to any of claims 1-83 of identifying a drug target among the group of biologically active polypeptides that are components of intracellular signalling pathways.

Figure 1

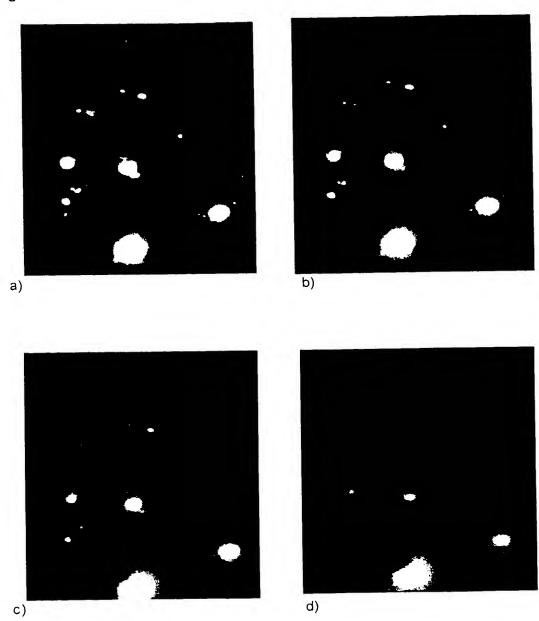
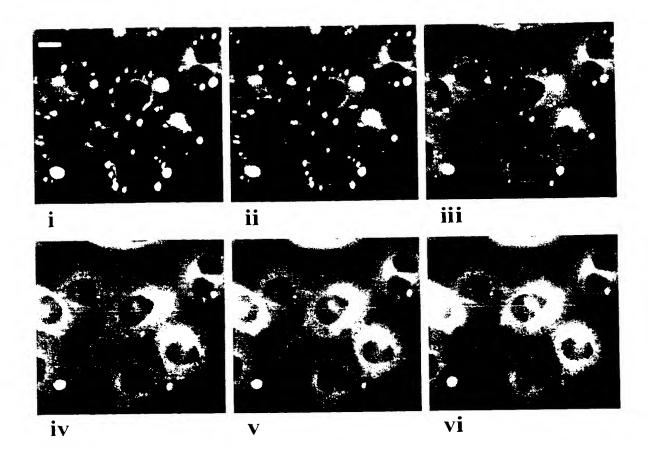
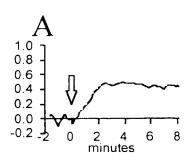


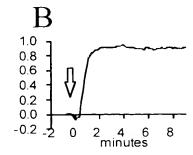
Figure 2

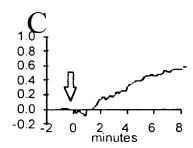


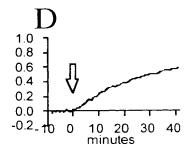
\$ 5 OKT. 1998

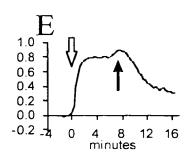
Figure 3

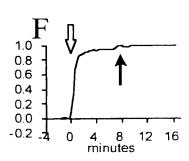


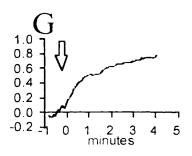












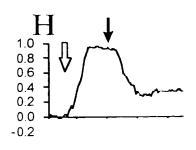
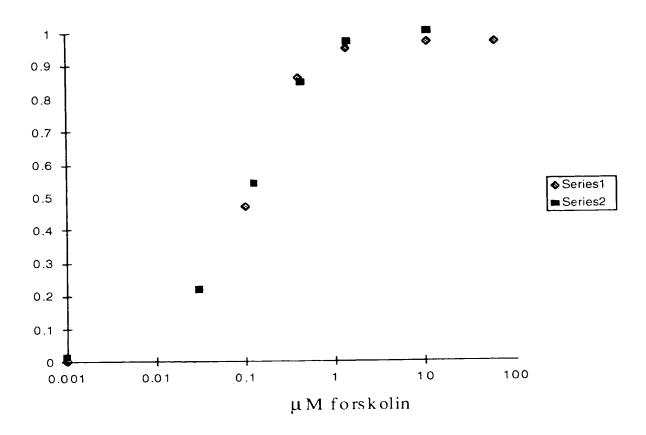


Figure 4



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Figure 5

[forskolin]µM	$t_{1/2max}/s$	t _{max} /s
1	115±21	310±31
10	69±14	224±47
50	47±10	125±28

Figure 6

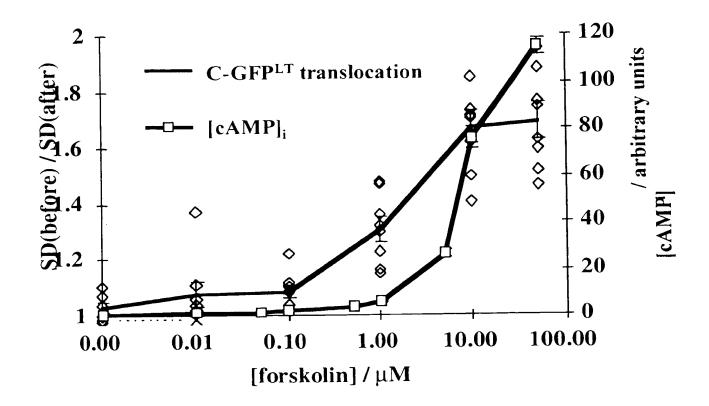
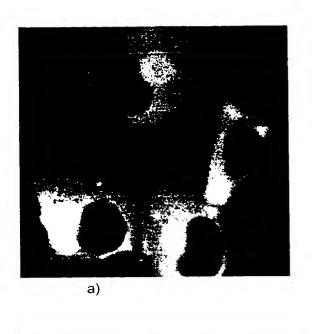
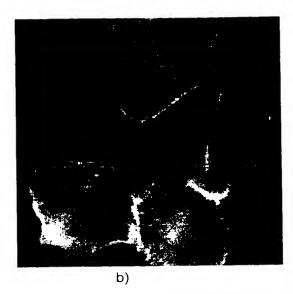
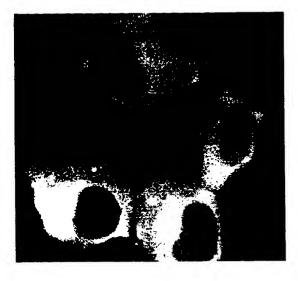


Figure 7







c)

Figure 8

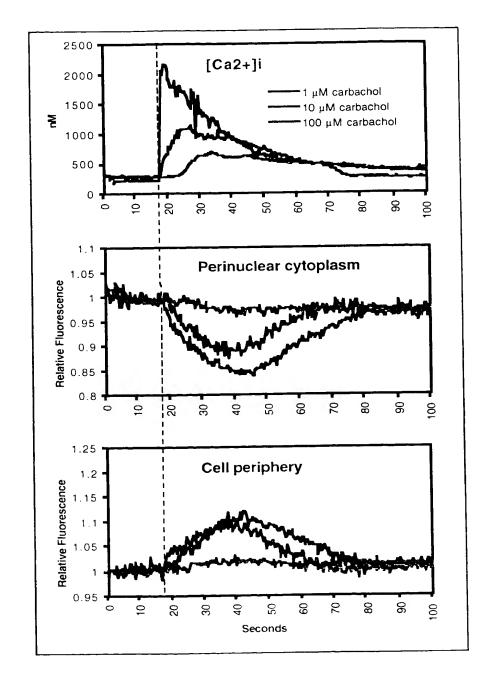
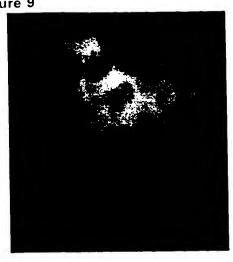
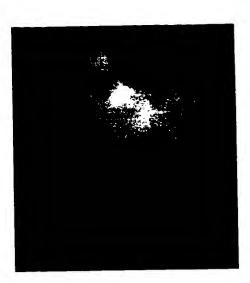


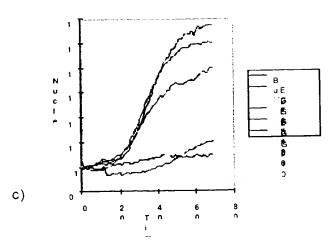
Figure 9





b)

a)



9

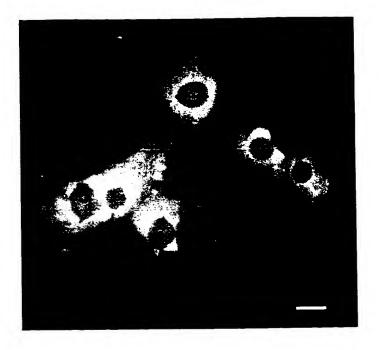
d)

Nucleus/Cyt.

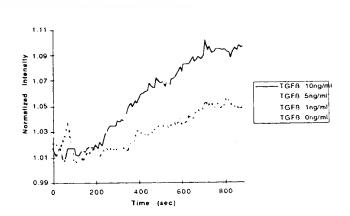
1.2

Figure 10

a)



b)



c)

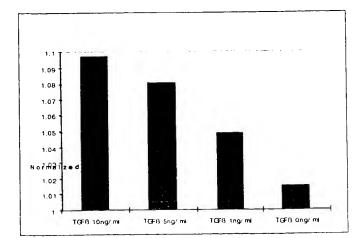


Figure 11

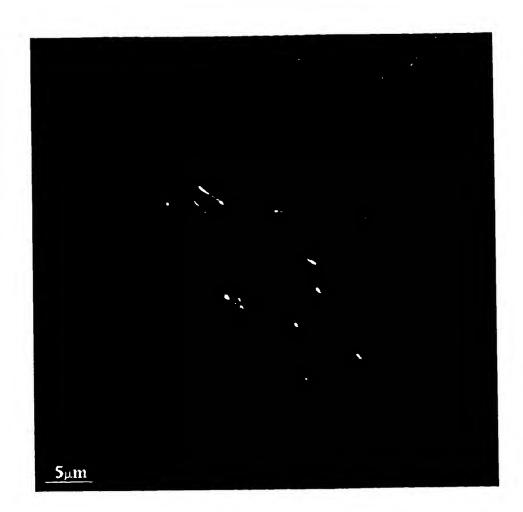
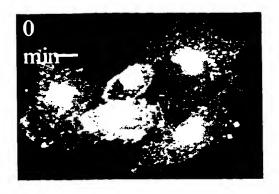
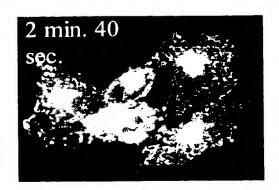
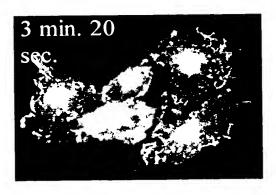


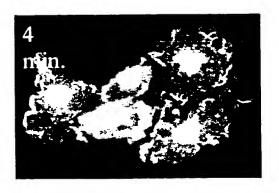
Fig. 12











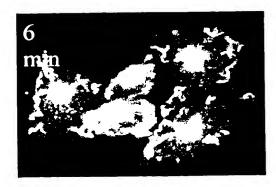


Figure 13

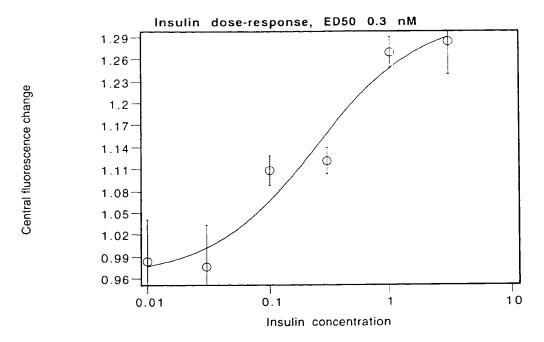


Figure 14

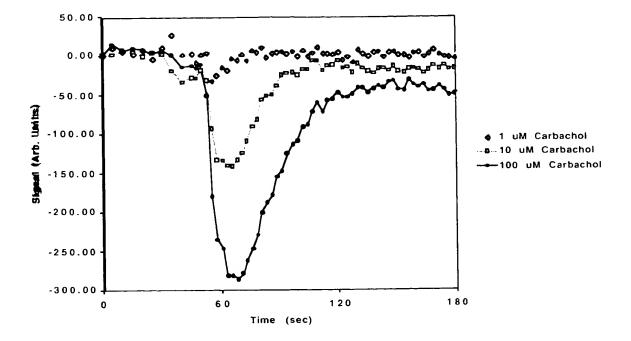


Figure 15

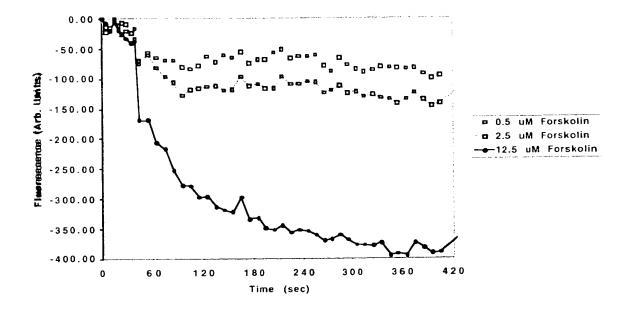
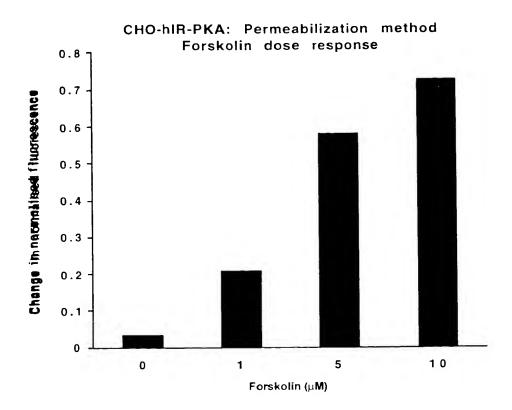


Figure 16



SEQUENCE LISTING

- (1) GENERAL INFORMATION
- (i) APPLICANT: NovoNordisk, BioImage
- (ii) TITLE OF THE INVENTION: An Improved Method of Detecting Cellular Translocation of Biologically Active Polypeptides Using Fluorescense Imaging
- (iii) NUMBER OF SEQUENCES: 165
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: NovoNordisk, BioImage
 - (B) STREET: Mørkhøjbygade 28
 - (C) CITY: Søborg
 - (D) STATE: DK
 - (E) COUNTRY: DENMARK
 - (F) ZIP: 2860
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Diskette
 - (B) COMPUTER: IBM Compatible
 - (C) OPERATING SYSTEM: DOS
 - (D) SOFTWARE: FastSEQ for Windows Version 2.0
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: , PV&P R
 - (B) REGISTRATION NUMBER:
 - (C) REFERENCE/DOCKET NUMBER:
 - (2) INFORMATION FOR SEQ ID NO:1:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 53 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

TTGGACACAA GCTTTGGACA CGGCGCGCCA TGAGTAAAGG AGAAGAACTT TTC

- (2) INFORMATION FOR SEQ ID NO:2:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 53 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

GTCATCTTCT CGAGTCTTAC TCCTGAGGTT TGTATAGTTC ATCCATGCCA TGT	53
(2) INFORMATION FOR SEQ ID NO:3:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 54 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:	
TTGGACACAA GCTTTGGACA CCCTCAGGAT ATGGGCAACG CCGCCGCCGC CAAG	54
(2) INFORMATION FOR SEQ ID NO:4:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 55 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
GTCATCTTCT CGAGTCTTTC AGGCGCGCCC AAACTCAGTA AACTCCTTGC CACAC	55
(2) INFORMATION FOR SEQ ID NO:5:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 55 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:	
TTGGACACAA GCTTTGGACA CCCTCAGGAT ATGGCTGACG TTTACCCGGC CAACG	55
(2) INFORMATION FOR SEQ ID NO:6:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 55 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:	
GTCATCTTCT CGAGTCTTTC AGGCGCGCCC TACTGCACTT TGCAAGATTG GGTGC	55
(2) INFORMATION FOR SEQ ID NO:7:	

(i) SEQUENCE CHARACTERISTICS:

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TYGGACACAA GCTTYGGACA CCCTCAGGAT ATGGCGGCGG CGGCGGCGGC TCCGGGGGGC GGGG	60 64
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(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 55 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
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(2) INFORMATION FOR SEQ ID NO:9:	
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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:	
TAGAATTCAA CCATGGCGGC GGCGGCGCGC	30
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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:	
TAGGATCCCT AGGGGGCCTC CAGCACTCC	29
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(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:	
TACTCGAGTA ACCATGGCGG CGGCGGCGGC G	31
(2) INFORMATION FOR SEQ ID NO:12:	
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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:	
TAGGATCCAT AGATCTGTAT CCTGG	25
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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:	
TAGGATCCTT AAGATCTGTA TCCTGG	26
(2) INFORMATION FOR SEQ ID NO:14:	
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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:	
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(2) INFORMATION FOR SEQ ID NO:15:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 28 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:	
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(2) INFORMATION FOR SEQ ID NO:16:	

(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 29 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:	
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(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 28 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:	
GTCTCGAGCC ATCATGAGCA GAAGCAAG	28
(2) INFORMATION FOR SEQ ID NO:18:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 27 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:	
GTGGATCCCA CTGCTGCACC TGTGCTA	27
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(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 40 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:	
CGCGAATTCC GCCACCATGA GTGCTGAGGG GTACCAGTAC	40
(2) INFORMATION FOR SEQ ID NO:21:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 32 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:	
CGCGGATCCT GTCGCCTCTG CTGTGCATAT AC	32
(2) INFORMATION FOR SEQ ID NO:22:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 30 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(vi) ORIGINAL SOURCE: (A) ORGANISM: p85-top-C	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	
GGGAGATCTA TGAGTGCTGA GGGGTACCAG	30
(2) INFORMATION FOR SEQ ID NO:23:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 34 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:	
GGGCGGATCC TCATCGCCTC TGCTGCAT ATAC	34
(2) INFORMATION FOR SEQ ID NO:24:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 33 base pairs(E) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

A TOP TO THE TOP TO TH	33
GTGAATTCGA CCATGTCGTC CATCTTGCCA TTC	33
(O) ANTIONNAMION FOR CEO ID NO. 25.	
(2) INFORMATION FOR SEQ ID NO:25:	
(i) SEQUENCE CHARACTERISTICS:	
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(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
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	2.1
GTGGTACCCA TGACATGCTT GAGCAACGCA C	31
(2) INFORMATION FOR SEQ ID NO:26:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 32 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:	
GTGGTACCTT ATGACATGCT TGAGCAACGC AC	32
(2) INFORMATION FOR SEQ ID NO:27:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 31 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(5) 101020011 1111011	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:	
GTGAATTCGT CAATGGAGCT GGAAAACATC G	31
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(A) LENGTH: 30 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(D) Totoboot. Timedi	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:	
(All) DESCRIBE PERSONS TO THE STATE OF THE	
GTGGATCCCT GCTGCTTCCG GTGGAGTTCG	30
0100,110001 001001.000 01001.	
(2) INFORMATION FOR SEO ID NO:29:	

(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 31 base pairs

(E) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
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(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 32 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
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(2) INFORMATION FOR SEQ ID NO:36:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 27 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:	
GTCTCGAGGC ACCATGAGCG ACGTGGC	27
(2) INFORMATION FOR SEQ ID NO:37:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 27 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:	
TGGGATCCGA GGCCGTGCTG CTGGCCG	27
(2) INFORMATION FOR SEQ ID NO:38:	

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1896 base pairs(B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...1891
 - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

ATG Met 1	GTG Val	AGC Ser	AAG Lys	GGC Gly 5	GAG Glu	GAG Glu	CTG Leu	TTC Phe	ACC Thr 10	GGG Gly	GTG Val	GTG Val	CCC Pro	ATC Ile 15	CTG Leu	48
GTC Val	GAG Glu	CTG Leu	GAC Asp 20	GGC Gly	GAC Asp	GTA Val	AAC Asn	GGC Gly 25	CAC His	AAG Lys	TTC Phe	AGC Ser	GTG Val 30	TCC Ser	GGC Gly	96
GAG Glu	GGC Gly	GAG Glu 35	GGC Gly	GAT Asp	GCC Ala	ACC Thr	TAC Tyr 40	GGC Gly	AAG Lys	CTG Leu	ACC Thr	CTG Leu 45	AAG Lys	TTC Phe	ATC Ile	144
TGC Cys	ACC Thr 50	ACC Thr	GGC Gly	AAG Lys	CTG Leu	CCC Pro 55	GTG Val	CCC Pro	TGG Trp	CCC Pro	ACC Thr 60	CTC Leu	GTG Val	ACC Thr	ACC Thr	192
CTG Leu 65	ACC Thr	TAC Tyr	GGC Gly	GTG Val	CAG Gln 70	TGC Cys	TTC Phe	AGC Ser	CGC Arg	TAC Tyr 75	CCC Pro	GAC Asp	CAC His	ATG Met	AAG Lys 80	240
CAG Gln	CAC	GAC Asp	TTC Phe	TTC Phe 85	AAG Lys	TCC Ser	GCC Ala	ATG Met	CCC Pro 90	GAA Glu	GGC Gly	TAC Tyr	GTC Val	CAG Gln 95	GAG Glu	288
CGC Arg	ACC Thr	ATC Ile	TTC Phe 100	Phe	AAG Lys	GAC Asp	GAC Asp	GGC Gly 105	Asn	TAC	: AAG	ACC Thr	CGC Arg	Ala	GAG Glu	336
GTG Val	AAG Lys	TTC Phe 115	Glu	GGC Gly	GAC Asp	ACC Thr	CTG Leu 120	Val	AAC Asn	CGC Arg	ATC	GAG Glu 125	Leu	AAG Lys	GGC	384
ATC Ile	GAC Asp 130	Phe	: AAG : Lys	GAG Glu	GAC Asp	GGC Gly 135	Asn	: ATC	: CTG	GGC Gly	G CAC / His 140	. Lys	CTC	G GAC	TAC Tyr	432
AAC Asr 145	тух	AAC Asr	AGC Ser	CAC His	: AAC : Asr 150	. Val	TAT Tyr	ATC	ATC Met	GCC Ala 155	a Asp	D AAC	G CAG	G AAC n Lys	AAC Asn 160	480
GG(Gly	ATC	AAC Lys	G GTC	AAC Asr 165	n Phe	AAC Lys	ATC	e Arg	CAC His 170	AS:	n Ile	GAC Glu	G GAG	C GGG p Gl ₂ 17	C AGC / Ser	528

GTG Val	CAG Gln	CTC Leu	GC Al 18	a A	AC (sp H	CAC '	TAC Tyr	CAG Gln	CAG Gln 185	As	C A n T	CC (CCC Pro	ATC Ile	GGC Gly 190	GA As	AC (3G(31 ₎	C 7	576
CCC Pro	GTG Val	CTG Leu 195	Le	G C u P	cc (GAC Asp	AAC Asn	CAC His 200	TAC Tyr	CT Le	G A	.GC . Ser	ACC Thr	CAG Gln 205	TCC	G(CC (CT(Le	G U	624
AGC Ser	AAA Lys 210	Asp	CC Pr	C A	AC Asn	GAG Glu	AAG Lys 215	CGC Arg	GAT Asp	CA Hi	CA	ATG 1et	GTC Val 220	CTG Leu	CTC	G. G	AG 1u	TT Ph	C e	672
GTG Val 225	Thr	GCC Ala	C GC	cc c	GG Gly	ATC Ile 230	ACT Thr	CTC Leu	GG(AT Me	et A	GAC Asp 235	GAG Glu	CTG Leu	TAC Tyi	C A	AG Ys	TC Se 24	er	720
GGA Gly	. CTC	AG Arg	A TY	er A	CGA Arg 245	GCT Ala	CAA Gln	GCT	TC(r A	AT 1 sn 1 50	ICA Ser	ACC Thr	ATG Met	GC(a A	CG la 255	GC A1	CG La	768
GCG Ala	GC' Ala	r CA	n G	GG (ly (60	GGC Gly	GGG Gly	GGC Gly	Gly	GA Gl 26	u P	cc (CGT Arg	AGA Arg	ACC Thr	GA G1 27	u	GG Gly	GT Vá	rc al	816
GG(Gly	C CC 7 Pr	G GG o Gl 27	y V	TC al	CCG Pro	GGG Gly	GAG Glu	GT(Va. 280	l Gl	G A u M	TG let	GTG Val	AAG Lys	GGG Gly 289	y G1	G (CCG Pro	T'	IC he	864
GA(As _l	C GT O Va 29	l Gl	у Р	CG ro	CGC Arg	TAC Tyr	ACC Thr 295	Gli	g TI n Le	rG C	AG 31n	TAC Tyr	11e	GG(G1;	C GA y Gl	u (GGC Gly	G A	CG la	912
ТА: Ту: 30	r Gl	C AT	G G	TC /al	AGC Ser	TCG Ser 310	Ala	та а ту	T GA	C C	CAC	GTG Val 315	Arg	C AA g Ly	G AC	eT ar	CGC Arg	V	TG Tal 20	960
GC Al	C AT	C A!	AG A /s I	AAG Jys	ATC Ile 325	Ser	CCC Pro	TT DPh	C G/ e G	lu F	CAT His 330	CAC Glr	ACO Th	C TA r Ty	C TY T C	GC ys	CAG Glr 335	1 4	r.ec r.ec	1008
AC Th	G C	rc co eu A:	rg (GAG Glu 340	Il€	CAC Gli	ı Il	C CT e Le	u L	eu i	Arg	Phe	e Ar	C CA g Hi	.s G	AG 1u 50	AAT Asr	r C	GTC /al	1056
AT Il	C G .e G	ly I	TC (le 2 55	CGA Arg	GAC Asp	AT'	r CT e Le	G CC u Ar 36	rg A	CG '	TCC Ser	ACC Th:	C CT r Le	G GA eu Gl	lu A	CC la	ATC	G A	AGA Arg	1104
G.F A.s	sp V	TC T al T 70	AC . yr	ATT Ile	GT(Va	G CA l Gl	G GA n As 37	p Le	rg A eu M	TG let	GAG Glu	AC Th	T GA r As 38	AC CT	rg T eu T	λt.	AA Ly	G '	TTG Leu	1152
L	rg A eu L 85	AA A ys S	.GC Ser	CAG Gln	CAG Gl:	G CT n Le 39	u Se	SC A	AT C	SAC Asp	CAT His	AT 11 39	e C	GC T. ys T	AC I	rTC Phe	CT Le	c u	TAC Tyr 400	1200
C. G	AG A ln I	TC C	TG .eu	CGG Arg	GG GG	C CI y Le	C AA	AG T /s T	AC A	ATC Ile	CAC	TC S Se	c G er A	CC A la A	AC (GTG Val	CI Le	C eu	CAC His	1248

415 410 405 CGA GAT CTA AAG CCC TCC AAC CTG CTC AGC AAC ACC ACC TGC GAC CTT Arg Asp Leu Lys Pro Ser Asn Leu Leu Ser Asn Thr Thr Cys Asp Leu 425 AAG ATT TGT GAT TTC GGC CTG GCC CGG ATT GCC GAT CCT GAG CAT GAC 1344 Lys Ile Cys Asp Phe Gly Leu Ala Arg Ile Ala Asp Pro Glu His Asp 440 CAC ACC GGC TTC CTG ACG GAG TAT GTG GCT ACG CGC TGG TAC CGG GCC 1392 His Thr Gly Phe Leu Thr Glu Tyr Val Ala Thr Arg Trp Tyr Arg Ala 455 CCA GAG ATC ATG CTG AAC TCC AAG GGC TAT ACC AAG TCC ATC GAC ATC 1440 Pro Glu Ile Met Leu Asn Ser Lys Gly Tyr Thr Lys Ser Ile Asp Ile 470 TGG TCT GTG GGC TGC ATT CTG GCT GAG ATG CTC TCT AAC CGG CCC ATC 1488 Trp Ser Val Gly Cys Ile Leu Ala Glu Met Leu Ser Asn Arg Pro Ile 485 TTC CCT GGC AAG CAC TAC CTG GAT CAG CTC AAC CAC ATT CTG GGC ATC 1536 Phe Pro Gly Lys His Tyr Leu Asp Gln Leu Asn His Ile Leu Gly Ile 505 500 CTG GGC TCC CCA TCC CAG GAG GAC CTG AAT TGT ATC ATC AAC ATG AAG 1584 Leu Gly Ser Pro Ser Gln Glu Asp Leu Asn Cys Ile Ile Asn Met Lys 520 515 GCC CGA AAC TAC CTA CAG TCT CTG CCC TCC AAG ACC AAG GTG GCT TGG 1632 Ala Arg Asn Tyr Leu Gln Ser Leu Pro Ser Lys Thr Lys Val Ala Trp 535 GCC AAG CTT TTC CCC AAG TCA GAC TCC AAA GCC CTT GAC CTG CTG GAC 1680 Ala Lys Leu Phe Pro Lys Ser Asp Ser Lys Ala Leu Asp Leu Leu Asp 555 550 CGG ATG TTA ACC TTT AAC CCC AAT AAA CGG ATC ACA GTG GAG GAA GCG 1728 Arg Met Leu Thr Phe Asn Pro Asn Lys Arg Ile Thr Val Glu Glu Ala 570 565 CTG GCT CAC CCC TAC CTG GAG CAG TAC TAT GAC CCG ACG GAT GAG CCA 1776 Leu Ala His Pro Tyr Leu Glu Gln Tyr Tyr Asp Pro Thr Asp Glu Pro 585 GTG GCC GAG GAG CCC TTC ACC TTC GCC ATG GAG CTG GAT GAC CTA CCT 1824 Val Ala Glu Glu Pro Phe Thr Phe Ala Met Glu Leu Asp Asp Leu Pro 600 AAG GAG CGG CTG AAG GAG CTC ATC TTC CAG GAG ACA GCA CGC TTC CAG 1872 Lys Glu Arg Leu Lys Glu Leu Ile Phe Gln Glu Thr Ala Arg Phe Gln 615 1896 CCC GGA GTG CTG GAG GCC C CCTAG

Pro Gly Val Leu Glu Ala Pro

(2) INFORMATION FOR SEQ ID NO:39:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 631 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 1.0 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 20 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 35 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 75 70 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 90 85 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 110 105 100 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 125 120 115 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 140 135 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 155 150 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 175 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Qly 190 185 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 205 200 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 220 215 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 235 230 Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Thr Met Ala Ala Ala 245 250 Ala Ala Gln Gly Gly Gly Gly Glu Pro Arg Arg Thr Glu Gly Val 265 270 260 Gly Pro Gly Val Pro Gly Glu Val Glu Met Val Lys Gly Gln Pro Phe 280 285 275 Asp Val Gly Pro Arg Tyr Thr Gln Leu Gln Tyr Ile Gly Glu Gly Ala 295 Tyr Gly Met Val Ser Ser Ala Tyr Asp His Val Arg Lys Thr Arg Val 315 310 Ala Ile Lys Lys Ile Ser Pro Phe Glu His Gln Thr Tyr Cys Gln Arg 330 325 Thr Leu Arg Glu Ile Gln Ile Leu Leu Arg Phe Arg His Glu Asn Val 345 340

Ile Gly Ile Arg Asp Ile Leu Arg Ala Ser Thr Leu Glu Ala Met Arg 355 360 365 Asp Val Tyr Ile Val Gln Asp Leu Met Glu Thr Asp Leu Tyr Lys Leu 370 375 380 Leu Lys Ser Gln Gln Leu Ser Asn Asp His Ile Cys Tyr Phe Leu Tyr 385 390 395 400 Gln Ile Leu Arg Gly Leu Lys Tyr Ile His Ser Ala Asn Val Leu His 410 415 405 Arg Asp Leu Lys Pro Ser Asn Leu Leu Ser Asn Thr Thr Cys Asp Leu 420 425 430 Lys Ile Cys Asp Phe Gly Leu Ala Arg Ile Ala Asp Pro Glu His Asp 435 440 His Thr Gly Phe Leu Thr Glu Tyr Val Ala Thr Arg Trp Tyr Arg Ala 460 450 455 Pro Glu Ile Met Leu Asn Ser Lys Gly Tyr Thr Lys Ser Ile Asp Ile 470 475 480 Trp Ser Val Gly Cys Ile Leu Ala Glu Met Leu Ser Asn Arg Pro Ile 485 490 495 Phe Pro Gly Lys His Tyr Leu Asp Gln Leu Asn His Ile Leu Gly Ile 500 505 510 Leu Gly Ser Pro Ser Gln Glu Asp Leu Asn Cys Ile Ile Asn Met Lys 515 520 525 Ala Arg Asn Tyr Leu Gln Ser Leu Pro Ser Lys Thr Lys Val Ala Trp 530 535 540 Ala Lys Leu Phe Pro Lys Ser Asp Ser Lys Ala Leu Asp Leu Leu Asp 545 550 555 560 Arg Met Leu Thr Phe Asn Pro Asn Lys Arg Ile Thr Val Glu Glu Ala 565 570 575 Leu Ala His Pro Tyr Leu Glu Gln Tyr Tyr Asp Pro Thr Asp Glu Pro 580 585 Val Ala Glu Glu Pro Phe Thr Phe Ala Met Glu Leu Asp Asp Leu Pro 595 600 605 Lys Glu Arg Leu Lys Glu Leu Ile Phe Gln Glu Thr Ala Arg Phe Gln 620 Pro Gly Val Leu Glu Ala Pro

(2) INFORMATION FOR SEQ ID NO:40:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1818 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...1815
 - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG
Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu

1 5 10 15

GTC Val	GAG Glu	CTG Leu	GAC Asp 20	GGC Gly	GAC Asp	GTA . Val .	AAC Asn	GGC Gly 25	CAC His	AAG Lys	TTC Phe	AGC Ser	GTG Val 30	TCC Ser	GGC Gly	96
GAG Glu	GGC Gly	GAG Glu 35	GGC Gly	GAT Asp	GCC Ala	ACC Thr	TAC Tyr 40	GGC Gly	AAG Lys	CTG Leu	ACC Thr	CTG Leu 45	AAG Lys	TTC Phe	ATC Ile	144
TGC Cys	ACC Thr 50	ACC Thr	GGC Gly	AAG Lys	CTG Leu	CCC Pro 55	GTG Val	CCC Pro	TGG Trp	CCC Pro	ACC Thr 60	CTC Leu	GTG Val	ACC Thr	ACC Thr	192
CTG Leu 65	ACC Thr	TAC Tyr	GGC Gly	GTG Val	CAG Gln 70	TGC Cys	TTC Phe	AGC Ser	CGC Arg	TAC Tyr 75	CCC Pro	GAC Asp	CAC	ATG Met	AAG Lys 80	240
CAG Gln	CAC His	GAC Asp	TTC Phe	TTC Phe 85	AAG Lys	TCC Ser	GCC Ala	ATG Met	CCC Pro 90	GAA Glu	GGC Gly	TAC Tyr	GTC Val	CAG Gln 95	GAG Glu	288
CGC Arg	ACC Thr	ATC	TTC Phe 100	Phe	AAG Lys	GAC Asp	GAC Asp	GGC Gly 105	Asn	TAC Tyr	AAC Lys	ACC Thr	CGC Arg	Ala	GAG Glu	336
GTC Val	AAG Lys	TTC Phe 115	e Glu	GGC Gly	GAC Asp	ACC Thr	CTG Leu 120	Val	AAC Asn	CGC Arg	ATC	GAC Glu 125	ı Let	AAC Lys	G GGC G Gly	384
)TA	GAC Asr 130	Phe	AAC Lys	GAG Glu	GAC Asp	GGC Gly 135	Asn	TATC	CTC	GG(G CAC / His 140	. Lys	G CTC	G GAG	З ТАС 1 ТУ1	432
AA(Asi 14	n Tyr	AAC Asr	C AGO	CAC His	AAC Asn 150	Val	ТАТ Туг	T ATC	ATC Met	GCC Ala 15	a As	C AAG p Ly:	G CAG s Gli	G AAG	G AAG s Asi 160	1
GG G1	C AT(C AAG e Ly:	G GTY	AAG Asr 165	n Phe	AAC Lys	ATC	C CGC E Arg	CAC His	s Ası	n Il	C GA	G GAG u Asj	C GG p G1 17	C AGG y Se: 5	528 r
GT Va	G CAG	G CTO	u Al	C GAG a Ası	o His	з Туз	Gli	G CAG n Gli 18!	n As	C AC	c cc r Pr	C AT o Il	C GG e Gl 19	y As	c GG p G1	C 576 Y
CC Pr	C GT	G CT l Le 19	u Le	G CC	C GAG o Ası	AA C ASI	C CAG n Hi 20	s Ty	C CT r Le	G AG u Se	C AC	C CA ir Gl 20	n Se	c GC r Al	C CT a Le	G 624 u
AC Se	C AA er Ly 21	s As	c cc p Pr	C AA o As	C GAG	G AAG u Ly 21	s Ar	C GA g As	T CA p Hi	C AT s Me	G G1 et Va 22	il Le	G CI eu Le	'G G? eu GI	AG TI lu Ph	rc 672 ie
Vá	NG AC al Th	c GC ar Al	CC GC .a Al	c GG a G1	G AT y Il 23	e Th	T CT r Le	c GG u G1	С АТ У Ме	G GA et As 23	sp G.	AG CT	rg T <i>i</i> eu Ty	AC AZ	AG TO ys Se 24	er
G(G:	GA CI ly Le	C AC	GA TO	T CG	A GT g Va	A AC 1 Th	C AT r Me	G GC	G GC .a Al	CG GC La Al	CG G(la A	CG GG la A	CG GG la A	CG G la G	GC CC ly Pi	CG 768

				245					250					255		
GAG .	ATG Met	GTC Val	CGC Arg 260	GGG Gly	CAG Gln	GTG Val	TTC Phe	GAC Asp 265	GTG Val	GGG Gly	CCG Pro	CGC Arg	TAC Tyr 270	ACT Thr	AAT Asn	816
CTC Leu	TCG Ser	TAC Tyr 275	ATC Ile	GGA Gly	GAA Glu	GGC Gly	GCC Ala 280	TAC Tyr	GGC Gly	ATG Met	GTT Val	TGT Cys 285	TCT Ser	GCT Ala	TAT Tyr	864
GAT Asp	AAT Asn 290	CTC Leu	AAC Asn	AAA Lys	GTT Val	CGA Arg 295	GTT Val	GCT Ala	ATC Ile	AAG Lys	AAA Lys 300	ATC Ile	AGT Ser	CCT Pro	TTT Phe	912
GAG Glu 305	CAC His	CAG Gln	ACC Thr	TAC Tyr	TGT Cys 310	CAG Gln	AGA Arg	ACC Thr	CTG Leu	AGA Arg 315	GAG Glu	ATA Ile	AAA Lys	ATC Ile	CTA Leu 320	960
CTG Leu	CGC Arg	TTC Phe	AGA Arg	CAT His 325	GAG Glu	AAC Asn	ATC Ile	ATC Ile	GGC Gly 330	ATC Ile	AAT Asn	GAC Asp	ATC Ile	ATC Ile 335	CGG Arg	1008
GCA Ala	CCA Pro	ACC Thr	ATT Ile 340	Glu	CAG Gln	ATG Met	AAA Lys	GAT Asp 345	GTA Val	TAT Tyr	ATA Ile	GTA Val	Glm 350	Asp	CTC Leu	1056
ATG Met	GAG Glu	ACA Thr	Asp	CTT Leu	TAC Tyr	AAG Lys	CTC Leu 360	Leu	AAG Lys	ACA Thr	CAG Gln	CAC His	Leu	AGC Ser	AAT Asn	1104
GAT Asp	CAT His	Il∈	TGC Cys	TAT Tyr	TTT Phe	CTT Leu 375	Туг	CAG	ATC	CTC Lev	AGA Arg 380	G13	TT?	A AAG 1 Lys	TAT Tyr	1152
ATA Ile 385	His	TCA Ser	A GCT	raa r	GTT Val	Leu	CAC His	CGT Arg	Asp Asp	CTC Lev 395	ı Lys	CC:	r TCC	C AAC r Asr	CTC Leu 400	1200
CTG Leu	CTC Lev	AAG 1 Ast	ACC Thi	C ACT	Cys	GAT Asp	CTC	AAG 1 Lys	ATC 116 410	S CA	GAC S Asi	TT'	r GG¢ ≥ Gl;	C CT y Lev 415	GCC Ala	1248
CGI Arg	GT Val	r GC/ 1 Ala	A GA A Ası 420	p Fro	A GAC	CAT His	GAS Asp	r CAT His 425	Thi	A GG(r Gly	G TTO y Phe	TTY E Le	G AC u Th 43	r Gl	TAT Tyr	1296
GTA Val	A GCC	C ACC a Th:	r Ar	r TG(g Tr]	Э ТАС Э Туі	AGA Arg	A GC' g Ala 440	a Pro	A GAZ	A AT	T ATG	G TT t Le 44	u As	T TC	C AAG r Lys	1344
GG1 Gly	г та у ту: 45	r Th	C AAG r Ly	G TCC s Se:	C ATT	r GA e Ası 459	o Il	T TG e Trj	G TC' p Se:	T GT r Va	G GG 1 G1 46	у Су	C AT	C CT e Le	G GCA u Ala	1392
GAG Glu 465	u Me	G CT t Le	A TC u Se	C AA r As	C AGO n Arg 47	g Pro	r AT o Il	C TT e Ph	C CC. e Pr	A GG o G1 47	A ГА	G CA s Hi	T TA S T	C CT	T GAC u Asp 480	1440

			CTG Leu						1488
			AAT Asn						1536
 			GTG Val						1584
			TTA Leu						1632
			GAA Glu 550						1680
			GAT Asp						1728
 	_	-	GAC Asp					ATT Ile	1776
			CGA Arg				TAA		1818

(2) INFORMATION FOR SEQ ID NO:41:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 605 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein
(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 15 1 5 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile **4**5 35 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 60 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 70 75 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 90 85

Arg 7	Thr	Ile	Phe 100	Phe	Lys	Asp		Gly 105	Asn	Tyr	Lys '		Arg 110	Ala	Glu
Val I		Phe		Gly	Asp	Thr			Asn	Arg		Glu 125	Leu	Lys	Gly
Ile A			Lys	Glu	Asp	Gly 135		Ile	Leu	Gly	His 140	Lys	Leu	Glu	Tyr
Asn '		Asn	Ser	His	Asn 150	Val	Tyr	Ile	Met	Ala 155	Asp	Lys	Gln	Lys	Asn 160
145 Gly	Ile	Lys	Val	Asn 165		Lys	Ile	Arg	His 170		Ile	Glu	Asp	Gly 175	Ser
Val	Gln	Leu	Ala 180		His	Tyr	Gln	Gln 185	Asn	Thr	Pro	Ile	Gly 190	Asp	Gly
		195	Leu				His 200					205			
	210					215	Arg				220				
Val 225	Thr	Ala	Ala	Gly	Ile 230	Thr	Leu	Gly	Met	Asp 235	Glu	Leu	Tyr	Lys	Ser 240
Gly	Leu	Arg	Ser	Arg 245	Val	Thr	Met	Ala	Ala 250	Ala	Ala	Ala	Ala	Gly 255	Pro
Glu	Met	Val	Arg 260	Gly	Gln	Val	Phe	Asp 265	Val	Gly	Pro	Arg	Tyr 270	Thr	Asn
		275					Ala 280					285			
	290					295					300				
305					310		Arg			315					320
				325	,		Ile		330					335	
			340					345					350		Leu
		355	,				360					365			Asn
	370					375	<u>, </u>				380				Tyr
385					390)				395					1 Leu 400
				405	5				410)				415	
			420)				425	5				430)	ı Tyr
		435	5				440)				445	5		Lys
_	450					459	5				460)			ı Ala
465					470)				475	5				1 Asp 480
				48	5				490	0				49	
			500)				50	5				51	0	r Leu
		51	5				520	C				525	5		a Asp
	530)				53	5				54	0			o His
Lys 545		g Il	e Gli	u Va	1 Gli 55		n Al	a Le	u Al	a Hi 55		о Ту	r Le	u Gl	u Gln 560

(2) INFORMATION FOR SEQ ID NO:42:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2529 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...2526
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

ATG Met 1	GTG Val	AGC Ser	AAG Lys	GGC Gly 5	GAG Glu	GAG Glu	CTG Leu	TTC Phe	ACC Thr 10	GGG Gly	GTG Val	GTG Val	CCC Pro	ATC Ile 15	CTG Leu	48
GTC Val	GAG Glu	CTG Leu	GAC Asp 20	GGC Gly	GAC Asp	GTA Val	AAC Asn	GGC Gly 25	CAC His	AAG Lys	TTC Phe	AGC Ser	GTG Val 30	TCC Ser	GGC Gly	96
GAG Glu	GGC Gly	GAG Glu 35	GGC Gly	GAT Asp	GCC Ala	ACC Thr	TAC Tyr 40	GGC Gly	AAG Lys	CTG Leu	ACC Thr	CTG Leu 45	AAG Lys	TTC Phe	ATC Ile	144
TGC Cys	ACC Thr 50	ACC Thr	GGC Gly	AAG Lys	CTG Leu	CCC Pro 55	GTG Val	CCC Pro	TGG Trp	CCC Pro	ACC Thr 60	CTC Leu	GTG Val	ACC Thr	ACC Thr	192
CTG Leu 65	ACC Thr	TAC Tyr	GGC Gly	GTG Val	CAG Gln 70	TGC Cys	TTC Phe	AGC Ser	CGC Arg	TAC Tyr 75	CCC Pro	GAC Asp	CAC His	ATG Met	AAG Lys 80	240
CAG Gln	CAC His	GAC Asp	TTC Phe	TTC Phe 85	AAG Lys	TCC Ser	GCC Ala	ATG Met	CCC Pro 90	GAA Glu	GGC Gly	TAC Tyr	GTC Val	CAG Gln 95	GAG Glu	288
CGC	ACC Thr	ATC Ile	TTC Phe 100	Phe	AAG Lys	GAC Asp	GAC Asp	GGC Gly 105	Asn	TAC Tyr	AAG Lys	ACC Thr	CGC Arg 110	Ala	GAG Glu	336
GTG Val	AAG Lys	TTC Phe 115	Glu	GGC Gly	GAC Asp	ACC Thr	CTG Leu 120	Val	AAC Asn	CGC Arg	ATC	GAG Glu 125	Leu	AAG Lys	GGC Gly	384
ATC Ile	GAC Asp	TTC Phe	AAG Lys	GAC Glu	GAC Asp	GGC Gly	: AAC / Asr	ATC	CTG	GGC Gly	CAC His	AAG Lys	CTC	GAC 1 Glu	TAC Tyr	432

130 135 140

														AAG Lys		480
GGC Gly	ATC Ile	AAG Lys	GTG Val	AAC Asn 165	TTC Phe	AAG Lys	ATC Ile	CGC Arg	CAC His 170	AAC Asn	ATC Ile	GAG Glu	GAC Asp	GGC Gly 175	AGC Ser	528
GTG Val	CAG Gln	CTC Leu	GCC Ala 180	GAC Asp	CAC His	TAC Tyr	CAG Gln	CAG Gln 185	AAC Asn	ACC Thr	CCC Pro	ATC Ile	GGC Gly 190	GAC Asp	GGC Gly	576
CCC Pro	GTG Val	CTG Leu 195	CTG Leu	CCC Pro	GAC Asp	AAC Asn	CAC His 200	TAC Tyr	CTG Leu	AGC Ser	ACC Thr	CAG Gln 205	TCC Ser	GCC Ala	CTG Leu	624
AGC Ser	AAA Lys 210	GAC Asp	CCC Pro	AAC Asn	GAG Glu	AAG Lys 215	CGC Arg	GAT Asp	CAC His	ATG Met	GTC Val 220	Leu	CTG Leu	GAG Glu	TTC Phe	672
GTG Val 225	Thr	GCC Ala	GCC Ala	GGG Gly	ATC Ile 230	ACT Thr	CTC Leu	GGC Gly	ATG Met	GAC Asp 235	Glu	CTG Leu	TAC Tyr	AAG Lys	TCC Ser 240	720
GGA Gly	CTC Leu	AGA Arg	TCT Ser	CGA Arg 245	Ala	CAA Gln	GCT Ala	TCG Ser	AAT Asn 250	Ser	TCA Ser	ATC Met	GAG Glu	CTG Leu 255	GAA Glu	768
AAC Asn	ATC	GTG Val	GCC Ala 260	Asn	ACG Thr	GTC Val	TTG Leu	Leu 265	Lys	GCC Ala	AGC Arg	GA# g Glu	4 GGG 1 Gly 270	gly,	GGA Gly	816
GG? Gly	AAG / Lys	CGC Arg 275	Lys	GGG Gly	. Yya	AGC Ser	AAC Lys 280	Lys	TGG Trp	AAA Lys	A GAA s Glu	A ATO 1 Ile 289	e Lei	AAC 1 Lys	TTC ; Phe	864
CCI	CAC His	Ιle	AGC Ser	CAG Glr	TGT Cys	GAA Glu 295	Asp	CTC Lev	CGA Arg	ACC	G ACC Thi	r Ile	A GAG e Asi	E AGA	A GAT J Asp	912
TAC Ty: 305	c Cys	AG1 Ser	TTF Lev	TGT Cys	GAC GAS GAS GAS GAS GAS GAS GAS GAS GAS GAS	Lys	G CAC	G CCA	A ATC	GG(Gl ₂ 31:	y Arg	G CTY	G CT u Lei	r TT(u Phe	CGG Arg 320	960
CA(Gl:	TTI	r TGT ≥ Cys	r GAA	A ACC 1 Th: 325	: Arg	G CCI G Pro	r GG(G CTO	G GAC u Glu 330	ı Су:	T TAC	C AT	T CAG	G TTO n Pho 33	CTG e Leu 5	1008
GA(As)	TCC pSe:	C GT(G GCA 1 Ala 340	a Glu	A TAT	r GAJ r Glu	A GT' u Val	T AC' 1 Th: 34!	r Pro	A GA O As	T GA	a AA u Ly	A CT s Le 35	u Gl	A GAG y Glu	1056
AA Ly	A GG s Gl	G AAG y Ly: 35	s Gl	A AT'	T ATY	G ACC	C AA0 r Ly: 36	s Ту	C CTO	C AC u Th	C CC r Pr	A AA o Ly 36	s Se	c cc r Pr	T GTT o Val	1104

Phe									GTC Val							1152
									TTT Phe							1200
									TTC Phe 410							1248
									AAG Lys							1296
									CGA Arg							1344
TTC Phe	GGG Gly 450	GAG Glu	GTC Val	TGT Cys	GCC Ala	TGC Cys 455	CAG Gln	GTT Val	CGG Arg	GCC Ala	ACG Thr 460	GGT Gly	AAA Lys	ATG Met	TAT Tyr	1392
									ATC Ile		Lys					1440
TCC Ser	ATG Met	GCC Ala	CTC Leu	AAT Asn 485	GAG Glu	AAG Lys	CAG Gln	ATC	CTC Leu 490	GAG Glu	AAG Lys	GTC Val	AAC Asn	AGT Ser 495	Gln	1488
TTT Phe	GTG Val	GTC Val	AAC Asn 500	Leu	GCC Ala	TAT Tyr	GCC	TAC Tyr 505	GAG Glu	ACC Thr	AAG Lys	GAT Asp	GCA Ala 510	Leu	TGC Cys	1536
TTG Leu	GTC Val	CTG Leu 515	Thr	ATC	ATG Met	AAT Asn	GGG Gly 520	, Gla	GAC Asp	CTC Lev	AAC Lys	TTC Phe 525	His	ATC	TAC Tyr	1584
AAC Asn	Met	GGC Gly	/ Asn	CCT Pro	Gly	Phe	GAC Glu	GAG Glu	GAG Glu	CGC Arg	GCC g Ala 540	a Lev	TTT 1 Phe	г тал Э Туг	GCG Ala	1632
GCA Ala 545	Glu	ATC Ile	CTC Lev	TGC Cys	GGC Gly 550	Let	GAA Glu	A GAC 1 Asp	CTC Lev	CAC His	s Arg	r GAG g Glu	G AAC L Asi	C ACC	GTC Val 560	1680
ТАС Туг	CGA Arg	GAT JAS	r CTC D Leu	5 AAA 1 Lys 565	Pro	GAA	AAA Laa L	TATY	CTC E Leu 570	ı Lev	A GA' ı Ası	r GA' p As	Т ТА' р Ту:	r GGG r Gly 57!	C CAC y His	1728
ATI Ile	AGC Arc	F ATO	TCA Ser 580	Asp	CTC	GGC Gly	TTY Le	G GC' u Ala 585	a Val	AAG L Ly:	G AT	c cc e Pr	C GA o G1 59	u Gl	A GAC y Asp	1776
CTC Lev	ATC	CGG Arg	g Gly	C CGC	GTY Va:	G GGG	AC' Y Th	T GT r Va	r GG(l Gly	TA Ty	C AT r Me	G GC t Al	C CC a Pr	C GA o Gl	A GTC u Val	1824

		595					600					605				
CTG A	AAC Asn 610	AAC Asn	CAG Gln	AGG Arg	Tyr	GGC Gly 615	CTG Leu	AGC Ser	CCC Pro	Asp	TAC Tyr 620	TGG Trp	GGC Gly	CTT Leu	GGC Gly	1872
TGC (Cys : 625	CTC Leu	ATC Ile	ТАТ Тут	GAG Glu	ATG Met 630	ATC Ile	GAG Glu	GGC Gly	CAG Gln	TCG Ser 635	CCG Pro	TTC Phe	CGC Arg	GGC Gly	CGT Arg 640	1920
AAG (Lys (GAG Glu	AAG Lys	GTG Val	AAG Lys 645	CGG Arg	GAG Glu	GAG Glu	GTG Val	GAC Asp 650	CGC Arg	CGG Arg	GTC Val	CTG Leu	GAG Glu 655	ACG Thr	1968
GAG Glu	GAG Glu	GTG Val	TAC Tyr 660	TCC Ser	CAC His	AAG Lys	TTC Phe	TCC Ser 665	GAG Glu	GAG Glu	GCC Ala	AAG Lys	TCC Ser 670	ATC Ile	TGC Cys	2016
AAG Lys																2064
GAG Glu	GGG Gly 690	GCT Ala	GCA Ala	GAG Glu	GTC Val	AAG Lys 695	AGA Arg	CAC His	CCC Pro	TTC Phe	TTC Phe 700	AGG Arg	AAC Asn	ATG Met	AAC Asn	2112
TTC Phe 705	AAG Lys	CGC	TTA Leu	GAA Glu	GCC Ala 710	GGG Gly	ATG Met	TTG Leu	GAC Asp	CCT Pro 715	CCC Pro	TTC Phe	GTT Val	CCA Pro	GAC Asp 720	2160
CCC Fro	CGC Arg	GCT Ala	GTG Val	TAC Tyr 725	TGT Cys	AAG Lys	GAC Asp	GTG Val	CTG Leu 730	GAC Asp	ATC Ile	GAG Glu	CAG Gln	TTC Phe 735	TCC Ser	2208
ACT Thr	GTG Val	AAG Lys	GGC Gly 740	Val	AAT Asn	CTG Leu	GAC Asp	CAC His 745	ACA Thr	GAC Asp	GAC Asp	GAC Asp	TTC Phe 750	Tyr	TCC Ser	2256
AAG Lys	TTC Phe	TCC Ser 755	ACG Thr	GGC	TCT Ser	GTG Val	TCC Ser 760	Ile	CCA Pro	TGG Trp	CAA Gln	AAC Asn 765	Glu	ATG Met	ATA Ile	2304
GAA Glu	ACA Thr	Glu	TGC Cys	TTT Phe	AAG Lys	GAG Glu 775	Leu	AAC Asn	GTG Val	TTT Phe	780	Pro	AAT Asr	GGT Gly	ACC Thr	2352
CTC Leu 785	CCG	CCA Pro	. GAT Asp	CTG Leu	AAC Asn 790	Arc	AAC Asr	CAC His	CCT Pro	795	Gli	A CCG	CCC Pro	C AAC D Lys	AAA Lys 800	2400
GGG Gly	CTC	CTC	CAC Glr	AGA Arg 805	Let	TTC Phe	C AAC	G CGC	G CAC g Glr 810	ı His	Г CAC s Glr	G AAC n Asr	a AA' n As:	r TCC n Sei 819	AAG Lys	2448
AGT Ser	TC(G CCC	Sei 820	c Ser	AAC Lys	G ACC	C AG	T TT:	e Ası	C CAC	C CAC	C ATA	A AA: e As: 83	n Se:	A AAC r Asn	2496

2529

CAT GTC AGC TCG AAC TCC ACC GGA AGC AGC TAG
His Val Ser Ser Asn Ser Thr Gly Ser Ser
835

(2) INFORMATION FOR SEQ ID NO:43:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 842 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 20 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 60 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 75 70 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 Arg Thr Ile Fhe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 110 100 105 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 125 120 115 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 130 135 140 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 145 150 155 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 190 185 180 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 205 195 200 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 220 210 215 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 235 225 230 Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Ser Met Glu Leu Glu 250 245 Asn Ile Val Ala Asn Thr Val Leu Leu Lys Ala Arg Glu Gly Gly 260 265 Gly Lys Arg Lys Gly Lys Ser Lys Lys Trp Lys Glu Ile Leu Lys Phe 280 285 275 Pro His Ile Ser Gln Cys Glu Asp Leu Arg Arg Thr Ile Asp Arg Asp 300 295 Tyr Cys Ser Leu Cys Asp Lys Gln Pro Ile Gly Arg Leu Leu Phe Arg 315 310

Gln	Phe	Cys	Glu	Thr	Arg	Pro	Gly :		Glu 330	Cys	Tyr	Ile	Gln	Phe 335	Leu
Asp	Ser	Val	Ala 340	Glu	Tyr	Glu		Thr 345	Pro	Asp	Glu	Lys	Leu 350	Gly	Glu
		355			Met		360					365			
	370				Gly	375					380				
385					Cys 390					395					400
				405	Arg				410					415	
			420		Phe			425					430		
		435			Phe		440					445			
	450				Ala	455					460				
465					Glu 470					475					480
				485	Glu				490					495	
			500		Ala			505					510		
		515)				520					525			Tyr
	530)				535					540				Ala
545					550					555	•				Val 560
				565	,				570)				575	
			580)				585					590)	Asp
		595	5				600					605	,		val
	610)				615	,				620)			ı Gly
625	5				630)				63	5				Arg 640
				649	5				650	O				65	
			66	0				665					670)	e Cys
		67	5				680)				68	5		n Glu
	69	0				695	5				70	0			t Asn
70	5				710)				71	5				o Asp 720
Pr	o Ar			72	5				73	C				73	-
			74	0				74	5				75	0	r Ser
		75	5				760)				76	5		t Ile
Gl	u Th		u Cy	s Ph	e Ly	s Gl:		ı As	n Va	l Ph	e G1 78	y Pr O	o As	n Gl	y Thr

(2) INFORMATION FOR SEQ ID NO:44:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1902 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...1899
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

					GGG Gly			48
					AAG Lys			96
					CTG Leu			144
					CCC Pro			192
					TAC Tyr 75			240
					GAA Glu			288
					TAC Tyr			336
					CGC Arg			384

ATC Ile	GAC Asp 130	TTC Phe	AAG Lys	GAG Glu	GAC Asp	GGC . Gly . 135	AAC Asn	ATC Ile	CTG Leu	GGG Gly	CAC His 140	AAG Lys	CTG Leu	GAG Glu	TAC Tyr	432
														AAG Lys		480
														GGC Gly 175		528
GTG Val	CAG Gln	CTC Leu	GCC Ala 180	GAC Asp	CAC His	TAC Tyr	CAG Gln	CAG Gln 185	AAC Asn	ACC Thr	CCC Pro	ATC Ile	GGC Gly 190	GAC Asp	GGC Gly	576
			Leu											GCC Ala		624
AGC Ser	AAA Lys 210	GAC Asp	CCC Pro	AAC Asn	GAG Glu	AAG Lys 215	CGC Arg	GAT Asp	CAC His	ATG Met	GTC Val 220	CTG Leu	CTG	GAG Glu	TTC Phe	672
GTG Val 225	Thr	GCC Ala	GCC Ala	GGG Gly	ATC Ile 230	ACT Thr	CTC Leu	GGC Gly	ATG Met	GAC Asp 235	Glu	CTG Leu	ТАС	AAG Lys	TCC Ser 240	720
GGA Gly	CTC Leu	AGA Arg	TCT Ser	CGA Arg 245	Ala	CGA Arg	GCC Ala	ATC	ATG Met 250	Ser	AGA Arg	AGC Ser	Lys	G CGT Arg 255	GAC Asp	768
AAC Asn	AAT Asn	TTI Phe	TAT Tyr 260	Ser	GTA Val	GAG Glu	ATT	GGA Gly 265	Asp	TCI Ser	ACA Thr	TTC Phe	ACA Thi	val	CTG Leu	816
AAA Lys	CGA Arg	TAT Ty: 275	Glr	AAT Asn	TTA Leu	AAA Lys	CCT Pro 280	Ile	Gly GGC	TCA Sei	GI	A GCT Ala 285	a Gl	A GGA n Gly	A ATA	864
GTA Va]	TG0 Cys 290	Ala	a Ala	TAT Tyr	Asp	Ala	Ile	CTI Lev	GAA Glu	A AGA	AA A Isa g 300	ı Va	r GC. l Al	A ATO	AAG Lys	912
AAC Lys 305	s Leu	A AGO	C CGA	A CCA	TTT Phe	e Gln	AAT Asr	CAC n Glr	ACT Thi	r CA' r Hi: 31	s Ala	a Ly:	G CG s Ar	g GC0 g Ala	TAC Tyr 320	960
AG. Arg	GA(Glu	G CT	A GTT u Val	r CTM Let 325	ı Met	AAA Lys	TGI Cys	r GTT s Val	AA? L Asi 330	n Hi	C AAI s Ly:	A AA' s As:	T AT n Il	A AT e Il 33	r GGC e Gly 5	1008
CT'	T TT u Lev	AA E	T GT' n Vai	l Phe	C ACA	A CCA	A CAG	34!	s Se	C CT r Le	A GA u Gl	A GA u Gl	A TT u Ph 35	e Gl	A GAT n Asp	1056
GT Va	Т ТА(1 Ту:	C AT	A GTO	C ATK	G GAG	G CTO	ATO	G GA' t As	T GC. p Al	A AA a As	T CT n Le	т тс u Су	C CA	A GT n Va	G ATT l Ile	1104

360 365 355 CAG ATG GAG CTA GAT CAT GAA AGA ATG TCC TAC CTT CTC TAT CAG ATG 1152 Gln Met Glu Leu Asp His Glu Arg Met Ser Tyr Leu Leu Tyr Gln Met 370 CTG TGT GGA ATC AAG CAC CTT CAT TCT GCT GGA ATT ATT CAT CGG GAC 1200 Leu Cys Gly Ile Lys His Leu His Ser Ala Gly Ile Ile His Arg Asp 390 385 TTA AAG CCC AGT AAT ATA GTA GTA AAA TCT GAT TGC ACT TTG AAG ATT 1248 Leu Lys Pro Ser Asn Ile Val Val Lys Ser Asp Cys Thr Leu Lys Ile 410 CTT GAC TTC GGT CTG GCC AGG ACT GCA GGA ACG AGT TTT ATG ATG ACG 1296 Leu Asp Phe Gly Leu Ala Arg Thr Ala Gly Thr Ser Phe Met Met Thr 425 CCT TAT GTA GTG ACT CGC TAC TAC AGA GCA CCC GAG GTC ATC CTT GGC 1344 Pro Tyr Val Val Thr Arg Tyr Tyr Arg Ala Pro Glu Val Ile Leu Gly 440 1392 ATG GGC TAC AAG GAA AAC GTG GAT TTA TGG TCT GTG GGG TGC ATT ATG Met Gly Tyr Lys Glu Asn Val Asp Leu Trp Ser Val Gly Cys Ile Met 455 450 GGA GAA ATG GTT TGC CAC AAA ATC CTC TTT CCA GGA AGG GAC TAT ATT Gly Glu Met Val Cys His Lys Ile Leu Phe Pro Gly Arg Asp Tyr Ile 470 465 GAT CAG TGG AAT AAA GTT ATT GAA CAG CTT GGA ACA CCA TGT CCT GAA 1488 Asp Gln Trp Asn Lys Val Ile Glu Gln Leu Gly Thr Pro Cys Pro Glu 490 TTC ATG AAG AAA CTG CAA CCA ACA GTA AGG ACT TAC GTT GAA AAC AGA 1536 Phe Met Lys Lys Leu Gln Pro Thr Val Arg Thr Tyr Val Glu Asn Arg 500 505 CCT AAA TAT GCT GGA TAT AGC TTT GAG AAA CTC TTC CCT GAT GTC CTT 1584 Pro Lys Tyr Ala Gly Tyr Ser Phe Glu Lys Leu Phe Pro Asp Val Leu 520 515 TTC CCA GCT GAC TCA GAA CAC AAA CTT AAA GCC AGT CAG GCA AGG 1632 Phe Pro Ala Asp Ser Glu His Asn Lys Leu Lys Ala Ser Gln Ala Arg 535 530 GAT TTG TTA TCC AAA ATG CTG GTA ATA GAT GCA TCT AAA AGG ATC TCT 1680 Asp Leu Leu Ser Lys Met Leu Val Ile Asp Ala Ser Lys Arg Ile Ser 555 545 550 1728 GTA GAT GAA GCT CTC CAA CAC CCG TAC ATC AAT GTC TGG TAT GAT CCT Val Asp Glu Ala Leu Gln His Pro Tyr Ile Asn Val Trp Tyr Asp Pro 570

TCT GAA GCA GAA GCT CCA CCA CCA AAG ATC CCT GAC AAG CAG TTA GAT Ser Glu Ala Glu Ala Pro Pro Pro Lys Ile Pro Asp Lys Gln Leu Asp

585

580

GAA AGG GAA CAC ACA ATA GAA GAG TGG AAA GAA TTG ATA TAT AAG GAA
Glu Arg Glu His Thr Ile Glu Glu Trp Lys Glu Leu Ile Tyr Lys Glu
595 600 605

GTT ATG GAC TTG GAG GAG AGA ACC AAG AAT GGA GTT ATA CGG GGG CAG
Val Met Asp Leu Glu Glu Arg Thr Lys Asn Gly Val Ile Arg Gly Gln
610 615 620

CCC TCT CCT TTA GCA CAG GTG CAG CAG TGA
18902

(2) INFORMATION FOR SEQ ID NO:45:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 633 amino acids

630

- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 1 5 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 45 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 60 55 Leu Thr Tyr Gly Val Gln Cys Fne Ser Arg Tyr Pro Asp His Met Lys 75 70 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 110 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 120 125 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 130 135 140 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 155 150 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 170 165 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 185 180 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 195 200 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 210 215 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 225 230 235 240 Gly Leu Arg Ser Arg Ala Arg Ala Ile Met Ser Arg Ser Lys Arg Asp 245 250

Asn Asn Phe Tyr Ser Val Glu Ile Gly Asp Ser Thr Phe Thr Val Leu 265 260 Lys Arg Tyr Gln Asn Leu Lys Pro Ile Gly Ser Gly Ala Gln Gly Ile 275 280 Val Cys Ala Ala Tyr Asp Ala Ile Leu Glu Arg Asn Val Ala Ile Lys 290 295 300 Lys Leu Ser Arg Pro Phe Gln Asn Gln Thr His Ala Lys Arg Ala Tyr 305 310 315 320 Arg Glu Leu Val Leu Met Lys Cys Val Asn His Lys Asn Ile Ile Gly 325 330 Leu Leu Asn Val Phe Thr Pro Gln Lys Ser Leu Glu Glu Phe Gln Asp 345 340 Val Tyr Ile Val Met Glu Leu Met Asp Ala Asn Leu Cys Gln Val Ile 355 360 Gln Met Glu Leu Asp His Glu Arg Met Ser Tyr Leu Leu Tyr Gln Met 375 Leu Cys Gly Ile Lys His Leu His Ser Ala Gly Ile Ile His Arg Asp 390 395 . 400 Leu Lys Pro Ser Asn Ile Val Val Lys Ser Asp Cys Thr Leu Lys Ile 405 410 415 Leu Asp Phe Gly Leu Ala Arg Thr Ala Gly Thr Ser Phe Met Met Thr 430 425 420 Pro Tyr Val Val Thr Arg Tyr Tyr Arg Ala Pro Glu Val Ile Leu Gly 445 435 440 Met Gly Tyr Lys Glu Asn Val Asp Leu Trp Ser Val Gly Cys Ile Met 455 460 Gly Glu Met Val Cys His Lys Ile Leu Phe Pro Gly Arg Asp Tyr Ile 465 470 475 Asp Gln Trp Asn Lys Val Ile Glu Gln Leu Gly Thr Pro Cys Pro Glu 485 490 Phe Met Lys Lys Leu Gln Pro Thr Val Arg Thr Tyr Val Glu Asn Arg 505 510 500 Pro Lys Tyr Ala Gly Tyr Ser Phe Glu Lys Leu Phe Pro Asp Val Leu 515 520 Phe Pro Ala Asp Ser Glu His Asn Lys Leu Lys Ala Ser Gln Ala Arg 530 535 540 Asp Leu Leu Ser Lys Met Leu Val Ile Asp Ala Ser Lys Arg Ile Ser 545 550 555 Val Asp Glu Ala Leu Gln His Pro Tyr Ile Asn Val Trp Tyr Asp Pro 570 575 565 Ser Glu Ala Glu Ala Pro Pro Pro Lys Ile Pro Asp Lys Gln Leu Asp 585 580 Glu Arg Glu His Thr Ile Glu Glu Trp Lys Glu Leu Ile Tyr Lys Glu 595 600 605 Val Met Asp Leu Glu Glu Arg Thr Lys Asn Gly Val Ile Arg Gly Gln 620 610 615 Pro Ser Pro Leu Ala Gln Val Gln Gln 630

(2) INFORMATION FOR SEQ ID NO:46:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1824 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence (B) LOCATION: 1...1821

(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

ATG Met 1	GTG Val	AGC Ser	AAG Lys	GGC Gly 5	GAG Glu	GAG Glu	CTG Leu	TTC Phe	ACC Thr 10	GGG Gly	GTG Val	GTG Val	CCC Pro	ATC Ile 15	CTG Leu	48
GTC Val	GAG Glu	CTG Leu	GAC Asp 20	GGC Gly	GAC Asp	GTA Val	AAC Asn	GGC Gly 25	CAC His	AAG Lys	TTC Phe	AGC Ser	GTG Val 30	TCC Ser	GGC Gly	96
GAG Glu	GGC Gly	GAG Glu 35	GGC Gly	GAT Asp	GCC Ala	ACC Thr	ТАС Туг 40	GGC Gly	AAG Lys	CTG Leu	ACC Thr	CTG Leu 45	AAG Lys	TTC Phe	ATC Ile	144
TGC Cys	ACC Thr 50	ACC Thr	GGC Gly	AAG Lys	CTG Leu	CCC Pro 55	GTG Val	CCC Pro	TGG Trp	CCC Pro	ACC Thr 60	CTC Leu	GTG Val	ACC Thr	ACC Thr	192
CTG Leu 65	ACC Thr	TAC Tyr	GGC Gly	GTG Val	CAG Gln 70	TGC Cys	TTC Phe	AGC Ser	CGC Arg	TAC Tyr 75	CCC Pro	GAC Asp	CAC	ATG Met	AAG Lys 80	240
CAG Gln	CAC His	GAC Asp	TTC Phe	TTC Phe 85	AAG Lys	TCC Ser	GCC Ala	ATG Met	CCC Pro 90	GAA Glu	GGC Gly	TAC Tyr	GTC Val	CAG Gln 95	GAG Glu	288
CGC Arg	ACC Thr	ATC	TTC Phe 100	Phe	AAG Lys	GAC Asp	GAC Asp	GGC Gly 105	Asn	TAC Tyr	AAG Lys	ACC Thr	CGC Arg 110	Ala	GAG Glu	336
GTG Val	AAG Lys	TTC Phe	Glu	GGC Gly	GAC Asp	ACC Thr	CTG Leu 120	Val	AAC Asn	CGC Arg	ATC Ile	GAG Glu 125	Leu	AAG Lys	GGC Gly	384
ATC Ile	GAC Asp 130	Phe	AAC Lys	G GAC	GAC Asp	GGC Gly 135	Asr	: ATC	CTG Leu	GGG Gly	CAC His	Lys	CTC Lev	GAC Glu	TAC Tyr	432
AAC Asn 145	Туг	AAC Asr	AGC N Sei	CAC His	AAC Asr 150	ı Val	TAT Tyi	T ATC	ATC Met	GCC Ala 155	a Asp	C AAC	G CAC	AAC Lys	AAC ASN 160	480
GGC Gly	ATC	AAC e Ly:	G GTG S Va	AAG l Asi 16	n Phe	AAC Lys	TA F	e Arg	CAC His	s Ası	n Ile	C GAG e Glu	G GAG	GGG Gl ₁ 17	C AGC y Ser S	528
GTC Va]	G CAG	G CTO	C GC0 u Ala 18	a Ası	C CAG	С ТАС 5 Туі	CAC	G CAC n Gli 185	n Ası	n Th	c cc	C ATY	GG(e Gl;	y As	c GGC p Gly	576
CC(GT Va	G CT	G CT u Le	G CC	C GAG	C AAG p Asi	C CA	C TAC s Ty:	CTO r Le	G AG u Se	C AC r Th	C CA r Gl	G TC n Se	C GC r Al	C CTG a Leu	624

195 200 205

AGC Ser	AAA Lys 210	GAC Asp	CCC Pro	AAC Asn	GAG Glu	AAG Lys 215	CGC Arg	GAT Asp	CAC His	ATG Met	GTC Val 220	CTG Leu	CTG Leu	GAG Glu	TTC Phe	672
GTG Val 225	ACC Thr	GCC Ala	GCC Ala	GGG Gly	ATC Ile 230	ACT Thr	CTC Leu	GGC Gly	ATG Met	GAC Asp 235	GAG Glu	CTG Leu	TAC Tyr	AAG Lys	TCC Ser 240	720
														TTC Phe 255		768
CGG Arg	CAG Gln	GAG Glu	CTG Leu 260	AAC Asn	AAG Lys	ACA Thr	ATC Ile	TG3 Trp 265	GAG Glu	GTG Val	CCC Pro	GAG Glu	CGT Arg 270	TAC Tyr	CAG Gln	816
														GCT Ala		864
TTT Phe	GAC Asp 290	ACA Thr	AAA Lys	ACG Thr	GGG Gly	TTA Leu 295	CGT Arg	GTG Val	GCA Ala	GTG Val	AAG Lys 300	AAG Lys	CTC	TCC Ser	AGA Arg	912
														CTG Leu		960
					Lys									GAC Asp 335		1008
				Arg					Phe					CTG Leu		1056
ACC Thr	CAT His	CTC Leu 355	Met	GGG Gly	GCA Ala	GAT Asp	CTG Leu 360	Asn	AAC Asn	ATT	GTG Val	AAA Lys 365	Cys	CAG Gln	AAG Lys	1104
CTT Leu	ACA Thr 370	Asp	'GAC	CAT His	GTT Val	CAG Gln 375	Phe	CTI Leu	ATC	TAC Tyr	CAA Glr 380	ıle	CTC	CGA Arg	GGT Gly	1152
CTA Leu 385	ιLys	TAT Tyr	T ATA	CAT His	TCA Ser 390	Ala	GAC	TATA	ATT	CAC His	Arg	GAC J Asp	CT?	A AAA 1 Lys	CCT Pro 400	1200
AGT Ser	TAA T	CTA	A GCT 1 Ala	GTC Val 405	. Asr	r GAA 1 Glu	GAC Asp	TGI Cys	GAC Glu 410	Lei	3 AAC 1 Lys	ATT	r CTG	G GAT u Asr 415	TTT Phe	1248
GG <i>I</i>	A CIC	GC: 1 Ala	r CGC Arg 420	g His	C ACA	A GAT	GAT Asp	GAA Glu 425	ı Met	ACA Thi	A GGC	С ТАС У Тул	C GTV r Va 43	l Ala	ACT Thr	1296

AGG Arg	TGG Trp	TAC Tyr 435	AGG Arg	GCT Ala	CCT Pro	GAG Glu	ATC Ile 440	ATG Met	CTG Leu	AAC Asn	TGG Trp	ATG Met 445	CAT His	TAC Tyr	AAC Asn	1344
			GAT Asp													1392
ACT Thr 465	GGA Gly	AGA Arg	ACA Thr	TTG Leu	TTT Phe 470	CCT Pro	GGT Gly	ACA Thr	GAC Asp	CAT His 475	ATT Ile	GAT Asp	CAG Gln	TTG Leu	AAG Lys 480	1440
CTC Leu	ATT Ile	TTA Leu	AGA Arg	CTC Leu 485	GTT Val	GGA Gly	ACC Thr	CCA Pro	GGG Gly 490	GCT Ala	GAG Glu	CTT Leu	TTG Leu	AAG Lys 495	AAA Lys	1488
ATC Ile	TCC Ser	TCA Ser	GAG Glu 500	TCT Ser	GCA Ala	AGA Arg	AAC Asn	ТАТ Туг 505	ATT	CAG Gln	TCT Ser	TTG Leu	ACT Thr 510	CAG Gln	ATG Met	1536
			Asn					Phe					Pro		GCT	1584
		Leu					Leu					Asp			ATT	1632
ACA Thr 545	Ala	GCC	CAA Gln	GCC Ala	CTT Leu 550	Ala	CAT	GCC Ala	TAC Tyr	TT1 Phe 555	Ala	CAC Glr	TAC Tyr	CAC His	GAT Asp 560	1680
					Val					Asp					AGC Ser	1728
				Ile					s Ser					Glu	A GTC ı Val	1776
ATC Ile	AGC Ser	TT7	e Val	Pro	CCF Pro	A CCC	CT: Lev 600	ı Ası	CAA o Glr	A GAZ n Glu	A GAO	ATY Met 60!	t Glu	TCC Ser	C TGA	1824

(2) INFORMATION FOR SEQ ID NO:47:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 607 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:

Met 1	Val	Ser	Lys	Gly 5	Glu	Glu	Leu		Thr 10	Gly	Val	Val	Pro	Ile 15	Leu
Val	Glu	Leu	Asp 20	Gly	Asp	Val		Gly 25	His	Lys	Phe	Ser	Val 30	Ser	Gly
Glu	Gly	Glu 35	Gly	Asp	Ala	Thr	Tyr 40	Gly	Lys	Leu	Thr	Leu 45	Lys	Phe	Ile
Cys	Thr 50	Thr	Gly	Lys	Leu	Pro 55	Val	Pro	Trp	Pro	Thr 60	Leu	Val	Thr	Thr
Leu 65	Thr	Tyr	Gly	Val	Gln 70	Cys	Phe	Ser	Arg	Tyr 75	Pro	Asp	His	Met	Lys 80
Gln				85	Lys				90					95	
_			100		Lys			105					110		
		115			Asp		120					125			
	130				Asp	135					140				
145					Asn 150					155					160
_				165	Phe				170					175	
			180		His			185					190		
		195			Asp		200					205			
	210)				215					220				Phe
225					230					235	,				240
				245					250)				255	
			260)				265					270)	Gln
		275	5				280					285	5		Ala
	290)				295					300)			Arg
305	5				310					31	5				320
				325	5				330	C				33	
			340)				345	5				350	J	u Val
		35	5				360)				36	5		n Lys
	37	0				375	5				386	3			g Gly
3.81	5				390)				39	5				s Pro 400
				40	5				41	0				41	
			42	0				42	5				43	0	a Thr
		43	5				440)				44	5		r Asn
Gl	n Th 45		l As	p Il	e Trj	9 Se: 45		ı Gl	у Су	s 11	.е ме 46	0 C A1	a Gl	u ⊾€	u Leu

Thr	Gly	Arg	Thr	Leu	Phe 470	Pro	Gly	Thr	Asp	His 475	Ile	Asp	Gln	Leu	Lys 480
Leu	Ile	Leu	Arg	Leu 485	Val	Gly	Thr	Pro	Gly 4 90	Ala	Glu	Leu	Leu	Lys 495	Lys
			500					505		Gln			510		
Pro	Lys	Met 515	Asn	Phe	Ala	Asn	Val 520	Phe	Ile	Gly	Ala	Asn 525	Pro	Leu	Ala
Val	Asp 530	Leu	Leu	Glu	Lys	Met 535	Leu	Val	Leu	Asp	Ser 540	Asp	Lys	Arg	Ile
Thr 545	Ala	Ala	Gln	Ala	Leu 550	Ala	His	Ala	Tyr	Phe 555	Ala	Gln	Tyr	His	Asp 560
Pro	Asp	Asp	Glu	Pro 565	Val	Ala	Asp	Pro	Tyr 570	Asp	Gln	Ser	Phe	Glu 575	Ser
Arg	Asp	Leu	Leu 580	Ile	Asp	Glu	Trp	Lys 585	Ser	Leu	Thr	Tyr	Asp 590	Glu	Val
Ile	Ser	Phe 595	Val	Pro	Pro	Pro	Leu 600	Asp	Gln	Glu	Glu	Met 605		Ser	
		(2) IN	FORM	OITA	n fo	R SE	Q ID	NO:	48:					
	(i) S	EQUE	NCE	CHAR	ACTE	RIST	ICS:							
	·			GTH:											
		(B)	TYP	E: n	ucle	ic a	cid								
		(C)	STR	ANDE	DNES	S:s	ingl	e							
		(D)	TOF	OLOG	Y: 1	inea	r								

- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...2904
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

ATG Met 1	GTG Val	AGC Ser	AAG Lys	GGC Gly 5	GAG Glu	GAG Glu	CTG Leu	TTC Phe	ACC Thr 10	GGG Gly	GTG Val	GTG Val	CCC Pro	ATC Ile 15	CTG Leu		48
GTC Val	GAG Glu	CTG Leu	GAC Asp 20	GGC Gly	GAC Asp	GTA Val	AAC Asn	GGC Gly 25	CAC His	AAG Lys	TTC Phe	AGC Ser	GTG Val 30	TCC Ser	GGC Gly		96
GAG Glu	GGC Gly	GAG Glu 35	GGC Gly	GAT Asp	GCC Ala	ACC Thr	TAC Tyr 40	GGC Gly	AAG Lys	CTG Leu	ACC Thr	CTG Leu 45	AAG Lys	TTC Phe	ATC Ile	1	44
TGC Cys	ACC Thr 50	ACC Thr	GGC Gly	AAG Lys	CTG Leu	CCC Pro 55	GTG Val	CCC Pro	TGG Trp	CCC Pro	ACC Thr 60	CTC Leu	GTG Val	ACC Thr	ACC Thr	1	192
CTG Leu 65	ACC Thr	TAC Tyr	GGC Gly	GTG Val	CAG Gln 70	TGC Cys	TTC Phe	AGC Ser	CGC Arg	TAC Tyr 75	CCC Pro	GAC Asp	CAC His	ATG Met	AAG Lys 80	Ź	240
CAG Gln	CAC His	GAC Asp	TTC Phe	TTC Phe	AAG Lys	TCC Ser	GCC Ala	ATG Met	CCC Pro	GAA Glu	GGC Gly	TAC Tyr	GTC Val	CAG Gln	GAG Glu	:	288

85 90 95

CGC Arg !	acc Thr	ATC Ile	TTC Phe 100	TTC Phe	AAG Lys	GAC Asp	GAC Asp	GGC Gly 105	AAC Asn	TAC Tyr	AAG Lys	ACC Thr	CGC Arg 110	GCC Ala	GAG Glu	336
GTG . Val :	AAG Lys	TTC Phe 115	GAG Glu	GGC Gly	GAC Asp	ACC Thr	CTG Leu 120	GTG Val	AAC Asn	CGC Arg	ATC Ile	GAG Glu 125	CTG Leu	AAG Lys	GGC Gly	384
Ile	GAC Asp 130	TTC Phe	AAG Lys	GAG Glu	GAC Asp	GGC Gly 135	AAC Asn	ATC Ile	CTG Leu	GGG Gly	CAC His 140	AAG Lys	CTG Leu	GAG Glu	TAC Tyr	432
AAC Asn 145	TAC Tyr	AAC Asn	AGC Ser	CAC His	AAC Asn 150	GTC Val	TAT Tyr	ATC Ile	ATG Met	GCC Ala 155	GAC Asp	AAG Lys	CAG Gln	AAG Lys	AAC Asn 160	480
GGC Gly	ATC Ile	AAG Lys	GTG Val	AAC Asn 165	Phe	AAG Lys	ATC Ile	CGC Arg	CAC His 170	Asn	ATC Ile	GAG Glu	GAC Asp	GGC Gly 175	AGC Ser	528
GTG Val	CAG Gln	CTC Leu	GCC Ala 180	GAC Asp	CAC His	TAC Tyr	CAG Gln	CAG Gln 185	AAC Asn	ACC Thr	CCC	ATC Ile	GGC Gly 190	GAC Asp	GGC Gly	576
CCC Pro	GTG Val	CTG Leu 195	Leu	CCC Pro	GAC Asp	AAC Asn	CAC His	Tyr	CTG Leu	AGC Ser	ACC Thr	CAG Gln 205	Ser	GCC Ala	CTG Leu	624
AGC Ser	AAA Lys 210	Asp	CCC Pro	AAC Asn	GAG Glu	AAG Lys 215	Arg	GAT Jak	CAC His	: ATG : Met	GTC Val 220	Leu	CTG Leu	GAG Glu	TTC Phe	672
GTG Val 225	Thr	GCC Ala	GCC Ala	GGG Gly	3 ATC 7 Ile 230	Thr	CTC	GGC GGL	Met	GAC Asp 235	Glu	G CTC	TAC	AAC Lys	S TCC S Ser 240	720
GGA Gly	CTC	AGA	A TCI g Ser	Met 245	Ser	GCT Ala	GAC Glu	G GG(u Gl)	TAC Ty:	c Glr	TAC	C AGA	A GCC g Ala	CTC Lev 259	TAT 1 Tyr	768
GAT Asp	ТАТ ТУТ	r AAJ Lys	A AAC 5 Lys 260	Gli	A AGA u Arg	A GAA	ı Gl	A GAS u Asi 265	o Ile	e Ası	TTO Le	G CA:	TTX s Let 270	ı Gly	r GAC y Asp	816
ATA Ile	TTC	3 AC' 1 Th: 27	r Val	AA E	r AAJ n Lys	A GGO	g TC y Se 28	r Le	A GTA	A GC' 1 Ala	r CT a Le	T GG: u G1; 28	y Phe	C AG	T GAT r Asp	864
GGA Gly	CAC Gli 290	n Gl	A GCC u Ala	AGG Ar	G CC' g Pr	r GA. o G1 ⁻ 29	u Gl	A AT u Il	r GG e Gl	C TG(y Tr]	G TT p Le 30	u As	T GG n Gl	С ТА у Ту	T AAT r Asn	912
GAA Glu 305	ı Th	C AC r Th	A GGG r Gl	G GA y Gl	A AG u Ar 31	g Gl	G GA y As	C TT p Ph	T CC e Pr	G GG o G1 31	y Th	T TA	C GT r Va	A GA 1 Gl	A TAT u Tyr 320	960

								Pro	ACA Thr 330							1008
									TCG Ser							1056
									CTT Leu							1104
									AAG Lys							1152
									TAC Tyr							1200
									GAT Asp 410							1248
									TTG Leu					Lys		1296
			Asp					Val	ATT				Val		AGT Ser	1344
GAA Glu	ATG Met 450	Ile	TCT Ser	TTA Leu	GCT Ala	CCA Pro 455	GAA Glu	GTA Val	CAA Gln	AGC Ser	TCC Ser 460	Glu	GAA Glu	. ТАТ ГУТ	ATT lle	1392
	Leu					Ile					Ile				TAT Tyr 480	1440
					Tyr					Phe					CAA Gln	1488
AC(Thi	TCC Ser	AGC Sei	AAA Lys 500	Asr	CTC Lev	TTC Lev	AAT Asr	GCA n Ala 505	Arg	. GTA Val	A CTC	TC: Sei	GA: Gl: 510	ı Ile	r TTC e Phe	1536
			t Lei					c Ala) Asi		r GAA r Glu	1584
AA(As)	c CTC n Let 530	ı Il	a aal e Ly:	A GTT	TATA	A GAA e Glu 535	111	r TTX e Lei	A ATC	TC.	A ACT r Thi 540	r Gl	A TG	G AA' p As:	T GAA n Glu	1632
CG.	A CAG	G CC	T GCA	A CCA	A GCA	A CIX	CC'	T CC:	r AAA o Lys	A CC s Pr	A CC	A AA o Ly	A CC s Pr	T AC o Th	T ACT r Thr	1680

545					550					555					560	
								AAT Asn								1728
								GAA Glu 585								1776
								GTA Val								1824
								AGG Arg								1872
								AAA Lys								1920
ACC Thr	TTC Phe	AGT Ser	TCT Ser	GTG Val 645	GTT Val	GAA Glu	TTA Leu	ATA Ile	AAC Asn 650	CAC His	TAC Tyr	CGG Arg	AAT Asn	GAA Glu 655	TCT Ser	1968
				Asn				GAT Asp 665						Pro	GTA Val	2016
TCC Ser	ДДА Гус	TAC Tyr 675	Gln	CAG Gln	GAT Asp	CAA Gln	GTT Val 680	Va1	AAA Lys	GAA Glu	GAT Asp	AAT Asn 685	11ϵ	GAA Glu	GCT Ala	2064
GTA Val	GGG Gly 690	Lys	. AAA Lys	. TTA Leu	CAT His	GAA Glu 695	Tyr	AAC Asn	ACT Thr	CAG Gln	TTT Phe 700	Gln	GAP Glu	A AAA 1 Lys	AGT Ser	2112
CGA Arg 705	Glu	ТАТ Туг	GAT Asp	AGA Arg	TTA Leu 710	Tyr	GAA Glu	A GAA 1 Glu	тат Туг	ACC Thr 715	Arg	ACA Thr	TC(CAC r Glr	G GAA Glu 720	2160
ATC Ile	CAA Gln	ATC	AAA Lys	AGG Arg 725	Thr	GCT Ala	`ATT	r GAÆ ∋ Glu	GCA Ala 730	Phe	RAA ? Asr	GAJ Glu	A ACC	C ATA r Ile 735	A AAA e Lys	2208
ATA Ile	A TTI e Phe	GAZ Glu	GAZ Glu 740	ı Glr	TGC Cys	CAC Glr	ACC Thi	CAF Glr 745	Glu	CGC Arg	TAC Tyr	AG(Se)	2 AA 2 Ly 75	s Gl	A TAC 1 Tyr	2256
ATA Ile	A GAA e Glu	AA0 1 Lys 75!	s Phe	Γ AAA ≥ Lys	CGT Arg	GAZ Glu	GG(1 Gl ₂ 76	y Asr	r GAC n Glu	AA 1 Lys	A GAZ S Glu	A ATZ 1 110 769	e Gl	A AG n Ar	G ATT g Ile	2304
ATX Met	G CAT His 770	: Ası	r TAT	r GA'	r AAC o Lys	779 779	ı Ly	G TC' s Se:	r Arg	ATO	C AG' e Se: 78	r Gl	TA A	T AT e Il	T GAC e Asp	2352

AGT . Ser . 785	AGA Arg	AGA Arg	AGA Arg	TTG Leu	GAA Glu 790	GAA Glu	GAC Asp	TTG Leu	AAG Lys	AAG Lys 795	CAG Gln	GCA Ala	GCT Ala	GAG Glu	ТАТ Туг 800	2400
CGA Arg	GAA Glu	ATT Ile	GAC Asp	AAA Lys 805	CGT Arg	ATG Met	AAC Asn	AGC Ser	ATT Ile 810	AAA Lys	CCA Pro	GAC Asp	CTT Leu	ATC Ile 815	CAG Gln	2448
												ACT Thr				2496
GTT Val	CGG Arg	CAA Gln 835	AAG Lys	AAG Lys	TTG Leu	AAC Asn	GAG Glu 840	TGG Trp	TTG Leu	GGC Gly	AAT Asn	GAA Glu 845	AAC Asn	ACT Thr	GAA Glu	2544
GAC Asp	CAA Gln 850	TAT Tyr	TCA Ser	CTG Leu	GTG Val	GAA Glu 855	GAT Asp	GAT Asp	GAA Glu	GAT Asp	TTG Leu 860	CCC Pro	CAT His	CAT His	GAT Asp	2592
						Gly						AAA Lys				2640
CTG Leu	TTG Leu	CGA Arg	. GGG Gly	AAG Lys 885	Arg	GAT Asp	GGC Gly	ACT Thr	TTT Phe 890	Leu	GTC Val	CGG Arg	GAG Glu	AGC Ser 895	Ser	2688
AAA Lys	CAG Gln	GGC Gly	TGC Cys 900	Tyr	GCC Ala	TGC Cys	TCT Ser	GTA Val	. Val	GTC Val	GAC Asp	GGC Gly	GAA Glu 910	Val	AAG Lys	2736
CAT His	TGT Cys	GTC Val	Ile	AAC Asr	AAA Lys	ACA Thr	GCA Ala 920	Thr	Gly	TAT: TYT	GGC Gly	TTT Phe 925	Ala	GAC	CCC Pro	2784
тат туг	AAC Asn 930	Let	TAC Tyr	AGC Sei	TCT Ser	CTC Lev 935	ı Lys	A GAJ Glu	Lev	G GTC	G CTA Lev 940	ı His	TAC Tyr	CA#	A CAC n His	2832
ACC Thr 945	Ser	CTT Let	r GTC i Val	G CAC	G CAC 1 His 950	. Asr	GAC n Asp	TCC Sei	CTC	2 AA 1 Asi 959	n Vai	C ACA	A CTA	A GCC	TAC Tyr 960	2880
					G CAC n Glr				A							2907

(2) INFORMATION FOR SEQ ID NO:49:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 968 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:

1	Leu
Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phere	Gly
Cys Thr Thr Gly Lys Leu Pro Val Pro Thr Pro Thr Leu Val Thr Thr Leu Val Thr Ss Thr Leu Val Thr Thr Leu Val Thr Leu Thr Leu Pro Chr Fro Gu Asp His Met 65	Ile
Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met 65 - - 70 - - 75 -	Thr
Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Glr 90 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Leu Lys Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Lys Ilio Lys Phe Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu 130 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys 145	80
Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Leu Lys L	
115 120 125 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu 130 135 140 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys 145 150 155	
130 135 140 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys 145 150 155	
145 150 155	
	160
Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gl	5
Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly As 180 185 190 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Al	
195 200 205 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Gl	
210 215 220 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Ly	
225 230 235 Gly Leu Arg Ser Met Ser Ala Glu Gly Tyr Gln Tyr Arg Ala Le	240
245 250 250 Asp Tyr Lys Lys Glu Arg Glu Glu Asp Ile Asp Leu His Leu Gl	5
260 265 270 Ile Leu Thr Val Asn Lys Gly Ser Leu Val Ala Leu Gly Phe Se	
275 280 285 Gly Gln Glu Ala Arg Pro Glu Glu Ile Gly Trp Leu Asn Gly Ty	
290 295 300 Glu Thr Thr Gly Glu Arg Gly Asp Phe Pro Gly Thr Tyr Val Gl	
305 310 315 Ile Gly Arg Lys Lys Ile Ser Pro Pro Thr Pro Lys Pro Arg Pr	320
325 330 3. Arg Pro Leu Pro Val Ala Pro Gly Ser Ser Lys Thr Glu Ala As	35
340 345 350 Glu Gln Gln Ala Leu Thr Leu Pro Asp Leu Ala Glu Gln Phe A	
355 360 365 Pro Asp Ile Ala Pro Pro Leu Leu Ile Lys Leu Val Glu Ala I	le Glu
370 375 380 Lys Lys Gly Leu Glu Cys Ser Thr Leu Tyr Arg Thr Gln Ser S	er Ser
385 390 395 Asn Leu Ala Glu Leu Arg Gln Leu Leu Asp Cys Asp Thr Pro S	400 er Val 15
405 410 4 Asp Leu Glu Met Ile Asp Val His Val Leu Ala Asp Ala Phe L 420 425 430	

Tyr	Leu	Leu 435	Asp	Leu	Pro	Asn	Pro 440	Val	Ile	Pro		Ala 445	Val	Tyr	Ser
Glu	Met 450		Ser	Leu	Ala	Pro 455		Val	Gln	Ser	Ser 460	Glu	Glu	Tyr	Ile
Gln 465	Leu	Leu	Lys	Lys	Leu 470	Ile	Arg	Ser	Pro	Ser 475	Ile	Pro	His	Gln	Tyr 480
	Leu	Thr	Leu	Gln 485		Leu	Leu	Lys	His 490	Phe	Phe	Lys	Leu	Ser 495	Gln
Thr	Ser	Ser	Lys 500		Leu	Leu	Asn	Ala 505	-	Val	Leu	Ser	Glu 510	Ile	Phe
Ser	Pro	Met 515	Leu	Phe	Arg	Phe	Ser 520	Ala	Ala	Ser	Ser	Asp 525	Asn	Thr	Glu
Asn	Leu 530	Ile	Lys	Val	Ile	Glu 535	Ile	Leu	Ile	Ser	Thr 540	Glu	Trp	Asn	Glu
Arg 545	Gln	Pro	Ala	Pro	Ala 550	Leu	Pro	Pro	Lys	Pro 555	Pro	Lys	Pro	Thr	Thr 560
Val	Ala	Asn	Asn	Gly 565	Met	Asn	Asn	Asn	Met 570	Ser	Leu	Gln	Asn	Ala 575	Glu
-	Tyr		580					585					590		
_	Thr	595					600					605			
	Gly 610					615					620				
11e 625	Lys	Ile	Phe	His	Arg 630	Asp	Gly	Lys	Tyr	Gly 635	Phe	Ser	Asp	Pro	Leu 640
Thr	Phe	Ser	Ser	Val 645		Glu	Leu	Ile	Asn 650		Tyr	Arg	Asn	Glu 655	Ser
Leu	Ala	Gln	Tyr 660		Pro	Lys	Leu	Asp 665		Lys	Leu	Leu	Tyr 670		Val
Ser	Lys	Tyr 675	Gln		Asp	Gln	Val 680		Lys	Glu	Asp	Asn 685		Glu	Ala
Val	Gly 690	Lys		Leu	His	Glu 695		Asn	Thr	Gln	Phe 700	Gln	Glu	Lys	Ser
Arg 705	Glu		Asp	Arg	Leu 710		Glu	Glu	Тут	Thr 715		Thr	Ser	Gln	Glu 720
Ile	Gln	Met	. Lys	725		Ala	Ile	Glu	Ala 730		Asn	Glu	Thr	735	Lys
			740)				745					750)	Tyr
		755	5				760)				765	, ,		Ile
Met	His 770		тут	Asp	Lys	775		Ser	Arg	, Il∈	Ser 780	Glu	ı Ile	e Ile	Asp
785	5				790)				795					800
Arg	g Glu	Ile	e Asp	Lys 805		, Met	Asr	ı Ser	11e		Pro) Asp) Let	1 Ile 815	Gln
Leu	ı Arg	Lys	820		g Asp	Glr	туг	Leu 825		Trp	Leu	Thi	Glr 830		Gly
Va]	Arg	Glr 835	ı Lys		s Leu	ı Asr	n Glu 840) Let	ı Gly	/ Asr	1 Glu 849		n Thi	Glu
Asp	Glr 850	туз		r Lei	ı Val	G1v 855		a Asp	o Glu	ı Asp	Lev 860		o Hi	s His	s Asp
Glu 869	ı Lys		r Trj	p As:	n Val	Gly		r Sei	c Asi	n Arg 879		ı Ly:	s Al	a Gl	Asn 880
		ı Arç	g Gl	y Lys 88:	s Arg		o Gly	y Thi	r Phe 890		u Val	l Ar	g Gl	u Se: 89	r Ser 5

Lys Gln Gly Cys Tyr Ala Cys Ser Val Val Val Asp Gly Glu Val Lys 905 900 His Cys Val Ile Asn Lys Thr Ala Thr Gly Tyr Gly Phe Ala Glu Pro 925 920 Tyr Asn Leu Tyr Ser Ser Leu Lys Glu Leu Val Leu His Tyr Gln His 935 940 Thr Ser Leu Val Gln His Asn Asp Ser Leu Asn Val Thr Leu Ala Tyr 945 950 955 Pro Val Tyr Ala Gln Gln Arg Arg 965 (2) INFORMATION FOR SEQ ID NO:50: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2160 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (ix) FEATURE: (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...2157 (D) OTHER INFOFMATION: (xi) SEQUENCE DESCRIPTION: SEQ ID NO:50: ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG 48 Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 1 5 10 GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 20 GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC 144 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 35 TGC ACC ACC GGC AAG CTG CCC GTG CCC TGG CCC ACC CTC GTG ACC ACC 192 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 50 240 CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 65 CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 90 CGC ACC ATC TTC TTC AAG GAC GGC AAC TAC AAG ACC CGC GCC GAG 336 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 GTG AAG TTC GAG GGC GAC ACC CTG GTG AAC CGC ATC GAG CTG AAG GGC 384

Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly

ATC GAC TTC AAG GAG GAC GGC AAC ATC CTG GGG CAC AAG CTG GAG TAC Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr AAC TAC AAC AGC CAC AAC GTC TAT ATC ATG GCC GAC AAG CAG AAG AAC Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn GGC ATC AAG GTG AAC TTC AAG ATC CGC CAC AAC ATC GAG GAC GGC AGC Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser GTG CAG CTC GCC GAC CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly CCC GTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu AGC AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe GTG ACC GCC GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TCC Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser GGA CTC AGA TCT CGA GCT CAA GCT TCG AAT TCG ACC ATG TCG TCC ATC Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Thr Met Ser Ser Ile TTG CCA TTC ACG CCG CCA GTT GTG AAG AGA CTG CTG GGA TGG AAG AAG Leu Pro Phe Thr Pro Pro Val Val Lys Arg Leu Leu Gly Trp Lys Lys TCA GCT GGT GGG TCT GGA GGA GCA GGC GGA GGA GAG CAG AAT GGG CAG Ser Ala Gly Gly Ser Gly Gly Ala Gly Gly Glu Gln Asn Gly Gln CAA GAA AAG TGG TGT GAG AAA GCA GTG AAA AGT CTG GTG AAG AAG CTA Glu Glu Lys Trp Cys Glu Lys Ala Val Lys Ser Leu Val Lys Lys Leu AAG AAA ACA GGA CGA TTA GAT GAG CTT GAG AAA GCC ATC ACC ACT CAA Lys Lys Thr Gly Arg Leu Asp Glu Leu Glu Lys Ala Ile Thr Thr Gln AAC TGT AAT ACT AAA TGT GTT ACC ATA CCA AGC ACT TGC TCT GAA ATT Asn Cys Asn Thr Lys Cys Val Thr Ile Pro Ser Thr Cys Ser Glu Ile TGG GGA CTG AGT ACA CCA AAT ACG ATA GAT CAG TGG GAT ACA ACA GGC Trp Gly Leu Ser Thr Pro Asn Thr Ile Asp Gln Trp Asp Thr Thr Gly

CTT Leu	TAC Tyr	AGC Ser 355	TTC Phe	TCT Ser	GAA (Glu (Gln '	ACC Thr 360	AGG Arg	TCT Ser	CTT Leu	GAT Asp	GGT Gly 365	CGT Arg	CTC Leu	CAG Gln	1104
GTA Val	TCC Ser 370	CAT His	CGA Arg	AAA Lys	GGA Gly	TTG Leu 375	CCA Pro	CAT His	GTT Val	ATA Ile	TAT Tyr 380	TGC Cys	CGA Arg	TTA Leu	TGG Trp	1152
CGC Arg 385	TGG Trp	CCT Pro	GAT Asp	CTT Leu	CAC His 390	AGT Ser	CAT His	CAT His	GAA Glu	CTC Leu 395	AAG Lys	GCA Ala	ATT	GAA Glu	AAC Asn 400	1200
TGC Cys	GAA Glu	TAT Tyr	GCT Ala	TTT Phe 405	AAT Asn	CTT Leu	AAA Lys	AAG Lys	GAT Asp 410	GAA Glu	GTA Val	TGT Cys	GTA Val	AAC Asn 415	CCT Pro	1248
TAC Tyr	CAC His	TAT Tyr	CAG Gln 420	AGA Arg	GTT Val	GAG Glu	ACA Thr	CCA Pro 425	GTT Val	TTG Leu	CCT	CCA Pro	GTA Val 430	TTA Leu	GTG Val	1296
CCC Pro	CGA	CAC His 435	Thr	GAG Glu	ATC Ile	CTA Leu	ACA Thr 440	Glu	CTT Leu	CCG Pro	CCI Pro	CTG Leu 445	Asp	GAC Asp	ТАТ Туг	1344
ACT Thr	CAC His	Ser	ATT	CCA Pro	GAA Glu	AAC Asn 455	ACT Thr	' AAC ' Asn	TTC Phe	CCA Pro	GCA Ala 460	a Gly	ATT	GAG	CCA Pro	1392
CAG Gln 465	Ser	`AAT Asr	TAT Tyr	'ATT	CCA Pro 470	GAA Glu	ACG Thr	CCA Pro	CCT Pro	CCT Pro 475	Gl	ч ТАТ у Тут	T ATC	AGT Ser	GAA Glu 480	1440
GAT Asr	GGA Gly	. GA≱ ⁄ Glι	A ACA	AGT Ser 485	Asp	CAA Gln	CAG Glr	TTG	AAT Asr 490	ı Glr	A AGʻ n Se:	T ATY	GAC t Asp	ACA Thi 495	A GGC Gly	1488
TCT Ser	CCF Fro	A GCA	A GAA a Glu 500	ı Let	TCT Ser	CCT	ACT	r ACT Thr 505	Lev	r TCC	C CC r Pr	T GT' o Vai	T AA' l Ası 510	n His	r AGC s Ser	1536
TTC	G GA' J Asi	r TT Le 51	u Glr	G CCA	A GTI o Val	ACT Thr	туз	r TCA r Sei	Gli	u Pr	o Al	A TT a Ph 52	e Tr	G TG' p Cy	T TCA s Ser	1584
AT.	A GC. e Al. 53	а Ту	T TA' r Ty:	r GAI r Gli	A TTA 1 Leu	A AAT 1 Asr 535	Gli	g AGG n Arg	G GT' g Va	T GG 1 G1	A GA у G1 54	u Th	C TT r Ph	C CA e Hi	T GCA s Ala	1632
TC. Se 54	r Gl	G CC n Pr	C TC. o Se	A CTO	C ACT u Thi 550	val	A GA' L As	T GG(p Gl)	TT y Ph	T AC e Th 55	r As	C CC p Pr	A TC	A AA r As	T TCA n Ser 560	1680
GA Gl	G AG u Ar	G TT g Ph	C TG e Cy	C TT s Le 56	u Gl	r TT y Lei	A CT 1 Le	C TC	C AA r As 57	n Va	T AA	AC CC	SA AA ng As	T GC n Al 57	C ACG a Thr	1728
GT Va	A GA 1 G1	A AT u Me	G AC	A AG	A AG g Ar	G CA'	T AT s Il	A GG e Gl	A AG y Ar	A GO	GA GT Ly Va	rg co al Ar	G TI	TA TA	C TAC	1776

580 585 590

ATA GGT GGG GAA GTT TTT GCT GAG TGC CTA AGT GAT AGT GCA ATC TTT Ile Gly Gly Glu Val Phe Ala Glu Cys Leu Ser Asp Ser Ala Ile Phe 600 GTG CAG AGC CCC AAT TGT AAT CAG AGA TAT GGC TGG CAC CCT GCA ACA Val Gln Ser Pro Asn Cys Asn Gln Arg Tyr Gly Trp His Pro Ala Thr 615 620 GTG TGT AAA ATT CCA CCA GGC TGT AAT CTG AAG ATC TTC AAC AAC CAG 1920 Val Cys Lys Ile Pro Pro Gly Cys Asn Leu Lys Ile Phe Asn Asn Gln 630 635 GAA TTT GCT GCT CTG GCT CAG TCT GTT AAT CAG GGT TTT GAA GCC 1968 Glu Phe Ala Ala Leu Leu Ala Gln Ser Val Asn Gln Gly Phe Glu Ala 650 645 GTC TAT CAG CTA ACT AGA ATG TGC ACC ATA AGA ATG AGT TTT GTG AAA 2016 Val Tyr Gln Leu Thr Arg Met Cys Thr Ile Arg Met Ser Phe Val Lys 660 665 GGG TGG GGA GCA GAA TAC CGA AGG CAG ACG GTA ACA AGT ACT CCT TGC 2064 Gly Trp Gly Ala Glu Tyr Arg Arg Gln Thr Val Thr Ser Thr Pro Cys 680 TGG ATT GAA CTT CAT CTG AAT GGA CCT CTA CAG TGG TTG GAC AAA GTA 2112 Trp Ile Glu Leu His Leu Asn Gly Pro Leu Gln Trp Leu Asp Lys Val 700 2160 TTA ACT CAG ATG GGA TCC CCT TCA GTG CGT TGC TCA AGC ATG TCA TAA Leu Thr Gln Met Gly Ser Pro Ser Val Arg Cys Ser Ser Met Ser 715 710 705

(2) INFCPMATION FOR SEQ ID NO:51:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 719 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:

 Met
 Val
 Ser
 Lys
 Gly
 Glu
 Glu
 Leu
 Phe
 Thr
 Gly
 Val
 Val
 Leu
 Leu
 Leu
 Asp
 Glu
 Asp
 Val
 Asp
 Val
 Asp
 Val
 Asp
 Val
 Asp
 Val
 Asp
 Gly
 His
 Lys
 Phe
 Ser
 Val
 Ser
 Gly

 Glu
 Gly
 Gly
 Asp
 Ala
 Thr
 Tyr
 Gly
 Lys
 Leu
 Thr
 Leu
 Thr
 Thr
 Thr
 Leu
 Thr
 Thr

Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Thr Met Ser Ser Ile Leu Pro Phe Thr Pro Pro Val Val Lys Arg Leu Leu Gly Trp Lys Lys Ser Ala Gly Gly Ser Gly Gly Ala Gly Gly Gly Glu Gln Asn Gly Gln Glu Glu Lys Trp Cys Glu Lys Ala Val Lys Ser Leu Val Lys Lys Leu Lys Lys Thr Gly Arg Leu Asp Glu Leu Glu Lys Ala Ile Thr Thr Gln Asn Cys Asn Thr Lys Cys Val Thr Ile Pro Ser Thr Cys Ser Glu Ile Trp Gly Leu Ser Thr Pro Asn Thr Ile Asp Gln Trp Asp Thr Thr Gly Leu Tyr Ser Phe Ser Glu Gln Thr Arg Ser Leu Asp Gly Arg Leu Gln 355 360 Val Ser His Arg Lys Gly Leu Pro His Val Ile Tyr Cys Arg Leu Trp 370 375 Arg Trp Pro Asp Leu His Ser His His Glu Leu Lys Ala Ile Glu Asn Cys Glu Tyr Ala Phe Asn Leu Lys Lys Asp Glu Val Cys Val Asn Pro Tyr His Tyr Gln Arg Val Glu Thr Pro Val Leu Pro Pro Val Leu Val Pro Arg His Thr Glu Ile Leu Thr Glu Leu Pro Pro Leu Asp Asp Tyr Thr His Ser Ile Pro Glu Asn Thr Asn Phe Pro Ala Gly Ile Glu Pro Gln Ser Asn Tyr Ile Pro Glu Thr Pro Pro Pro Gly Tyr Ile Ser Glu Asp Gly Glu Thr Ser Asp Gln Gln Leu Asn Gln Ser Met Asp Thr Gly Ser Pro Ala Glu Leu Ser Pro Thr Thr Leu Ser Pro Val Asn His Ser Leu Asp Leu Gln Pro Val Thr Tyr Ser Glu Pro Ala Phe Trp Cys Ser Ile Ala Tyr Tyr Glu Leu Asn Gln Arg Val Gly Glu Thr Phe His Ala

545					550			Gly		555					560
	_			565				Ser	570					575	
Val	Glu	Met	Thr 580	Arg	Arg	His	Ile	Gly 585	Arg	Gly	Val	Arg	Leu 590	Tyr	Tyr
Ile	Gly	Gly 595	Glu	Val	Phe	Ala	Glu 600	Cys	Leu	Ser	Asp	Ser 605	Ala	Ile	Phe
Val	Gln 610	Ser	Pro	Asn	Суѕ	Asn 615	Gln	Arg	Tyr	Gly	Trp 620	His	Pro	Ala	Thr
Val 625	Суѕ	Lys	Ile	Pro	Pro 630	Gly	Суѕ	Asn	Leu	Lys 635	Ile	Phe	Asn	Asn	Gln 640
Glu	Phe	Ala	Ala	Leu 645	Leu	Ala	Gln	Ser	Val 650	Asn	Gln	Gly	Phe	Glu 655	Ala
	-		660					Thr 665					670		
Gly	Trp	Gly 675	Ala	Glu	Tyr	Arg	Arg 680	Gln	Thr	Val	Thr	Ser 685	Thr	Pro	Cys
Trp	Ile 690	Glu	Leu	His	Leu	Asn 695	Gly	Pro	Leu	Gln	Trp 700	Leu	Asp	Lys	Val
Leu 705		Gln	Met	Gly	Ser 710	Pro	Ser	Val	Arg	Cys 715		Ser	Met	Ser	

(2) INFORMATION FOR SEQ ID NO:52:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2421 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...2418
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:

ATG Met 1	GTG Val	AGC Ser	AAG Lys	GGC Gly 5	GAG Glu	GAG Glu	CTG Leu	TTC Phe	ACC Thr 10	GGG Gly	GTG Val	GTG Val	CCC Pro	ATC Ile 15	CTG Leu	48	}
GTC Val	GAG Glu	CTG Leu	GAC Asp 20	GGC Gly	GAC Asp	GTA Val	AAC Asn	GGC Gly 25	CAC His	AAG Lys	TTC Phe	AGC Ser	GTG Val 30	TCC Ser	GGC Gly	96	5
							TAC Tyr 40									144	1
							GTG Val									191	2
CTG Leu	ACC Thr	TAC Tyr	GGC Gly	GTG Val	CAG Gln	TGC Cys	TTC Phe	AGC Ser	CGC Arg	TAC Tyr	CCC Pro	GAC Asp	CAC His	ATG Met	AAG Lys	24	C

65	70	75	80
-*	he Lys Ser Ala M	ATG CCC GAA GGC TAC (let Pro Glu Gly Tyr \ 90	
	he Lys Asp Asp G	GGC AAC TAC AAG ACC (Gly Asn Tyr Lys Thr (
		GTG AAC CGC ATC GAG (/al Asn Arg Ile Glu : 125	
		ATC CTG GGG CAC AAG (le Leu Gly His Lys 1 140	
		ATC ATG GCC GAC AAG Ile Met Ala Asp Lys 155	
Gly Ile Lys Val A		CGC CAC AAC ATC GAG Arg His Asn Ile Glu 170	
	sp His Tyr Gln (CAG AAC ACC CCC ATC Gln Asn Thr Pro Ile 185	
-		TAC CTG AGC ACC CAG Tyr Leu Ser Thr Gln 205	
		GAT CAC ATG GTC CTG Asp His Met Val Leu 220	
		GGC ATG GAC GAG CTG Gly Met Asp Glu Leu 235	
Gly Leu Arg Ser A		TCG AAT TCG AAT TCA Ser Asn Ser Asn Ser 250	
	hr Asn Thr Pro	ACA AGT AAT GAT GCC Thr Ser Asn Asp Ala 265	
		AGA CAA GGT GGA GAG Arg Gln Gly Gly Glu 285	
		TTG GTA AAG AAG CTG Leu Val Lys Lys Leu 300	

AAA Lys 305	GAT Asp	GAA Glu	TTG Leu	GAT Asp	TCT Ser 310	TTA . Leu	ATA Ile	ACA Thr	GCT Ala	ATA Ile 315	ACT Thr	ACA Thr	TAA Asn	GGA Gly	A	СТ 1а 20	960
CAT His	CCT Pro	AGT Ser	AAA Lys	TGT Cys 325	GTT Val	ACC Thr	ATA Ile	CAG Gln	AGA Arg 330	ACA Thr	TTG Leu	GAT Asp	GGG Gly	AGG Arg 335	L	TT eu	1008
CAG Gln	GTG Val	GCT Ala	GGT Gly 340	CGG Arg	AAA Lys	GGA Gly	TTT Phe	CCT Pro 345	CAT His	GTG Val	ATC	TAT Tyr	GCC Ala 350	CGT	C C	eu .eu	1056
TGG Trp	AGG Arg	TGG Trp 355	CCT Pro	GAT Asp	CTT Leu	CAC His	AAA Lys 360	AAT Asn	GAA Glu	CTA Leu	AAA Lys	CAT His 365	GTT Val	AA/ Lys	r A	TAT Tyr	1104
TGT Cys	CAG Gln 370	Tyr	GCG Ala	TTT Phe	GAC Asp	TTA Leu 375	AAA Lys	TGT Cys	GAT Asp	AGT Ser	GTC Val 380	Cys	GTG Val	AA' Ası	r (CCA Pro	1152
тат Туг 385	His	TAC Tyr	GAA Glu	CGA Arg	GTT Val 390	GTA Val	TCA Ser	CCT Pro	GGA Gly	ATT Ile 395	Asp	CTC Leu	TCA Ser	GG;	λ 1	TTA Leu 400	1200
ACA Thr	CTG Leu	CAG Gln	AGT Ser	AAT Asn 405	GCT Ala	CCA Pro	TCA Ser	AGT Ser	ATG Met 410	Met	GTC Val	AAC Lys	GAT Asp	GA Gl 41	u '	TAT Tyr	1248
GTG Val	CAT His	GAC	TTT Phe 420	Glu	GGA Gly	CAG Gln	CCA Pro	TCG Ser 425	Leu	TCC Ser	Thi	r GAA	430	y Hi	T S	TCA Ser	1296
PTA 11	CAA Glr	A ACC Thi 435	Ile	CAG Gln	CAT	CCA Pro	CCA Pro 440	Ser	raa °	CGT Arg	r GCZ	A TCC a Set 44	r Thi	A GA r Gl	.u	ACA Thr	1344
ТАС Туз	AGC Ser 450	Thi	C CCA	A GCT	CTC	TTA Leu 455	Ala	CCA Pro	TCT Ser	GAG	G TC L Se: 46	r As	r GC n Al	T AC a Th	CC nr	AGC Ser	1392
ACT Thi 465	c Ala	C AAG	TTT	e Pro	AAC Asr 470	ATT n Ile	Pro	r GTC o Val	l Ala	a Se	r Th	A AG r Se	T CA r Gl	G CC n Pi	CT ro	GCC Ala 480	1440
AG' Se:	r ATA	e Le	G GGK u Gly	G GGG Y Gly 485	/ Sei	CAT r His	: AG	r GAZ r Glu	A GGI u G1: 49	y Le	G TT u Le	G CA u Gl	G AT n Il	e A	CA 1a 95	TCA Ser	1488
GG Gl	g CC' y Pr	T CA	G CC. n Pro 50	o Gl	A CAG	G CAC	G CAG	a AA' n Asi 50	n Gl	A TT y Ph	T AC e Th	T GG	т СА y Gl 51	n P	CA ro	GCT Ala	1536
AC T'n	т та r ту	C CA r Hi 51	s Hi	T AA s As:	C AG n Se	C AC'	r AC r Th 52	r Th	C TG r Tr	g AC p Th	T GO LT Gl	SA AC Ly Se 52	er Au	G A	CT hr	GCA Ala	1584
CC Pr	A TA O Ty	C AC	A CC	T AA o As	T TT n Le	G CC'	T CA o Hi	C CA s Hi	C CA s Gl	A AA n As	AC GC sn Gl	SC CA	AT C' is L	rr c eu G	AG In	CAC	1632

535 530 540 CAC CCG CCT ATG CCG CCC CAT CCC GGA CAT TAC TGG CCT GTT CAC AAT His Pro Pro Met Pro Pro His Pro Gly His Tyr Trp Pro Val His Asn 555 550 545 GAG CTT GCA TTC CAG CCT CCC ATT TCC AAT CAT CCT GCT CCT GAG TAT Glu Leu Ala Phe Gln Pro Pro Ile Ser Asn His Pro Ala Pro Glu Tyr 565 TGG TGT TCC ATT GCT TAC TTT GAA ATG GAT GTT CAG GTA GGA GAG ACA 1776 Trp Cys Ser Ile Ala Tyr Phe Glu Met Asp Val Gln Val Gly Glu Thr 585 TTT AAG GTT CCT TCA AGC TGC CCT ATT GTT ACT GTT GAT GGA TAC GTG 1824 Phe Lys Val Pro Ser Ser Cys Pro Ile Val Thr Val Asp Gly Tyr Val 595 600 GAC CCT TCT GGA GGA GAT CGC TTT TGT TTG GGT CAA CTC TCC AAT GTC 1872 Asp Pro Ser Gly Gly Asp Arg Phe Cys Leu Gly Gln Leu Ser Asn Val 615 610 CAC AGG ACA GAA GCC ATT GAG AGA GCA AGG TTG CAC ATA GGC AAA GGT 1920 His Arg Thr Glu Ala Ile Glu Arg Ala Arg Leu His Ile Gly Lys Gly 625 GTG CAG TTG GAA TGT AAA GGT GAA GGT GAT GTT TGG GTC AGG TGC CTT 1968 Val Gln Leu Glu Cys Lys Gly Glu Gly Asp Val Trp Val Arg Cys Leu 645 650 AGT GAC CAC GCG GTC TTT GTA CAG AGT TAC TAC TTA GAC AGA GAA GCT 2016 Ser Asp His Ala Val Phe Val Gln Ser Tyr Tyr Leu Asp Arg Glu Ala 665 660 GGG CGT GCA CCT GGA GAT GCT GTT CAT AAG ATC TAC CCA AGT GCA TAT 2064 Gly Arg Ala Pro Gly Asp Ala Val His Lys Ile Tyr Pro Ser Ala Tyr 680 ATA AAG GTC TTT GAT TTG CGT CAG TGT CAT CGA CAG ATG CAG CAG 2112 Ile Lys Val Phe Asp Leu Arg Gln Cys His Arg Gln Met Gln Gln Gln 690 695 GCG GCT ACT GCA CAA GCT GCA GCA GCT GCC CAG GCA GCA GCC GTG GCA 2160 Ala Ala Thr Ala Gln Ala Ala Ala Ala Ala Gln Ala Ala Ala Val Ala 715 GGA AAC ATC CCT GGC CCA GGA TCA GTA GGT GGA ATA GCT CCA GCT ATC 2208 Gly Asn Ile Pro Gly Pro Gly Ser Val Gly Gly Ile Ala Pro Ala Ile 730 AGT CTG TCA GCT GCT GGA ATT GGT GTT GAT GAC CTT CGT CGC TTA 2256 Ser Leu Ser Ala Ala Ala Gly Ile Gly Val Asp Asp Leu Arg Arg Leu 740 TGC ATA CTC AGG ATG AGT TTT GTG AAA GGC TGG GGA CCG GAT TAC CCA 2304 Cys Ile Leu Arg Met Ser Phe Val Lys Gly Trp Gly Pro Asp Tyr Pro

760

765

AGA CAG AGC ATC AAA GAA ACA CCT TGC TGG ATT GAA ATT CAC TTA CAC

Arg Gln Ser Ile Lys Glu Thr Pro Cys Trp Ile Glu Ile His Leu His

770 775 780

CGG GCC CTC CAG CTC CTA GAC GAA GTA CTT CAT ACC ATG CCG ATT GCA

Arg Ala Leu Gln Leu Leu Asp Glu Val Leu His Thr Met Pro Ile Ala

785 790 795 800

GAC CCA CAA CCT TTA GAC TGA

Asp Pro Gln Pro Leu Asp

(2) INFORMATION FOR SEQ ID NO:53:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 806 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:53:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 5 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 20 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 35 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 60 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 70 75 80 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 110 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 115 120 125 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 130 135 140 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 145 150 155 160 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 175 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 180 185 190 Fro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 195 200 205 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 210 215 220 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 225 230 235 240 Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Asn Ser Thr Met Asp 250 255

Asn	Met	Ser	Ile 260	Thr	Asn	Thr	Pro	Thr 265	Ser	Asn	Asp	Ala	Cys 270	Leu	Ser
		275	Ser				280					285		Glu	
	290					295					300			Glu	
Lys 305	Asp	Glu	Leu	Asp	Ser 310	Leu	Ile	Thr	Ala	Ile 315	Thr	Thr	Asn	Gly	Ala 320
His	Pro	Ser	Lys	Cys 325	Val	Thr	Ile	Gln	Arg 330	Thr	Leu	Asp	Gly	Arg 335	Leu
Gln	Va1	Ala	Gly 340	Arg	Lys	Gly	Phe	Pro 345	His	Val	Ile	Tyr	Ala 350	Arg	Leu
Trp	Arg	Trp 355	Pro	Asp	Leu	His	Lуs 360	Asn	Glu	Leu	Lys	His 365	Val	Lys	Tyr
	370					375					380			Asn	
385					390					395				Gly	400
				405					410					Glu 415	
			420					425					430		
		435					440					445		Glu	
	450					455					460			Thr	
465					470					475				Pro	480
				485					490					Ala 495	
			500					505					510		
	_	515					520					525		Thr	
	530					535					540			Gln	
545	·				550					555					Asn 560
				565	·				570)				575	
			580)				585	•				590)	Thr
		595	5				600)				605)		· Val
	610)				615	5				620)			Val
62!	5				630)				635	•				640
				645	5				650)				655	
			660)				665	5				670	0	ı Ala
		675	5				680)				685	5		a Tyr
	69	С				695	5				700)			n Gln
A1. 70		a Th:	r Ala	a Gl	n Ala 710		a Ala	a Ala	a Ala	a Glr 71		a Al	a Al	a Vai	1 Ala 720

											_,	- 1 -	D	73-	* 3 -	
Gly A				725					730					735		
Ser I		•	740					745					750			
Cys 1		755					760					765				
Arg (31n :	Ser :	Ile	Lys		Thr 775	Pro	Cys	Trp	Ile	Glu 780	Ile	His	Leu	His	
Arg 2		Leu (Gln	Leu	Leu 790	Asp	Glu	Val	Leu	His 795	Thr	Met	Pro	Ile	Ala 800	
Asp 1	Pro (Gln		Leu 805	Asp											
		(2)	INF	CRMA	MOIT	FOF	SEÇ	Q ID	NO:5	54:						
	(i	(A) (B) (C)	LENG TYPE STRA	TH: E: nu ANDEI	HARA 3120 iclei ONESS	bas .c ac .s: s:	se pa cid ingle	airs								
	•	i) M x) F			TYPI	E: cl	ONA									
		(B)	LOO OTI	CATIO	ON: :	l : RMAT	3117 ION:	eque			5 A					
								: SE								
ATG Met 1	GTG Val	AGC Ser	AAG Lys	GGC Gly 5	GAG Glu	GAG Glu	CTG Leu	TTC Phe	ACC Thr 10	GJA	GTC Val	GTG Val	Pro	ATC D Ile 15	CTG Leu	48
GTC	GAG	CTG	GAC	GGC	GAC	GTA	AAC	GGC	CVC	AAG	TTO	AGC	GT	G TCC	GGC Gly	96
Val	GIU	Leu	20	GIÀ	ASP	Vai	ASII	25 25	nis	. Lys	, ,,,,		30		. 017	
GAG Glu	GGC Gly	GAG Glu 35	GGC Gly	GAT Asp	GCC Ala	ACC Thr	TAC Tyr 40	GGC Gly	: AAC	CTC Lev	ACC Thi	C CTC Let 45	AA(G TTO S Pho	C ATC	144
TGC Cys	ACC Thr 50	ACC Thr	GGC Gly	AAG Lys	CTG Leu	CCC Pro	GTC Val	CCC Pro	TG(CCC Pro	C ACC Th:	CTY r Lei	C GTv ı Va	G AC	C ACC r Thr	192
CTG	ACC	TAC	GGC	GTG	CAG	TGC	TTC	: AGC	CG(TAC	CC	C GA	CA	C AT	G AAG	240
Leu 65	Thr	Tyr	Gly	Val	Gln 70	Cys	: Phe	e Ser	: Arq	тур 75	r Pr	o Ası	o Hi	s Me	t Lys 80	
CAG Gln	CAC His	GAC Asp	TTC Phe	TTC Phe 85	AAC Lys	TCC Sei	GCC Ala	ATC a Met	90	G GAI o Glu	e GG u Gl	С ТА У ТУ	C GT r Va	C CA 1 G1 95	G GAG n Glu	288
CGC Arg	ACC Thr	ATC Ile	TTC Phe	Ph∈	AAC Lys	GAC Asi	GA(GGG Gl ₂ 10!	y As	o Tag n Tyt	C AA r Ly	G AC s Th	C CG r Ar 11	g Al	C GAG a Glu	336

GTG AAG TTC GAG GGC GAC ACC CTG GTG AAC CGC ATC GAG CTG AAG GGC Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 115 120 125	384
ATC GAC TTC AAG GAG GAC GGC AAC ATC CTG GGG CAC AAG CTG GAG TAC Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 130 135 140	432
AAC TAC AAC AGC CAC AAC GTC TAT ATC ATG GCC GAC AAG CAG AAG AAC ASn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 145	480
GGC ATC AAG GTG AAC TTC AAG ATC CGC CAC AAC ATC GAG GAC GGC AGC Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165	528
GTG CAG CTC GCC GAC CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 180	576
CCC GTG CTG CTC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 195 200 205	624
AGC AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 210 215 220	672
GTG ACC GCC GCC GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TCC Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 235 230	720
GGA CTC AGA TCT ACC ATG GCG GGC TGG ATC CAG GCC CAG CAG CTG CAG Gly Leu Arg Ser Thr Met Ala Gly Trp Ile Gln Ala Gln Gln Leu Gln 245 250 255	768
GGA GAC GCG CTG CGC CAG ATG CAG GTG CTG TAC GGC CAG CAC TTC CCC Gly Asp Ala Leu Arg Gln Met Gln Val Leu Tyr Gly Gln His Phe Pro 260 265 270	816
ATC GAG GTC CGG CAC TAC TTG GCC CAG TGG ATT GAG AGC CAG CCA TGG Ile Glu Val Arg His Tyr Leu Ala Gln Trp Ile Glu Ser Gln Pro Trp 275 280 285	864
GAT GCC ATT GAC TTG GAC AAT CCC CAG GAC AGA GCC CAA GCC ACC CAG Asp Ala Ile Asp Leu Asp Asn Pro Gln Asp Arg Ala Gln Ala Thr Gln 290 295 300	912
CTC CTG GAG GGC CTG GTG CAG GAG CTG CAG AAG AAG GCG GAG CAC CAG Leu Leu Glu Gly Leu Val Gln Glu Leu Gln Lys Lys Ala Glu His Gln 305 310 315	960
GTG GGG GAA GAT GGG TTT TTA CTG AAG ATC AAG CTG GGG CAC TAC GCC Val Gly Glu Asp Gly Phe Leu Leu Lys Ile Lys Leu Gly His Tyr Ala 325	1008
ACG CAG CTC CAG AAA ACA TAT GAC CGC TGC CCC CTG GAG CTG GTC CGC Thr Gln Leu Gln Lys Thr Tyr Asp Arg Cys Pro Leu Glu Leu Val Arg	1056

TGC ATC CGG CAC ATT CTG TAC AAT GAA CAG AGG CTG GTC CGA GAA GCC Cys Ile Arg His Ile Leu Tyr Asn Glu Gln Arg Leu Val Arg Glu Ala AAC AAT TGC AGC TCT CCG GCT GGG ATC CTG GTT GAC GCC ATG TCC CAG Asn Asn Cys Ser Ser Pro Ala Gly Ile Leu Val Asp Ala Met Ser Gln AAG CAC CTT CAG ATC AAC CAG ACA TTT GAG GAG CTG CGA CTG GTC ACG Lys His Leu Gln Ile Asn Gln Thr Phe Glu Glu Leu Arg Leu Val Thr CAG GAC ACA GAG AAT GAG CTG AAG AAA CTG CAG CAG ACT CAG GAG TAC Gln Asp Thr Glu Asn Glu Leu Lys Lys Leu Gln Gln Thr Gln Glu Tyr TTC ATC ATC CAG TAC CAG GAG AGC CTG AGG ATC CAA GCT CAG TTT GCC Phe Ile Ile Gln Tyr Gln Glu Ser Leu Arg Ile Gln Ala Gln Phe Ala CAG CTG GCC CAG CTG AGC CCC CAG GAG CGT CTG AGC CGG GAG ACG GCC Gln Leu Ala Gln Leu Ser Pro Gln Glu Arg Leu Ser Arg Glu Thr Ala CTC CAG CAG AAG CAG GTG TCT CTG GAG GCC TGG TTG CAG CGT GAG GCA Leu Gln Gln Lys Gln Val Ser Leu Glu Ala Trp Leu Gln Arg Glu Ala CAG ACA CTG CAG CAG TAC CGC GTG GAG CTG GCC GAG AAG CAC CAG AAG Gln Thr Leu Gln Gln Tyr Arg Val Glu Leu Ala Glu Lys His Gln Lys ACC CTG CAG CTG CTG CGG AAG CAG CAG ACC ATC ATC CTG GAT GAC GAG Thr Leu Gln Leu Leu Arg Lys Gln Gln Thr Ile Ile Leu Asp Asp Glu CTG ATC CAG TGG AAG CGG CGG CAG CAG CTG GCC GGG AAC GGC GGG CCC Leu Ile Gln Trp Lys Arg Arg Gln Gln Leu Ala Gly Asn Gly Gly Pro CCC GAG GGC AGC CTG GAC GTG CTA CAG TCC TGG TGT GAG AAG TTG GCC Pro Glu Gly Ser Leu Asp Val Leu Gln Ser Trp Cys Glu Lys Leu Ala GAG ATC ATC TGG CAG AAC CGG CAG CAG ATC CGC AGG GCT GAG CAC CTC Glu Ile Ile Trp Gln Asn Arg Gln Gln Ile Arg Arg Ala Glu His Leu TGC CAG CAG CTG CCC ATC CCC GGC CCA GTG GAG GAG ATG CTG GCC GAG Cys Gln Gln Leu Pro Ile Pro Gly Pro Val Glu Glu Met Leu Ala Glu GTC AAC GCC ACC ATC ACG GAC ATT ATC TCA GCC CTG GTG ACC AGC ACA Val Asn Ala Thr Ile Thr Asp Ile Ile Ser Ala Leu Val Thr Ser Thr

TTC Phe	ATC Ile	ATT Ile	GAG Glu 580	AAG Lys	CAG Gln	CCT Pro	CCT Pro	CAG Gln 585	GTC Val	CTG Leu	AAG Lys	ACC Thr	CAG Gln 590	ACC Thr	AAG Lys	1776
TTT Phe	GCA Ala	GCC Ala 595	ACC Thr	GTA Val	CGC Arg	CTG Leu	CTG Leu 600	GTG Val	GGC Gly	GGG Gly	AAG Lys	CTG Leu 605	AAC Asn	GTG Val	CAC His	1824
ATG Met	AAT Asn 610	CCC Pro	CCC Pro	CAG Gln	GTG Val	AAG Lys 615	GCC Ala	ACC Thr	ATC Ile	ATC Ile	AGT Ser 620	GAG Glu	CAG Gln	CAG Gln	GCC Ala	1872
AAG Lys 625	TCT Ser	CTG Leu	CTT Leu	AAA Lys	AAT Asn 630	GAG Glu	AAC Asn	ACC Thr	CGC Arg	AAC Asn 635	GAG Glu	TGC Cys	AGT Ser	GGT Gly	GAG Glu 640	1920
ATC Ile	CTG Leu	AAC Asn	AAC Asn	TGC Cys 645	TGC Cys	GTG Val	ATG Met	GAG Glu	TAC Tyr 650	CAC His	CAA Gln	GCC Ala	ACG Thr	GGC Gly 655	ACC Thr	1968
CTC Leu	AGT Ser	GCC Ala	CAC His 660	TTC Phe	AGG Arg	AAC Asn	ATG Met	TCA Ser 665	CTG Leu	AAG Lys	AGG Arg	ATC Ile	AAG Lys 670	Arg	GCT Ala	2016
GAC Asp	CGG Arg	CGG Arg 675	Gly	GCA Ala	GAG Glu	TCC Ser	GTG Val 680	Thr	GAG Glu	GAG Glu	AAG Lys	TTC Phe 685	Thr	GTC Val	CTG Leu	2064
TTT Phe	GAG Glu 690	Ser	CAG Gln	TTC Phe	AGT Ser	GTT Val 695	GGC Gly	AGC Ser	AAT Asn	GAG Glu	CTT Leu 700	Val	TTC Phe	CAC Glr	GTG Val	2112
AAG Lys 705	Thr	CTC Leu	TCC Ser	CTA	CCT Pro 710	Val	GTT Val	GTC Val	Ile	GTC Val 715	His	Gly	: AGC / Sei	CAC Glr	GAC Asp 720	2160
CAC His	AAT Asn	GCC Ala	ACG Thr	GCT Ala 725	Thr	GTG Val	CTG Leu	TGG Trp	GAC Asp 730	Asr	GCC Ala	TTT Phe	r GCT e Ala	F GAG a Glu 73!	CCG Pro	2208
GGC	AGG Arg	GTC Val	G CCA 1 Pro 740	Phe	GCC Ala	GTG Val	Pro	GAC Asp 745	Lys	GTC Val	CTC Lev	TG(750 750	5 Gl	G CTG n Leu	2256
TGT Cys	GAC Glu	GC0 1 Ala 75	a Leu	AAC 12A L	ATC Met	AAA Lys	770 Phe 760	e Lys	GCC Ala	GAI Glu	A GTC 1 Val	G CA0 L Gli 76!	n Se	C AA r As	n Arg	2304
G17	CTC / Let 770	ı Th	C AAC	G GA(AAC LAST	775	ı Val	TTY Phe	CTC e Lei	GCC 1 Ala	G CAG a Glr 780	ı Ly	A CT s Le	G TT u Ph	c AAC e Asn	2352
AA0 Asi 78!	ı Sei	c AG r Se	C AGG	C CAC	C CTC s Lev 790	ı Glı	GA(С ТА(р Ту:	C AG:	r GG r Gl; 79	y Le	G TC u Se	C GT r Va	G TC 1 Se	C TGG r Trp 800	2400
TC(Se:	C CAG	3 TT n Ph	C AA(e As:	c AG n Ar	G GA(g Gl)	G AA(u Asi	TT Lei	g CCG u Pr	G GG(o Gl;	TG Y Tr	G AA p As:	C TA n Ty	C AC r Th	C TT ir Ph	C TGG le Trp	2448

		805		810		815	
CAG TGG	TTT GAC Phe Asp 820	GGG GTG Gly Val	ATG GAG Met Glu	GTG TTG Val Leu 825	AAG AAG Lys Lys	CAC CAC AAG His His Lys 830	CCC 2496 Pro
His Trp .	AAT GAT Asn Asp 835	GGG GCC Gly Ala	ATC CTA Ile Leu 840	GGT TTT Gly Phe	Val Asn	AAG CAA CAG Lys Gln Gln 845	GCC 2544 Ala
CAC GAC His Asp 850	CTG CTC Leu Leu	ATC AAC Ile Asn	AAG CCC Lys Pro 855	GAC GGG Asp Gly	ACC TTC Thr Phe 860	TTG TTG CGC Leu Leu Arg	TTT 2592 Phe
AGT GAC Ser Asp 865	TCA GAA Ser Glu	ATC GGG Ile Gly 870	GGC ATC Gly Ile	ACC ATC	GCC TGG Ala Trp 875	AAG TTT GAC Lys Phe Asp	TCC 2640 Ser 880
CCG GAA Pro Glu	CGC AAC Arg Asn	CTG TGG Leu Trp 885	AAC CTG Asn Leu	AAA CCA Lys Pro 890	Phe Thr	ACG CGG GAT Thr Arg Asp 895	Phe
TCC ATC Ser Ile	AGG TCC Arg Ser 900	Leu Ala	GAC CGG Asp Arg	CTG GGG Leu Gly 905	G GAC CTG y Asp Leu	AGC TAT CTC Ser Tyr Let 910	ATC 2736
TAT GTG Tyr Val	TTT CCT Phe Pro 915	GAC CGC Asp Arg	CCC AAG Pro Lys 920	Asp Gl	G GTC TTC u Val Phe	TCC AAG TAG Ser Lys Tyn 925	TAC 2784
ACT CCT Thr Pro 930	GTG CTC Val Let	GCT AAA 1 Ala Lys	A GCT GTT : Ala Val 935	T GAT GG. l Asp Gl	A TAT GTG y Tyr Val 940	AAA CCA CA Lys Pro Gl	S ATC 2832 n Ile
AAG CAA Lys Gln 945	GTG GTG Val Va	C CCT GAG 1 Pro Glu 950	ı Phe Vai	G AAT GC l Asn Al	A TCT GCA a Ser Ala 955	A GAT GCT GG A Asp Ala Gl	G GGC 2880 y Gly 960
AGC AGC Ser Ser	GCC AC	G TAC ATO r Tyr Me	G GAC CAC t Asp Gli	G GCC CC n Ala Pr 97	o Ser Pro	A GCT GTG TG D Ala Val Cy 97	s Pro
CAG GCT Gln Ala	CCC TA Pro Ty 98	r Asn Me	G TAC CC. t Tyr Pr	A CAG AA o Gln As 985	C CCT GAC	CAT GTA CT His Val Le 990	C GAT 2976 u Asp
CAG GAT Gln Asp	GGA GA Gly Gl 995	A TTC GA u Phe As	C CTG GA p Leu As 100	p Glu Th	CC ATG GA' nr Met As	T GTG GCC AC p Val Ala Ar 1005	G CAC 3024 g His
GTG GAG Val Glu 1010	ı Glu Le	C TTA CG eu Leu Ar	C CGA CC g Arg Pr 1015	A ATG GA O Met As	AC AGT CT sp Ser Le 102	T GAC TCC CC u Asp Ser Ai 0	GC CTC 3072 rg Leu
TCG CCC Ser Pro 1025	CCT GC	CC GGT CT la Gly Le 103	eu Phe Th	CC TCT GO	CC AGA GG la Arg Gl 1035	C TCC CTC T y Ser Leu S	CA TGA 3120 er 1

(2) INFORMATION FOR SEQ ID NO:55:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1039 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 20 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 60 55 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 75 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 105 100 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 125 115 120 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 140 135 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 150 155 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 170 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 185 190 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 200 205 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 220 215 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 235 230 Gly Leu Arg Ser Thr Met Ala Gly Trp Ile Gln Ala Gln Gln Leu Gln 245 250 Gly Asp Ala Leu Arg Gln Met Gln Val Leu Tyr Gly Gln His Phe Pro 265 Ile Glu Val Arg His Tyr Leu Ala Gln Trp Ile Glu Ser Gln Pro Trp 275 280 Asp Ala Ile Asp Leu Asp Asn Fro Gln Asp Arg Ala Gln Ala Thr Gln 295 Leu Leu Glu Gly Leu Val Gln Glu Leu Gln Lys Lys Ala Glu His Gln 315 310 Val Gly Glu Asp Gly Phe Leu Leu Lys Ile Lys Leu Gly His Tyr Ala 325 330 Thr Gln Leu Gln Lys Thr Tyr Asp Arg Cys Pro Leu Glu Leu Val Arg 345

Cys	Ile	Arg 355	His	Ile	Leu		Asn 360	Glu	Gln	Arg		Val 365	Arg	Glu	Ala
	370	Cys				Ala 375	Gly				380				
Lys	His	Leu	Gln	Ile	Asn	Gln	Thr	Phe	Glu	Glu	Leu	Arg	Leu	Val	Thr
385					390				_	395	~ 1.	m 1	G3-		400
				405	Glu				410					415	
			420		Gln			425					430		
		435			Ser		440					445			
Leu	Glr 450		Lys	Gln	Val	Ser 455	Leu	Glu	Ala	Trp	Leu 460	Gln	Arg	Glu	Ala
Gln 465	Thr	Lev	Gln	Gln	Tyr 470	Arg	Val	Glu	Leu	Ala 475		Lys	His	Gln	Lys 480
Thr	Let	ı Glr	Leu	Leu 485	Arg	Lys	Gln	Gln	Thr 490		Ile	Leu	Asp	Asp 495	Glu
Lev	ılle	e Glr	Trp 500	Lys	Arg	Arg	Gln	Gln 505	Leu	Ala	Gly	Asn	Gly 510	Gly	Pro
Pro	Gl:	Gly 515	/ Ser		Asp	Val	Leu 520	Gln	Ser	Trp	Cys	Glu 525	Lys	Leu	Ala
Glu	1 Ile 53	e Ile	Trp	Gln	Asn	Arg 535		Gln	Ile	Arg	Arg 540		Glu	His	Leu
CVS	s Gli	n Gli	ı Leu	Pro	Ile			Pro	Val	Glu	Glu	Met	Leu	Ala	Glu
549	5				550					555	•				560
Va.	l As	n Ala	a Thr	: Ile	Thr	Asp	Ile	Ile	Ser 570		Leu	. Val	Thr	Ser 575	Thr
			580	Lys	Gln			585					590)	Lys
		59	5				600)				605)		His
	61	0				615	5				620)			Ala
62	5				630	1				635	5				Glu 640
11	e Le	u As	n Ası	n Cys 649		Va]	l Met	Glu	ту: 650		s Glr	n Ala	a Thi	655 Gly	/ Thr
Le	u Se	r Al	a His		e Arg	Ası	n Met	Ser 665		ı Ly:	s Arg	g Ile	e Lys 670	s Arg O	g Ala
		67	5				680)				68	5		l Leu
	69	0				69	5				700	C			n Val
70	5				710)				71	5				n Asp 720
Hi	s As			72	5				73	0				73	
G]	y Au	g Va	al Pr 74		e Ala	a Va	l Pr	o As 74		s Va	l Le	u Tr	p Pr 75	o Gl 0	n Leu
		75	55				76	0				76	5		n Arg
	7	70				77	5				78	0			e Asn
78	35				79	0				79	95				r Trp 800
Se	er G	ln Pl	ne As	n Ar 80		u As	n Le	eu Pr		.у Ті .0	np As	η Τζ	r Th	r Ph 81	e Trp

Gln	Trp	Phe	Asp 820	Gly	Val	Met	Glu	Val 825	Leu	Lys	Lys	His	His 830	Lys	Pro	
His	Trp	Asn 835	Asp	Gly	Ala	Ile	Leu 840	Gly	Phe	Val	Asn	Lys 845	Gln	Gln	Ala	
His	Asp 850	Leu	Leu	Ile	Asn	Lys 855	Pro	Asp	Gly	Thr	Phe 860	Leu	Leu	Arg	Phe	
Ser 865	Asp	Ser	Glu	Ile	Gly 870	Gly	Ile	Thr	Ile	Ala 875	Trp	Lys	Phe	Asp	Ser 880	
Pro	Glu	Arg	Asn	Leu 885	Trp	Asn	Leu	Lys	Pro 890	Phe	Thr	Thr	Arg	Asp 895	Phe	
Ser	Ile	Arg	Ser 900	Leu	Ala	Asp	Arg	Leu 905	Gly	Asp	Leu	Ser	Tyr 910	Leu	Ile	
Tyr	Val	Phe 915	Pro	Asp	Arg	Pro	Lys 920	Asp	Glu	Val	Phe	Ser 925	Lys	Tyr	Tyr	
Thr	Pro 930	Val	Leu	Ala	Lys	Ala 935	Val	Asp	Gly	Tyr	Val 940		Pro	Gln	Ile	
Lys 945	Gln	Val	Val	Pro	Glu 950	Phe	Val	Asn	Ala	Ser 955	Ala	Asp	Ala	Gly	Gly 960	
Ser	Ser	Ala	Thr	Tyr 965	Met	Asp	Gln	Ala	Pro 970	Ser	Pro	Ala	Val	Cys 975	Pro	
Gln	Ala	Pro	Tyr 980	Asn	Met	Tyr	Pro	Gln 985	Asn	Pro	Asp	His	Val 990	Leu	Asp	
Gln	Asp	Gly 995	Glu	Phe	Asp		Asp 1000	Glu	Thr	Met	_	Val 1005	Ala	Arg	His	
	Glu 1010	Glu	Leu	Leu		Arg 1015	Pro	Met	Asp		Leu 1020	Asp	Ser	Arg	Leu	
Ser 025	Pro	Pro	Ala	_	Leu 1030	Phe	Thr	Ser		Arg 1035	Gly	Ser	Leu		1	
		(2)) IN	FORM	ATIOI	v FOI	R SE(Q ID	NO:	56:						
	(:	i) SI	EQUEI	VCE (CHAR	ACTE	RIST	ICS:								
						basic ac	se pa	airs								
		(C)	STRA	ANDEI	ONES!		ingle	е								
	(-					E: cI										
			FEAT													
		(B)	LO	TATIO	ON: 3	Codi: L: MAT:		equei	nce							
	(:	ki) s	SEQUI	ENCE	DESC	RIP	rion	: SE	Q ID	NO:	56:					
	GCG Ala															48
1				5					10					15		
	ACT Thr	Ala														96

AAG GGG CAG CCA TTC GAT GTG GGC CCA CGC TAC ACG CAG CTG CAG TAC Lys Gly Gln Pro Phe Asp Val Gly Pro Arg Tyr Thr Gln Leu Gln Tyr 35 40 45

ATC Ile	GGC Gly 50	GAG Glu	GGC Gly	GCG Ala	TAC Tyr	GGC Gly 55	ATG Met	GTC Val	AGC Ser	TCA Ser	GCT Ala 60	TAT Tyr	GAC Asp	CAC His	GTG Val	192
CGC Arg 65	AAG Lys	ACC Thr	AGA Arg	GTG Val	GCC Ala 70	ATC Ile	AAG Lys	AAG Lys	ATC Ile	AGC Ser 75	CCC Pro	TTT Phe	GAG Glu	CAT His	CAA Gln 80	240
ACC Thr	TAC Tyr	TGT Cys	CAG Gln	CGC Arg 85	ACG Thr	CTG Leu	AGG Arg	GAG Glu	ATC Ile 90	CAG Gln	ATC Ile	TTG Leu	CTG Leu	CGA Arg 95	TTC Phe	288
CGC Arg	CAT His	GAG Glu	AAT Asn 100	GTT Val	ATA Ile	GGC Gly	ATC Ile	CGA Arg 105	GAC Asp	ATC Ile	CTC Leu	AGA Arg	GCG Ala 110	CCC	ACC Thr	336
CTG Leu	GAA Glu	GCC Ala 115	Met	AGA Arg	GAT Asp	GTT Val	ТАС Туг 120	ATT Ile	GTT Val	CAG Gln	GAC Asp	CTC Leu 125	ATG Met	GAG Glu	ACA Thr	384
GAC Asp	CTG Leu 130	Tyr	AAG Lys	CTG Leu	CTT Leu	AAA Lys 135	AGC Ser	CAG Gln	CAG Gln	CTG Leu	AGC Ser 140	Asn	GAC Asp	CAC	ATC	432
TGC Cys	Tyr	TTC Phe	CTC	TAC Tyr	CAG Gln 150	Ile	CTC Leu	CGG Arg	GGC Gly	CTC Leu 155	Lys	TAT Tyr	ATA	CAC His	TCA Ser 160	480
GCC Ala	C AAT AST	GTC Val	G CTC	CAC His 165	Arg	GAC Asp	CTG Leu	AAG Lys	Pro	Ser	CAA:	CTC	CTI Let	175	AAC Asn	528
AC(C ACC	TG(C GAG S Asp 180	Leu	'AAC	ATC Ile	TGI Cys	GAT Asp 185	Ph∈	GGC Gly	CTC Lev	G GCC	CGC Arg 190	g Ile	r GCT e Ala	576
GA(Ası	CC' Pro	GA(Gl)	u Hi:	C GAC s Asp	CAC His	ACT Thr	GG(G1)	/ Phe	CTC	ACC 1 Thi	G GAG	TA' 1 Ty: 20!	r Vai	G GCG	C ACA a Thr	624
CG(TG Tr 21	o Ty	C CG.	A GCC g Ala	CCA Pro	A GAC O Glu 215	ılle	C ATC	G CTI	AA 1 ASI	r TCC n Se: 22:	r Ly	G GGG S Gl	с та у ту	C ACC r Thr	672
AA Ly 22	s Se	C AT	C GA e As	C ATO	TG(Tr) 23	o Sei	r GTY	G GGK	TG(y Cy:	E AT's Ile	e Le	G GC u Al	T GA: a Gl	G AT u Me	G CTC t Leu 240	720
TC Se	C AA r As	c cg n Ar	G CC g Pr	C ATY O Ile 24	e Ph	c cc e Pr	C GG G G1	C AAG y Ly:	G CAG S Hi: 25	s Ту	C CT r Le	G GA u As	C CA p Gl	G CT n Le 25	C AAC eu Asn 55	768
CA Hi	C AT s Il	T CT e Le	'A GG eu G1 26	y Il	C TT e Le	G GG u Gl	T TC y Se	C CC. r Pr 26	o Se	C CA r Gl	G GA n Gl	.G GA .u As	C CT sp Le 27	eu As	AT TGC sn Cys	816
AT Il	C AI e Il	T AA e As	C AT n Me	G AA t Ly	G GC s Al	C CG a Ar	A AA g As	C TA n Ty	C CT	G CA u Gl	.G TC .n Se	T CT	rg co eu Pr	CC TO	CG AAA er Lys	864

275 280 285

Thr	AAG Lys 290	GTG Val	GCT Ala	TGG Trp	GCC Ala	AAG Lys 295	CTC Leu	TTT Phe	CCT Pro	Lys	TCT Ser 300	GAC Asp	TCC Ser	AAA Lys	GCT Ala	912
CTT Leu 305	GAC Asp	CTG Leu	CTG Leu	GAC Asp	CGG Arg 310	ATG Met	TTA Leu	ACC Thr	TTC Fhe	AAC Asn 315	CCA Pro	AAC Asn	AAG Lys	CGC Arg	ATC Ile 320	960
ACA Thr	GTA Val	GAG Glu	GAA Glu	GCG Ala 325	CTG Leu	GCT Ala	CAC His	CCT Pro	TAC Tyr 330	CTG Leu	GAA Glu	CAG Gln	TAC Tyr	TAC Tyr 335	GAT Asp	1008
CCG Pro	ACA Thr	TAD qzA	GAG Glu 340	CCA Pro	GTG Val	GCC Ala	GAG Glu	GAG Glu 345	CCA Pro	TTC Phe	ACC Thr	TTC Phe	GAC Asp 350	ATG Met	GAG Glu	1056
CTG Leu	GAT Asp	GAC Asp 355	CTC Leu	CCC Pro	AAG Lys	GAG Glu	CGG Arg 360	CTG Leu	AAG Lys	GAG Glu	TTG Leu	ATC Ile 365	TTC Phe	CAG Gln	GAG Glu	1104
ACA Thr	GCC Ala 370	CGC Arg	TTC Phe	CAG Gln	CCA Pro	GGG Gly 375	GCG Ala	CCA Pro	GAG Glu	GGC Gly	CCC Pro 380	Gly	CGC Arg	GCC Ala	ATG Met	1152
AGT Ser 385	AAA Lys	GGA Gly	GAA Glu	GA.A Glu	CTT Leu 390	TTC Phe	ACT Thr	GGA Gly	GTT Val	GTC Val 395	CCA Pro	ATT	CTT	GTT Val	GAA Glu 400	1200
TTA Leu	GAT Asp	GGC Gly	GAT Asp	GTI Val	. Asn	GGG Gly	CAA Gln	. AAA . Lys	TTC Phe 410	Ser	GTT Val	AGT Ser	GGA Gly	GAC Glu 415	GGT 1 Gly	1248
GAA Glu	GGT Gly	GAT Asp	GCA Ala 420	Thi	TAC	GGA Gly	AAA Lys	CTT Lev 425	1 Thr	CTT Leu	`AAA	A TTI S Phe	116 430	Cy:	ACT Thr	1296
ACT Thr	G17 GGG	AAC Lys 435	Leu	CC.	r GTT o Val	CCA Pro	TGC Trp 440	Pro	A ACC	CTI	GTC Val	C ACT l Thr 445	Thi	r CTY	C ACT u Thr	1344
ТАТ Туг	GGT Gl ₃ 450	/ Val	CAF Glr	A TGG	TTI S Phe	TCT Ser 459	Arg	TAC	c cca r Pro	A GAT	CA' Hi:	s Met	G AAI E Ly:	A CA	G CAT n His	1392
GAC Asi 465	Fhe	r TT(∋ Phe	AAC E Lys	G AG'	T GCC r Ala 470	a Me	CCC Pro	G GA	A GG' u Gl	TA:	c Va	A CAG	g GA n Gl	A AG u Ar	A ACT g Thr 480	1440
ATA Ile	A TT e Pho	r TAC e Tyr	C AAA	A GA s As 48	p Ası	GGG Gl:	AA E Y As:	C TA n Ty	C AAG r Ly: 49	s Thi	A CG r Ar	T GC	T GA a Gl	A GT u Va 49	C AAG 1 Lys 5	1488
TT" Fh	r GA e Gl	A GG u Gl	T GA Y As; 50	p Th	C CT	r GT u Va	T AA 1 As	T AG n Ar 50	g Il	C GA	G TT u Le	'A AA :u Ly	A GG s Gl 51	λIJ	T GAT .e Asp	1536

TTT Phe	AAA Lys	GAA Glu 515	GAT Asp	GGA Gly	AAC Asn	ATT Ile	CTT Leu 520	GGA Gly	CAC His	AAA Lys	ATG Met	GAA Glu 525	TAC Tyr	AAT Asn	TAT Tyr		1584
AAC Asn	TCA Ser 530	CAT His	AAT Asn	GTA Val	TAC Tyr	ATC Ile 535	ATG Met	GCA Ala	GAC Asp	AAA Lys	CCA Pro 540	AAG Lys	AAT Asn	GGC Gly	ATC Ile		1632
AAA Lys 545	GTT Val	AAC Asn	TTC Phe	AAA Lys	ATT Ile 550	AGA Arg	CAC His	AAC Asn	ATT Ile	AAA Lys 555	GAT Asp	GGA Gly	AGC Ser	GTT Val	CAA Gln 560		1680
TTA Leu	GCA Ala	GAC Asp	CAT His	TAT Tyr 565	CAA Gln	CAA Gln	AAT Asn	ACT Thr	CCA Pro 570	ATT Ile	GGC Gly	GAT Asp	GGC Gly	CCT Pro 575	GTC Val		1728
CTT Leu	TTA Leu	CCA Pro	GAC Asp 580	Asn	CAT	TAC Tyr	CTG Leu	TCC Ser 585	Thr	CAA Gln	TCT Ser	GCC	CTT Leu 590	TCC Ser	AAA Lys		1776
GAT Asp	CCC	AAC Asn 595	Glu	. AAG . Lys	AGA Arg	GAT Asp	CAC His	Met	ATC Ile	CTT Leu	CTI Leu	GAG Glu 605	Phe	GTA Val	ACA Thr		1824
GCT Ala	GCT Ala	Gly	ATI / Ile	ACA Thr	CAT His	GGC Gly 615	Met	GAT Asp	GAA	CTA Leu	TAC TYI 620	Lys	CCT Pro	CAG Glr	GAG Glu	Т	1873
AA																	1875

(2) INTFORMATION FOR SEQ ID NO:57:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 624 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:

Met Ala Ala Ala Ala Ala Pro Gly Gly Gly Gly Glu Pro Arg 10 Gly Thr Ala Gly Val Val Pro Val Val Pro Gly Glu Val Glu Val Val 20 25 Lys Gly Gln Pro Phe Asp Val Gly Pro Arg Tyr Thr Gln Leu Gln Tyr 40 Ile Gly Glu Gly Ala Tyr Gly Met Val Ser Ser Ala Tyr Asp His Val 55 Arg Lys Thr Arg Val Ala Ile Lys Lys Ile Ser Pro Phe Glu His Gln 70 Thr Tyr Cys Gln Arg Thr Leu Arg Glu Ile Gln Ile Leu Leu Arg Phe 90 Arg His Glu Asn Val Ile Gly Ile Arg Asp Ile Leu Arg Ala Pro Thr 105 110 100 Leu Glu Ala Met Arg Asp Val Tyr Ile Val Gln Asp Leu Met Glu Thr

		115					120					125			
	130					135					140				
Cys 7					150					155					160
Ala A	Asn	Val	Leu	His 165	Arg	Asp	Leu	Lys	Pro 170	Ser	Asn	Leu	Leu	11e 175	Asn
Thr S			180					185					190		
Asp !		195					200					205			
	210					215					220				
Lys : 225					230					235					240
Ser .				245					250					255	
His			260					265					270		
		275					Asn 280					285			
	290					295	Leu				300				
305					310		Leu			315					320
				325			His		330					335	
			340				Glu	345					350		
		355					Arg 360					365			
	370					375					380				
385					390					395)				Glu 400
				405	,				410)				415	
			420					425					430)	Thr
		435					440					445	•		Thr
	450					455	5				460)			His
465					470	ı				475	ō				Thr 480
				485	5				490)				499	
			500)				505	5				510)	Asp
		515	<u>,</u>				520)				525	5		ı Tyr
	530)				53	5				540)			y Ile
545					55()				55	5				1 Gln 560
				56	5				57	0				57	
Leu	Lev	Pro	a Ası	o Asi	n His	s Ty	r Lei	ı Se:	r Th	r Gl	n Se	r Al	a Le	u Se	r Lys

		5	80					85					90			
Asp P	5	95				ϵ	00				6	05				
Ala A 6	la (Sly 1	le 7	Thr F		Sly M 515	iet A	sp G	Slu I		Tyr I 520	ys F	ro (3ln (Glu	
		(2)	INFO	ORMA'	NOIT	FOR	SEQ	ID 1	10:58	3:						
		(A) ! (B) ' (C) .	LENG' TYPE STRAI	FH: : nu : NDED	HARAC 1815 cleic NESS : lin	base ac: sir	e pai id									
		i) M ×) F			TYPE	: cDi	AV									
		(B)	LOC	OITA	Y: C N: 1 NFOR	1	811	quen	ce							
	(×	i) S	EQUE	NCE	DESC	RIPT	ION:	SEQ	ID	NO:5	8:					
ATG Met .	GCG Ala	GCG Ala	GCG Ala	GCG Ala 5	GCG Ala	GCG Ala	GGC Gly	CCG Pro	GAG Glu 10	ATG Met	GTC Val	CGC Arg	GGG Gly	CAG Gln 15	GTG Val	48
TTC Phe	GAC Asp	GTG Val	GGG Gly 20	CCG Pro	CGC Arg	TAC Tyr	ACT Thr	AAT Asn 25	CTC Leu	TCG Ser	TAC Tyr	ATC Ile	GGA Gly 30	GAA Glu	GGC Gly	96
GCC Ala	TAC Tyr	GGC Gly 35	ATG Met	GTT Val	TGT Cys	TCT Ser	GCT Ala 40	тат туг	GAT Asp	AAT Asn	CTC Leu	AAC Asn 45	AAA Lys	GTT Val	CGA Arg	144
GTT Val	GCT Ala 50	ATC Ile	AAG Lys	AAA Lys	ATC Ile	AGT Ser 55	CCT Pro	TTT Phe	GAG Glu	CAC His	CAG Gln 60	ACC Thr	TAC Tyr	TGT Cys	CAG Gln	192
AGA Arg 65	ACC Thr	CTG Leu	AGA Arg	GAG Glu	ATA Ile 70	AAA Lys	ATC Ile	CTA Leu	CTG Leu	CGC Arg 75	TTC Phe	AGA Arg	CAT His	GAG Glu	AAC Asn 80	240
ATC Ile	ATC Ile	GGC Gly	ATC Ile	AAT Asn 85	GAC Asp	ATC Ile	ATC Ile	CGG Arg	GCA Ala 90	CCA Pro	ACC Thr	ATT	GAG Glu	CAG Gln 95	ATG Met	288
AAA Lys	GAT Asp	GTA Val	TAT Tyr 100	Ile	GTA Val	CAG Gln	GAC Asp	CTC Leu 105	Met	GAG Glu	ACA Thr	GAT Asp	CTI Leu 110	ı Tyr	AAG Lys	336
CTC Leu	TTG	AAG Lys 115	Thr	CAG Gln	CAC His	CTC	AGC Ser 120	Asn	'GAT	CAT His	TATC	TGC Cys 125	Туг	TTT	r CTT e Leu	384
TAT Tyr	CAC Glr 130	ıle	CTG	AGA Arg	GGA Gly	. TTA Leu 135	Lys	TAT Tyr	T ATA	A CAT	TCA Ser 140	: Ala	CAA C	r GT n Val	r CTG l Leu	432

						TCC Ser										480
						GGC Gly										528
						ACA Thr										576
						AAT Asn								_		624
						ATC Ile 215		_	_							672
						TAC Tyr										720
						CAG Gln										768
						CTT Leu										816
						AAC Asn										864
						AAC Asn 295										912
	Leu			Pro		CTG Leu		Gln		Tyr	Asp					960
						TTC										1008
						GAA Glu										1056
			Tyr			ATG Met							Met			1104
AAG	GGC	GAG	GAG	CTG	TTC	ACC	GGG	GTG	GTG	CCC	ATC	CTG	GTC	GAG	CTG	1152

Lys	Gly 370	Glu	Glu	Leu	Phe	Thr 375	Gly	Val	Val	Pro	Ile 380	Leu	Val	Glu	Leu	
GAC Asp 385	GGC Gly	GAC Asp	GTA Val	AAC Asn	GGC Gly 390	CAC His	AAG Lys	TTC Phe	AGC Ser	GTG Val 395	TCC Ser	GGC Gly	GAG Glu	GGC Gly	GAG Glu 400	1200
GGC Gly	GAT Asp	GCC Ala	ACC Thr	TAC Tyr 405	GGC Gly	AAG Lys	CTG Leu	ACC Thr	CTG Leu 410	AAG Lys	TTC Phe	ATC Ile	TGC Cys	ACC Thr 415	ACC Thr	1248
GGC Gly	AAG Lys	CTG Leu	CCC Pro 420	GTG Val	CCC Pro	TGG Trp	CCC Pro	ACC Thr 425	CTC Leu	GTG Val	ACC Thr	ACC Thr	CTG Leu 430	ACC Thr	TAC Tyr	1296
					AGC Ser											1344
TTC Phe	TTC Phe 450	AAG Lys	TCC Ser	GCC Ala	ATG Met	CCC Pro 455	GAA Glu	GGC Gly	TAC Tyr	GTC Val	CAG Gln 460	Glu	CGC	ACC Thr	ATC Ile	1392
TTC Phe 465	Phe	AAG Lys	GAC Asp	GAC Asp	GGC Gly 470	AAC Asn	TAC Tyr	AAG Lys	ACC Thr	CGC Arg 475	GCC Ala	GAG Glu	GTG Val	AAG Lys	TTC Phe 480	1440
GAG Glu	GGC Gly	GAC Asp	ACC Thr	CTG Leu 485	Val	AAC Asn	CGC Arg	ATC Ile	GAG Glu 490	Leu	AAC Lys	GGC Gly	ATC	GAC Asp 495	TTC Phe	1488
AAG Lys	GAG Glu	GAC Asp	GGC Gly 500	Asn	: ATC	CTG Leu	GGG Gly	CAC His 505	Lys	CTG Leu	GAC Glu	TAC 1 Tyr	AAC Asr 510	Ty:	AAC Asn	1536
AGC Ser	CAC His	AAC Asn 515	Val	TAT Tyr	ATC	ATG Met	GCC Ala 520	Asp	AAG Lys	CAG Glr	AAC Lys	3 AA0 3 Asr 525	ı Gly	TATO	C AAG e Lys	1584
GTC Val	AAC Asn 530	Phe	AAG Lys	ATC	CGC Arg	CAC His	Asn	: ATC	GAG Glu	GAC Asp	GG(G1) 540	y Se	C GTV	G CAG	G CTC n Leu	1632
GC0 Ala 545	a Asp	CAC His	TAC	CAC Glr	G CAG n Glr 550	Asn	ACC Thr	CCC	ATC	GGG Gly 555	y As	C GG(p Gl;	C CC y Pr	C GT o Va	G CTG 1 Leu 560	
CT(Le	G CCC	GAC Asi	AAC Asr	CAC h His 565	з Туг	CTC Lev	AGC Sei	C ACC	CAC Glr 570	n Se	C GC r Al	C CT a Le	G AG u Se	C AA r Ly 57	A GAC s Asp 5	1728
CC(C AAC o Asi	GA(G AAC Lys 580	s Ar	C GAT	r CAC	TA C	G GTX t Val 585	l Le	J Le	G GA u Gl	G TT u Ph	C GT e Va 59	1 Th	C GCC	1776
			e Thi		C GG(u Gl <u>)</u>			p Gl					A			1815

- (2) INFORMATION FOR SEQ ID NO:59:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 604 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:59:

Met Ala Ala Ala Ala Ala Gly Pro Glu Met Val Arg Gly Gln Val 10 5 1 Phe Asp Val Gly Pro Arg Tyr Thr Asn Leu Ser Tyr Ile Gly Glu Gly 25 20 Ala Tyr Gly Met Val Cys Ser Ala Tyr Asp Asn Leu Asn Lys Val Arg 40 Val Ala Ile Lys Lys Ile Ser Pro Phe Glu His Gln Thr Tyr Cys Gln Arg Thr Leu Arg Glu Ile Lys Ile Leu Leu Arg Phe Arg His Glu Asn 75 Ile Ile Gly Ile Asn Asp Ile Ile Arg Ala Pro Thr Ile Glu Gln Met 90 Lys Asp Val Tyr Ile Val Gln Asp Leu Met Glu Thr Asp Leu Tyr Lys 100 105 Leu Leu Lys Thr Gln His Leu Ser Asn Asp His Ile Cys Tyr Phe Leu 120 125 Tyr Gln Ile Leu Arg Gly Leu Lys Tyr Ile His Ser Ala Asn Val Leu 140 135 His Arg Asp Leu Lys Pro Ser Asn Leu Leu Leu Asn Thr Thr Cys Asp 155 150 Leu Lys Ile Cys Asp Phe Gly Leu Ala Arg Val Ala Asp Pro Asp His 170 165 Asp His Thr Gly Phe Leu Thr Glu Tyr Val Ala Thr Arg Trp Tyr Arg 185 180 Ala Pro Glu Ile Met Leu Asn Ser Lys Gly Tyr Thr Lys Ser Ile Asp 200 205 Ile Trp Ser Val Gly Cys Ile Leu Ala Glu Met Leu Ser Asn Arg Pro 215 220 Ile Phe Pro Gly Lys His Tyr Leu Asp Gln Leu Asn His Ile Leu Gly 230 235 lle Leu Gly Ser Pro Ser Gln Glu Asp Leu Asn Cys Ile Ile Asn Leu 245 250 Lys Ala Arg Asn Tyr Leu Leu Ser Leu Pro His Lys Asn Lys Val Pro 265 260 Trp Asn Arg Leu Phe Pro Asn Ala Asp Ser Lys Ala Leu Asp Leu Leu 280 285 Asp Lys Met Leu Thr Phe Asn Pro His Lys Arg Ile Glu Val Glu Gln 295 Ala Leu Ala His Pro Tyr Leu Glu Gln Tyr Tyr Asp Pro Ser Asp Glu 310 315 Pro Ile Ala Glu Ala Pro Phe Lys Phe Asp Met Glu Leu Asp Asp Leu 330

Pro Lys Glu Lys Leu Lys Glu Leu Ile Phe Glu Glu Thr Ala Arg Phe

340 345 Gln Pro Gly Tyr Arg Ser Met Asp Pro Pro Val Ala Thr Met Val Ser 355 360 Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu 375 Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu 390 395 Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr 405 410 Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr 420 425 Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp 435 440 Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile 460 455 Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe 465 470 475 Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe 490 495 485 Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn 505 510 500 Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys 520 515 Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu 535 540 Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu 555 545 550 Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp 570 575 565 Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala 580 585 Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 600

(2) INFORMATION FOR SEQ ID NO:60:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2511 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...2508
 - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:

ATG GAG CTG GAA AAC ATC GTG GCC AAC ACG GTC TTG CTG AAA GCC AGG
Met Glu Leu Glu Asn Ile Val Ala Asn Thr Val Leu Leu Lys Ala Arg

1 5 10 15

								CAG Gln								144
								TGT Cys								192
								ACC Thr								240
								GAA Glu								288
					_			ATT Ile 105								336
			_		_	_		GTT Val								384
								CCG Pro								432
								CTG Leu								480
								CGC Arg								528
		_		_				ACT Thr 185			_			_		576
							Val	TGT Cys								624
								TTG Leu								672
								AAT Asn								720
								CTG Leu								768
GAT	GCA	CTG	TGC	TTG	GTC	CTG	ACC	ATC	ATG	AAT	GGG	GGT	GAC	CTG	AAG	816

Asp	Ala	Leu	Суs 260	Leu	Val	Leu	Thr	Ile 265	Met	Asn	Gly	Gly	Asp 270	Leu	Lys	
								CCT Pro								864
								TGC Cys								912
								AAA Lys								960
								GAC Asp								1008
								CGG Arg 345								1056
								AGG Arg								1104
		Leu						GAG Glu								1152
	Arg					Lys		AAG Lys			Glu					1200
					Glu			TCC Ser							Ala	1248
AAC Lys	TCC Ser	ATC	TGC Cys 420	Lys	ATG Met	CTG Leu	CTC Leu	ACG Thr 425	Lys	GAT Asp	GCG Ala	AAC Lys	G CAC Glr 430	Arg	CTG Leu	1296
			Glu					a Glu					s Pro		TTC Phe	1344
AGC Arg	3 AAC 3 Asr 450	n Met	AAC Asr	TTC h Fhe	AAC Lys	G CGC Arg 455	Le	A GAF ı Glu	GCC Ala	GG(G ATC / Met 460	Lev	G GAG	C CC	CCC Pro	1392
	e Val					g Ala					s Asp				C ATC p Ile 480	1440
					· Val					ı Le					C GAC p Asp 5	1488

						TCC Ser										1536
						GAA Glu										1584
		_				CCA Pro 535										1632
						CTC Leu										1680
						CCC Pro										1728
						AGC Ser										1776
						GTG Val										1824
			_			GAG Glu 615										1872
						GGC Gly										1920
						ACC Thr										1968
						ACC Thr										2016
						CAC His									GAA Glu	2064
						ACC Thr 695										2112
						AAG Lys										2160
ATC	GAG	CTG	AAG	GGC	ATC	GAC	TTC	AAG	GAG	GAC	GGC	AAC	ATC	CTG	GGG	2208

Ile	Glu	Leu	Lys	Gly 725	Ile	Asp	Phe	Lys	Glu 730	Asp	Gly	Asn	Ile	Leu 735	Gly	
									CAC His							2256
									AAC Asn							2304
									GAC Asp							2352
									CCC Pro		Asn					2400
									AAC Asn 810						ATG Met	2448
				Phe					Gly					Met	GAC Asp	2496
	CTG Leu		Lys		.											2511

(2) INFORMATION FOR SEQ ID NO:61:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 836 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:61:

 Met
 Glu
 Leu
 Glu
 Asn
 I le
 Val
 Ala
 Asn
 Thr
 Val
 Leu
 Leu
 Lys
 Ala
 Arg
 Lys
 Gly
 Lys
 Lys
 Bar
 Lys
 Gly
 Lys
 Ser
 Lys
 Lys
 Lys
 Glu
 Arg
 Lys
 Glu
 Asp
 Leu
 Arg
 Arg
 Thr
 Arg
 Glu
 Lys
 Glu
 Asp
 Leu
 Arg
 Arg
 Thr
 Arg</t

			100					105					110		
Lys	Ser	Pro 115	Val	Phe	Ile	Ala	Gln 120	Val	Gly	Gln	Asp	Leu 125	Val	Ser	Gln
Thr	Glu 130	Glu	Lys	Leu	Leu	Gln 135	Lys	Pro	Cys	Lys	Glu 140	Leu	Phe	Ser	Ala
Cys 145	Ala	Gln	Ser	Val	His 150	Glu	Tyr	Leu	Arg	Gly 155	Glu	Pro	Phe	His	Glu 160
_			Ser	165				_	170					175	
			Pro 180			_		185				_	190		
_	_	195	Gly				200	_		_		205			
_	210		Tyr			215					220				
Arg 225	Lys	Gly	Glu	Ser	Met 230	Ala	Leu	Asn	Glu	Lys 235	Gln	He	Leu	GIu	Lys 240
	Asn	Ser	Gln	Phe 245		Val	Asn	Leu	Ala 250		Ala	Tyr	Glu	Thr 255	
Asp	Ala	Leu	Суs 260	Leu	Val	Leu	Thr	11e 265	Met	Asn	Gly	Gly	Asp 270	Leu	Lys
		275	Tyr				280					285			
	290		Ala			295					300				
305			Val		310					315					320
-	_	_	His	325					330					335	
		_	Asp 340					345					350		
		355	Val				360	_	_			365			
	370		Gly			375					380				
385	_	_	Arg Thr	_	390	_		_		395					400
			Cys	405					410					415	
			420 Glu					425					430		
		435					440					445			Pro
_	450					455					460				
465					470					475					11e 480
				485					490					495	Asp
			500					505					510		Gln
		515					520					525			Gly
	530					535					540				Glu
545					550					555					Gln 560
Asn	Asn	ser	rys	ser	ser	PIO	ser	ser	гÀг	unr	ser	rne	ASN	пıS	His

570 565 Ile Asn Ser Asn His Val Ser Ser Asn Ser Thr Gly Ser Ser Arg Asp 580 585 590 Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly **59**5 600 Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys 610 615 620 Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu 630 635 Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro 645 650 655 Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr 660 665 670 Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu 675 680 Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr 690 695 700 Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg 705 710 715 720 Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly 725 730 735 His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala 740 745 750 Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn 755 760 765 Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr 770 775 780 Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser 790 795 800 Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met 805 810 815 Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp 825 820 Glu Leu Tyr Lys 835

(2) INFORMATION FOR SEQ ID NO:62:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1893 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: CDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...1890
 - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:

ATG AGC AGA AGC AAG CGT GAC AAC AAT TTT TAT AGT GTA GAG ATT GGA
Met Ser Arg Ser Lys Arg Asp Asn Asn Phe Tyr Ser Val Glu Ile Gly
1 5 10 15

GAT TOT ACA TTO ACA GTO CTG AAA CGA TAT CAG AAT TTA AAA COT ATA

Asp	Ser	Thr	Phe 20	Thr	Val	Leu	Lys	Arg 25	Tyr	Gln	Asn	Leu	Lys 30	Pro	Ile	
GGC Gly	TCA Ser	GGA Gly 35	GCT Ala	CAA Gln	GGA Gly	ATA Ile	GTA Val 40	TGC Cys	GCA Ala	GCT Ala	TAT Tyr	GAT Asp 45	GCC Ala	ATT Ile	CTT Leu	144
GAA Glu	AGA Arg 50	AAT Asn	GTT Val	GCA Ala	ATC Ile	AAG Lys 55	AAG Lys	CTA Leu	AGC Ser	CGA Arg	CCA Pro 60	TTT Phe	CAG Gln	AAT Asn	CAG Gln	192
ACT Thr 65	CAT His	GCC Ala	AAG Lys	CGG Arg	GCC Ala 70	TAC Tyr	AGA Arg	GAG Glu	CTA Leu	GTT Val 75	CTT Leu	ATG Met	AAA Lys	TGT Cys	GTT Val 80	240
AAT Asn	CAC His	AAA Lys	AAT Asn	ATA Ile 85	ATT Ile	GGC Gly	CTT Leu	TTG Leu	AAT Asn 90	GTT Val	TTC Phe	ACA Thr	CCA Pro	CAG Gln 95	AAA Lys	288
TCC Ser	CTA Leu	GAA Glu	GAA Glu 100	TTT Phe	CAA Gln	GAT Asp	GTT Val	TAC Tyr 105	ATA Ile	GTC Val	ATG Met	GAG Glu	CTC Leu 110	Met	GAT Asp	336
GCA Ala	AAT Asn	CTT Leu 115	Cys	CAA Gln	GTG Val	ATT	CAG Gln 120	ATG Met	GAG Glu	CTA Leu	GAT Asp	CAT His	Glu	AGA Arg	ATG Met	384
TCC Ser	ТАС Тут 130	Leu	CTC	тат Туг	CAG Gln	ATG Met 135	CTG Leu	TGT Cys	GGA Gly	ATC Ile	AAG Lys 140	His	CTI Lev	CAT His	TCT Ser	432
	Gly					Asp					Asr				A AAA Lys 160	480
TCT Ser	GAT Asp	TGC Cys	ACT Thr	TTC Lev 165	Lys	ATT	CTI Leu	GAC Asp	TTC Phe 170	G17	CTC	G GCC	a Arg	G ACT g Thi 175	GCA Ala	528
GG# Gly	ACC Thr	AGT Sei	TTT Phe	Met	ATG : Met	ACG Thr	CCT	TAT Tyr 185	· Val	GT(G ACT	r Ar	TAC TY1 190	r Ty	AGA Arg	576
GCA Ala	A CCC	GAG Glv 195	ı Val	ATC	CTI Leu	GGC Gly	ATC Met	Gly	TAC Tyr	Ly:	G GA	A AA u As 20	n Va	G GA' l As	T TTA p Leu	624
TG(Tr)	G TCT Ser 210	· Va	G GGC	FTG0	TATT	T ATC Met 215	Gl	A GAA y Glu	A ATO	GT Va	T TG 1 Cy 22	s Hi	C AA s Ly	TA A S Il	C CTC e Leu	672
TT Pho 22	e Pro	A GG.	A AGO y Arg	GA(JAS	тап р Туз 230	c Ile	r GA′ ∂ Asj	T CAC	G TG(G AA O As 23	n Ly	A GT s Va	T AT	T GA e Gl	A CAG u Gln 240	720
CT Le	T GG u Gl	A AC y Th	A CCZ r Pro	A TG' 5 Cy: 24	s Pro	r GAX o Gli	A TT	C ATY	3 AAG t Ly: 250	s Ly	A CT s Le	G CA u Gl	A CC n Pr	A AC o Th	A GTA r Val	768

AGG Arg	ACT Thr	TAC Tyr	GTT Val 260	GAA Glu	AAC Asn	AGA Arg	Pro	AAA Lys 265	TAT Tyr	GCT Ala	GGA Gly	TAT Tyr	AGC Ser 270	TTT Phe	GAG Glu	816
AAA Lys	CTC Leu	TTC Phe 275	CCT Pro	GAT Asp	GTC Val	CTT Leu	TTC Phe 280	CCA Pro	GCT Ala	GAC Asp	TCA Ser	GAA Glu 285	CAC His	AAC Asn	AAA Lys	864
								TTG Leu								912
GAT Asp 305	GCA Ala	TCT Ser	AAA Lys	AGG Arg	ATC Ile 310	TCT Ser	GTA Val	GAT Asp	GAA Glu	GCT Ala 315	CTC Leu	CAA Gln	CAC His	CCG Pro	TAC Tyr 320	960
ATC Ile	AAT Asn	GTC Val	TGG Trp	TAT Tyr 325	GAT Asp	CCT Pro	TCT Ser	GAA Glu	GCA Ala 330	GAA Glu	GCT Ala	CCA Pro	CCA Pro	CCA Pro 335	Lys	1008
ATC Ile	CCT Pro	GAC Asp	AAG Lys 340	Gln	TTA Leu	GAT Asp	GAA Glu	AGG Arg 345	GAA Glu	CAC His	ACA Thr	ATA Ile	GAA Glu 350	Glu	TGG Trp	1056
AAA Lys	GAA Glu	TTC Lev 355	ıle	TAT	AAG Lys	GAA Glu	GTT Val 360	ATG Met	GAC Asp	TTG	GAG Glu	GAG Glu 365	Arc	ACC Thr	AAG Lys	1104
AAT Asn	GGA Gly 370	Va]	T ATA	A CGG Arg	GGG Gly	CAG Gln 375	Pro	TCT Ser	CCT	TTA Lev	GCA Ala 380	Glr	GTC 1 Val	G CAG	G CAG n Gln	1152
TGC Trp 385) Asp	CCA Pro	A CCC	G GTC	GCC Ala 390	Thr	ATG Met	GTG Val	AGC Ser	AAC Lys 395	Gly	GAC	G GAG	G CTY	G TTC u Phe 400	1200
AC(Thi	c Gly	GTY Va	G GTV	G CCC 1 Pro 405	o Il€	CTG Lev	GTC Val	GAG Glu	CTC Lev 410	. Ası	617 C GGC	GA(C GT.	A AA 1 As 41	c GGC n Gly 5	1248
CA(C AAK s Lys	G TT s Ph	e Se	r Va	l Sei	c Gly	/ Glu	G GGC 1 Gly 425	/ Glu	GGGG	C GAT	r GC(C AC a Th 43	r Ty	c GGC r Gly	1296
AA(Ly	G CTO	G AC u Th 43	r Le	G AA u Ly	G TTO	TA C	TG(E Cy:	s Thi	C ACC	c GG r Gl	C AA(y Ly:	G CT s Le 44	u Pr	C GI	G CCC il Pro	1344
TG Tr	G CC p Pr 45	o Th	C CT	C GT u Va	G AC	C ACC r Th: 45	r Le	G ACC	C TAC r Ty:	c GG r Gl	C GT y Va 46	1 G1	.G TO n Cy	C TI	C AGC ne Ser	1392
CG Ar 46	g Ty	C CC	C GA	C CA sp Hi	C AT s Me 47	t Ly	G CA s Gl	G CAO	C GA s As	C TT p Ph 47	e Ph	C AA e Ly	G TO	CC GC er Al	CC ATG la Met 480	
CC	C GA	A GC	SC TA	C GI	C CA	.g ga	G CG	C AC	C AT	C TI	C TI	'C A	AG GA	AC G	AC GGC	1488

Pro	Glu	Gly	Tyr	Val 485	Gln	Glu	Arg	Thr	Ile 490	Phe	Phe	Lys	Asp	Asp 495	Gly	
AAC Asn	TAC Tyr	AAG Lys	ACC Thr 500	CGC Arg	GCC Ala	GAG Glu	GTG Val	AAG Lys 505	TTC Phe	GAG Glu	GGC Gly	GAC Asp	ACC Thr 510	CTG Leu	GTG Val	1536
AAC Asn	CGC Arg	ATC Ile 515	GAG Glu	CTG Leu	AAG Lys	GGC Gly	ATC Ile 520	GAC Asp	TTC Phe	AAG Lys	GAG Glu	GAC Asp 525	GGC Gly	AAC Asn	ATC Ile	1584
CTG Leu	GGG Gly 530	CAC His	AAG Lys	CTG Leu	GAG Glu	ТАС Туг 535	AAC Asn	TAC Tyr	AAC Asn	AGC Ser	CAC His 540	AAC Asn	GTC Val	ТАТ Туг	ATC Ile	1632
ATG Met 545	GCC Ala	GAC Asp	AAG Lys	CAG Gln	AAG Lys 550	AAC Asn	GGC Gly	ATC Ile	AAG Lys	GTG Val 555	AAC Asn	TTC Phe	AAG Lys	ATC Ile	CGC Arg 560	1680
CAC His	AAC Asn	ATC Ile	GAG Glu	GAC Asp 565	Gly	AGC Ser	GTG Val	CAG Gln	CTC Leu 570	GCC Ala	GAC Asp	CAC His	TAC Tyr	CAG Gln 575	Gln	1728
AAC Asn	ACC Thr	CCC	ATC 1le 580	Gly	GAC Asp	GGC Gly	CCC	GTG Val 585	Leu	CTG Leu	CCC Pro	GAC Asp	AAC Asn 590	His	TAC Tyr	1776
CTG Leu	AGC Ser	ACC Thr 595	Gln	TCC Ser	GCC Ala	CTC Leu	AGC Ser 600	Lys	. GAC	CCC Pro	AAC Asi	GAG n Glu 605	Lys	CGC Arg	GAT J Asp	1824
CAC His	ATC Met	Va.	CTC Lev	CTC Let	G GAC	TTC Phe 619	e Val	ACC Thr	GCC Ala	GCC Alá	GG(G1) 620	y Ile	ACT Thi	CTC	GGC Gly	1872
	Asp	o Glu	G CTC	і Туі	630	S)		70 71	> NO	. 63 -						1893

(2) INFORMATION FOR SEQ ID NO:63:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 630 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:63:

Glu Arg Asn Val Ala Ile Lys Lys Leu Ser Arg Pro Phe Gln Asn Gln Thr His Ala Lys Arg Ala Tyr Arg Glu Leu Val Leu Met Lys Cys Val Asn His Lys Asn Ile Ile Gly Leu Leu Asn Val Phe Thr Pro Gln Lys Ser Leu Glu Glu Phe Gln Asp Val Tyr Ile Val Met Glu Leu Met Asp Ala Asn Leu Cys Gln Val Ile Gln Met Glu Leu Asp His Glu Arg Met Ser Tyr Leu Leu Tyr Gln Met Leu Cys Gly Ile Lys His Leu His Ser Ala Gly Ile Ile His Arg Asp Leu Lys Pro Ser Asn Ile Val Val Lys Ser Asp Cys Thr Leu Lys Ile Leu Asp Phe Gly Leu Ala Arg Thr Ala Gly Thr Ser Phe Met Met Thr Pro Tyr Val Val Thr Arg Tyr Tyr Arg Ala Pro Glu Val Ile Leu Gly Met Gly Tyr Lys Glu Asn Val Asp Leu Trp Ser Val Gly Cys Ile Met Gly Glu Met Val Cys His Lys Ile Leu Phe Pro Gly Arg Asp Tyr Ile Asp Gln Trp Asn Lys Val Ile Glu Gln Leu Gly Thr Pro Cys Pro Glu Phe Met Lys Lys Leu Gln Pro Thr Val Arg Thr Tyr Val Glu Asn Arg Pro Lys Tyr Ala Gly Tyr Ser Phe Glu Lys Leu Phe Pro Asp Val Leu Phe Pro Ala Asp Ser Glu His Asn Lys Leu Lys Ala Ser Gln Ala Arg Asp Leu Leu Ser Lys Met Leu Val Ile Asp Ala Ser Lys Arg Ile Ser Val Asp Glu Ala Leu Gln His Pro Tyr Ile Asn Val Trp Tyr Asp Pro Ser Glu Ala Glu Ala Pro Pro Pro Lys Ile Pro Asp Lys Gln Leu Asp Glu Arg Glu His Thr Ile Glu Glu Trp Lys Glu Leu Ile Tyr Lys Glu Val Met Asp Leu Glu Glu Arg Thr Lys Asn Gly Val Ile Arg Gly Gln Pro Ser Pro Leu Ala Gln Val Gln Gln Trp Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Fhe Glu Gly Asp Thr Leu Val

			500					505					510			
Asn	Arg	Ile 515	Glu	Leu	Lys	Gly	Ile 520		Phe	Lys	Glu	Asp 525		Asn	Ile	
Leu	Gly 530	His	Lys	Leu	Glu	Tyr 535	Asn	Tyr	Asn	Ser	His 540	Asn	Val	Tyr	Ile	
Met 545	Ala	Asp	Lys	Gln	Lys 550	Asn	Gly	Ile	Lys	Val 555	Asn	Phe	Lys	Ile	Arg 560	
His	Asn	Ile	Glu	Asp 565	Gly	Ser	Val	Gln	Leu 570	Ala	Asp	His	Tyr	Gln 575	Gln	
Asn	Thr	Pro	Ile 580	Gly	Asp	Gly	Pro	Val 585	Leu	Leu	Pro	Asp	Asn 590	His	Tyr	
Leu	Ser	Thr 595	Gln	Ser	Ala	Leu	Ser 600	Lys	Asp	Pro	Asn	Glu 605	Lys	Arg	qzA	
His	Met 610	Val	Leu	Leu	Glu	Phe 615	Val	Thr	Ala	Ala	Gly 620	Ile	Thr	Leu	Gly	
Met 625	Asp	Glu	Leu	Tyr	Lys 630											
		(2) IN	FORM	IOITA	N FOR	R SE(Q ID	NO : 6	54:						
	(.	i) SI (A)					RIST:									
		(B)	TYPI	E: nı	ıcle:	ic ac	_									
						inear	_	-								
		ii) N ix) B			TYPI	E: cI	ANC									
		(A)	NAN	Œ/KI	EY: C	Codir	ng Se	eguer	ice							
		(B)	LO	CATIO	ON: 1	Codir LI	818	eguer	ice							
	()	(B)	LOC OTH	CATIO HER I	ON: I	11 MATI	1818 ION:			NO:6	54:					
	TCT	(B) (D) xi) S CAG	LOX OTF SEQUE GAG	CATIONER I	ON: 1 INFOR DESC CCC	l1 RMATI CRIPI ACG	1818 ION: TION: TTC	: SEQ TAC) ID	CAG	GAG					48
	TCT	(B) (D) xi) S	LOX OTF SEQUE GAG	CATIONER I	ON: 1 INFOR DESC CCC	l1 RMATI CRIPI ACG	1818 ION: TION: TTC	: SEQ TAC) ID	CAG	GAG					48
Met 1 ATC	TCT Ser	(B) (D) xi) S CAG Gln	LOX OTH SEQUE GAG Glu	CATION HER INCE AGG Arg 5	DN: INFOR	ACG Thr	L818 LON: TION: TTC Phe TAC	: SEQ TAC Tyr CAG	CGG Arg 10	CAG Gln CTG	GAG Glu TCT	Leu	Asn GTG	Lys 15 GGC	Thr	48
Met 1 ATC	TCT Ser	(B) (D) xi) S CAG Gln	LOX OTH SEQUE GAG Glu	CATION HER INCE AGG Arg 5	DN: INFOR	ACG Thr	L818 LON: TION: TTC Phe TAC	: SEQ TAC Tyr CAG	CGG Arg 10	CAG Gln CTG	GAG Glu TCT	Leu	Asn GTG	Lys 15 GGC	Thr	
Met 1 ATC Ile	TCT Ser TGG Trp	(B) (D) (Xi) S CAG Gln GAG Glu TAT	GAG GTG Val 20	CATIC HER I ENCE AGG Arg 5 CCC Pro	ON: INFOR	CRIPT ACG Thr CGT Arg	1818 ION: TION: TTC Phe TAC TYr	TAC Tyr CAG Gln 25	CGG Arg 10 AAC Asn	CAG Gln CTG Leu	GAG Glu TCT Ser	Leu CCA Pro	Asn GTG Val 30 ACG	Lys 15 GGC Gly	Thr TCT Ser	
Met 1 ATC Ile	TCT Ser TGG Trp	(B) (D) xi) S CAG Gln GAG Glu	GAG GTG Val 20	CATIC HER I ENCE AGG Arg 5 CCC Pro	ON: INFOR	CRIPT ACG Thr CGT Arg	1818 ION: TION: TTC Phe TAC TYr	TAC Tyr CAG Gln 25	CGG Arg 10 AAC Asn	CAG Gln CTG Leu	GAG Glu TCT Ser	Leu CCA Pro	Asn GTG Val 30 ACG	Lys 15 GGC Gly	Thr TCT Ser	96
Met 1 ATC Ile GGC Gly	TCT Ser TGG Trp GCC Ala	(B) (D) (A) (CAG GIN GAG GIU TAT Tyr 35 GCA	LOC OTHER SECULIAR SECU	CCC Pro	DESC CCC Pro GAG Glu GTG Val	11 CRIPI ACG Thr CGT Arg TGT Cys	1818 CON: TTC Phe TAC TYr GCT Ala 40 TCC	TAC Tyr CAG Gln 25 GCT Ala	CCA	CAG Gln CTG Leu GAC Asp	GAG Glu TCT Ser ACA Thr	CCA Pro AAA Lys 45	Asn GTG Val 30 ACG Thr	Lys 15 GGC Gly GGG Gly	Thr TCT Ser TTA Leu CAT	96
Met 1 ATC Ile GGC Gly	TCT Ser TGG Trp GCC Ala	(B) (D) (Xi) S CAG Gln GAG Glu TAT Tyr 35	LOC OTHER SECULIAR SECU	CCC Pro	DESC CCC Pro GAG Glu GTG Val	11 CRIPI ACG Thr CGT Arg TGT Cys	1818 CON: TTC Phe TAC TYr GCT Ala 40 TCC	TAC Tyr CAG Gln 25 GCT Ala	CCA	CAG Gln CTG Leu GAC Asp	GAG Glu TCT Ser ACA Thr	CCA Pro AAA Lys 45	Asn GTG Val 30 ACG Thr	Lys 15 GGC Gly GGG Gly	Thr TCT Ser TTA Leu CAT	96 144
Met 1 ATC Ile GGC Gly CGT Arg	TCT Ser TGG Trp GCC Ala GTG Val 50	(B) (D) (A) (A) (B) (D) (A) (A) (A) (B) (A) (B) (B) (B) (B) (A) (A) (A) (A) (A) (A) (A) (A) (A) (A	LOC OTT CONTROL OF CON	EATICE AGG Arg 5 CCC Pro	DESC CCC Pro GAG Glu GTG Val	11 CRIPI ACG Thr CGT Arg TGT Cys CTC Leu 55	1818 ION: TTON TTC Phe TAC Tyr GCT Ala 40 TCC Ser	TAC Tyr CAG Gln 25 GCT Ala AGA Arg	CGG Arg 10 AAC Asn TTT Phe CCA Pro	CAG Gln CTG Leu GAC Asp	GAG Glu TCT Ser ACA Thr CAG Gln 60	CCA Pro AAA Lys 45 TCC Ser	Asn GTG Val 30 ACG Thr ATC Ile	Lys 15 GGC Gly GGG Gly ATT Ile	Thr TCT Ser TTA Leu CAT His	96 144
Met 1 ATC Ile GGC Gly CGT Arg	TCT Ser TGG Trp GCC Ala GTG Val 50	(B) (D) xi) S CAG Gln GAG Glu TAT Tyr 35 GCA Ala	LOC OTT CONTROL OF CON	EATICE AGG Arg 5 CCC Pro	DESC CCC Pro GAG Glu GTG Val	11 CRIPI ACG Thr CGT Arg TGT Cys CTC Leu 55	1818 ION: TTON TTC Phe TAC Tyr GCT Ala 40 TCC Ser	TAC Tyr CAG Gln 25 GCT Ala AGA Arg	CGG Arg 10 AAC Asn TTT Phe CCA Pro	CAG Gln CTG Leu GAC Asp	GAG Glu TCT Ser ACA Thr CAG Gln 60	CCA Pro AAA Lys 45 TCC Ser	Asn GTG Val 30 ACG Thr ATC Ile	Lys 15 GGC Gly GGG Gly ATT Ile	Thr TCT Ser TTA Leu CAT His	96 144 192
Met 1 ATC Ile GGC Gly CGT Arg GCG Ala 65	TCT Ser TGG Trp GCC Ala GTG Val 50 AAA Lys	(B) (D) (A) (A) (B) (D) (A) (A) (A) (B) (A) (B) (B) (B) (B) (A) (A) (A) (A) (A) (A) (A) (A) (A) (A	LOC OTHER SEQUENTS OF THE SEQU	CATIC AGG Arg 5 CCC Pro TCT Ser AAG Lys TAC Tyr	DN: 1 INFOR DESC CCC Pro GAG Glu GTG Val AAG Lys AGA Arg 70	11 CRIPI ACG Thr CGT Arg TGT Cys CTC Leu 55 GAA Glu	1818 ION: TTC Phe TAC Tyr GCT Ala 40 TCC Ser CTG Leu	TAC Tyr CAG Gln 25 GCT Ala AGA Arg CGG Arg	CCA Pro	CAG Gln CTG Leu GAC Asp TTT Phe CTT Leu 75	GAG Glu TCT Ser ACA Thr CAG Gln 60 AAAA Lys	CCA Pro AAA Lys 45 TCC Ser CAT His	Asn GTG Val 30 ACG Thr ATC Ile ATG Met	Lys 15 GGC Gly GGG Gly ATT Ile AAA Lys	Thr TCT Ser TTA Leu CAT His CAT His 80 CTG	96 144 192

GAG Glu	GAA Glu	TTC Phe	AAT Asn 100	GAT Asp	GTG '	TAT Tyr	CTG Leu	GTG Val 105	ACC Thr	CAT His	CTC Leu	ATG Met	GGG Gly 110	GCA Ala	GAT Asp	336
CTG Leu	AAC Asn	AAC Asn 115	ATT Ile	GTG Val	AAA Lys	TGT Cys	CAG Gln 120	AAG Lys	CTT Leu	ACA Thr	GAT Asp	GAC Asp 125	CAT His	GTT Val	CAG Gln	384
TTC Phe	CTT Leu 130	ATC Ile	TAC Tyr	CAA Gln	ATT Ile	CTC Leu 135	CGA Arg	GGT Gly	CTA Leu	AAG Lys	ТАТ Туг 140	ATA Ile	CAT His	TCA Ser	GCT Ala	432
GAC Asp 145	ATA Ile	ATT Ile	CAC His	AGG Arg	GAC Asp 150	CTA Leu	AAA Lys	CCT Pro	AGT Ser	AAT Asn 155	CTA Leu	GCT Ala	GTG Val	AAT Asn	GAA Glu 160	480
GAC Asp	TGT Cys	GAG Glu	CTG Leu	AAG Lys 165	ATT Ile	CTG Leu	GAT Asp	TTT Phe	GGA Gly 170	CTG Leu	GCT Ala	CGG Arg	CAC His	ACA Thr 175	GAT Asp	528
GAT Asp	GAA Glu	ATG Met	ACA Thr 180	GGC Gly	TAC Tyr	GTG Val	GCC Ala	ACT Thr 185	AGG Arg	TGG Trp	TAC Tyr	AGG Arg	GCT Ala 190	Pro	GAG Glu	576
ATC Ile	ATG Met	CTG Leu 195	Asn	TGG Trp	ATG Met	CAT His	TAC Tyr 200	Asn	CAG Gln	ACA Thr	GTT Val	GAT Asp 205	Ile	TGC Trp	TCA Ser	624
GTG Val	GGA Gly 210	Cys	ATA	ATG Met	GCC Ala	GAG Glu 215	Leu	TTG Leu	ACT Thr	GGA Gly	AGA Arg 220	Thi	TTC	TT.	CCT Pro	672
GGT Gly 225	Thr	GAC Asp	CAT His	ATI	GAT Asp 230	Gln	TTC Lev	AAG Lys	CTC Lev	11e 235	Leu	A AGA	A CTY	C GT' u Vai	r GGA 1 Gly 240	720
ACC Thr	CCA Pro	Gly	GCI / Ala	GAC Glu 245	ı Leu	TTC	AAC Lys	AAA Lys	A ATC 5 Ile 250	e Ser	TC#	A GAG	G TC' u Se:	T GC. r Al	A AGA a Arg 5	768
AAC raA	TAT Tyr	ATT	CAC Glr 260	ı Sei	r TTC	ACT Thr	CAC	ATC Met 269	Pro	AA(Lys	S ATC	G AA t As	C TT n Ph 27	e Al	G AAT a Asn	816
GTA Val	A TTT	T ATT	e Gly	r GC0 / Ala	C AAT a Asr	CCC Pro	28	a Ala	r GTG a Val	C GAG l Ası	C TTO	G CT u Le 28	u Gl	G AA u Ly	G ATG	864
CT: Lei	r GTA u Val 290	l Le	G GAG	TC p Se:	A GAT	r AAG 5 Ly: 29:	s Ar	A AT	T AC.	A GCO	G GC a Al 30	a Gl	A GC n Al	C CI a Le	T GCA eu Ala	912
CA' Hi: 30	s Ala	С ТА в Ту	C TT	T GC e Al	T CAG a Gli 310	а Ту	C CA r Hi	C GA s As	T CC p Pr	T GA o As 31	p As	T GA p Gl	A CC u Pr	IA GT	rG GC0 al Ala 320	3.
G.A	T CC'	АТ Т	T GA	т са	G TC	C TT	T GA	A AG	C AG	G GA	C CI	rc cr	T A	ra G	AT GAG	G 1008

Asp	Pro	Tyr	Asp	Gln 325	Ser	Phe	Glu	Ser	Arg 330	Asp	Leu	Leu	Ile	Asp 335	Glu	
														CCA Pro		1056
														ACC Thr		1104
														CTG Leu		1152
														GGC Gly		1200
														ATC Ile 415		1248
		_												ACC Thr		1296
														AAG Lys		1344
														GAG Glu		1392
ACC																
														GAG Glu		1440
Thr 465 AAG	Ile	Phe GAG	Phe GGC	Lys GAC	Asp 470 ACC	Asp CTG	Gly GTG	Asn AAC	Tyr CGC	Lys 475 ATC	Thr GAG	Arg CTG	Ala AAG		Val 480 ATC	1440
Thr 465 AAG Lys	TTC Phe	Phe GAG Glu AAG	Phe GGC Gly GAG	GAC Asp 485 GAC	Asp 470 ACC Thr	Asp CTG Leu AAC	Gly GTG Val	Asn AAC Asn CTG	Tyr CGC Arg 490 GGG	Lys 475 ATC Ile CAC	Thr GAG Glu AAG	Arg CTG Leu CTG	Ala AAG Lys GAG	Glu GGC Gly	Val 480 ATC Ile	
Thr 465 AAG Lys GAC Asp	TTC Phe TTC Phe	Phe GAG Glu AAG Lys	GGC Gly GAG Glu 500 CAC	GAC Asp 485 GAC Asp	Asp 470 ACC Thr GGC Gly	Asp CTG Leu AAC Asn	Gly GTG Val ATC Ile	ASn AAC ASn CTG Leu 505	CGC Arg 490 GGG Gly	Lys 475 ATC Ile CAC His	Thr GAG Glu AAG Lys	Arg CTG Leu CTG Leu CAG	Ala AAG Lys GAG Glu 510	Glu GGC Gly 495 TAC	Val 480 ATC Ile AAC Asn	1488
Thr 465 AAG Lys GAC Asp TAC Tyr	TTC Phe TTC Phe AAC ASD	GAG Glu AAG Lys AGC Ser 515	GGC Gly GAG Glu 500 CAC His	GAC Asp 485 GAC Asp AAC Asn	ASP 470 ACC Thr GGC Gly GTC Val	Asp CTG Leu AAC Asn TAT Tyr	GTG Val ATC Ile ATC Ile 520	Asn AAC Asn CTG Leu 505 ATG Met	CGC Arg 490 GGG Gly GCC Ala	Lys 475 ATC Ile CAC His GAC Asp	GAG Glu AAG Lys AAG Lys	CTG Leu CTG Leu CAG Gln 525 GAC	Ala AAG Lys GAG Glu 510 AAG Lys	GGC Gly 495 TAC Tyr	Val 480 ATC Ile AAC Asn GGC Gly	1488 1536

GTG CTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG AGC
Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser
575

AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC GTG
Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val
580

ACC GCC GCC GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TAA

1821
Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys
595

(2) INFORMATION FOR SEQ ID NO:65:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 606 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:65:

Met Ser Gln Glu Arg Pro Thr Phe Tyr Arg Gln Glu Leu Asn Lys Thr 10 Ile Trp Glu Val Pro Glu Arg Tyr Gln Asn Leu Ser Pro Val Gly Ser 25 20 Gly Ala Tyr Gly Ser Val Cys Ala Ala Phe Asp Thr Lys Thr Gly Leu 40 Arg Val Ala Val Lys Lys Leu Ser Arg Pro Phe Gln Ser Ile Ile His 60 Ala Lys Arg Thr Tyr Arg Glu Leu Arg Leu Leu Lys His Met Lys His 70 **7**5 Glu Asn Val Ile Gly Leu Leu Asp Val Phe Thr Pro Ala Arg Ser Leu 85 90 Glu Glu Phe Asn Asp Val Tyr Leu Val Thr His Leu Met Gly Ala Asp 105 110 100 Leu Asn Asn Ile Val Lys Cys Gln Lys Leu Thr Asp Asp His Val Gln 120 125 115 Phe Leu Ile Tyr Gln Ile Leu Arg Gly Leu Lys Tyr Ile His Ser Ala 135 140 Asp Ile Ile His Arg Asp Leu Lys Pro Ser Asn Leu Ala Val Asn Glu 150 155 160 Asp Cys Glu Leu Lys Ile Leu Asp Phe Gly Leu Ala Arg His Thr Asp 165 170 175 Asp Glu Met Thr Gly Tyr Val Ala Thr Arg Trp Tyr Arg Ala Pro Glu 185 190 180 Ile Met Leu Asn Trp Met His Tyr Asn Gln Thr Val Asp Ile Trp Ser 195 200 205 Val Gly Cys Ile Met Ala Glu Leu Leu Thr Gly Arg Thr Leu Phe Pro 215 220 Gly Thr Asp His Ile Asp Gln Leu Lys Leu Ile Leu Arg Leu Val Gly 230 235 Thr Pro Gly Ala Glu Leu Leu Lys Lys Ile Ser Ser Glu Ser Ala Arg

250 245 Asn Tyr Ile Gln Ser Leu Thr Gln Met Pro Lys Met Asn Phe Ala Asn 260 265 Val Phe Ile Gly Ala Asn Pro Leu Ala Val Asp Leu Leu Glu Lys Met 280 285 275 Leu Val Leu Asp Ser Asp Lys Arg Ile Thr Ala Ala Gln Ala Leu Ala 290 295 300 His Ala Tyr Phe Ala Gln Tyr His Asp Pro Asp Asp Glu Pro Val Ala 315 310 Asp Pro Tyr Asp Gln Ser Phe Glu Ser Arg Asp Leu Leu Ile Asp Glu 330 335 325 Trp Lys Ser Leu Thr Tyr Asp Glu Val Ile Ser Phe Val Pro Pro 345 340 Leu Asp Gln Glu Glu Met Glu Ser Glu Asp Pro Pro Val Ala Thr Met 355 360 365 Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val 380 375 Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu 385 390 395 400 Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys 410 415 Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu 430 425 420 Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln 445 440 His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg 460 455 Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val 475 465 470 Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile 495 490 485 Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn 510 505 500 Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly 525 520 Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val 540 530 535 Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro 545 550 555 Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser 570 565 Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val 585 590 580 Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 600 595

- (2) INFORMATION FOR SEQ ID NO:66:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2913 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence

(B) LOCATION: 1...2910

(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:66:

														AAA Lys 15		48
														ACT Thr		96
AAT Asn	AAA Lys	GGG Gly 35	TCC Ser	TTA Leu	GTA Val	GCT Ala	CTT Leu 40	GGA Gly	TTC Phe	AGT Ser	GAT Asp	GGA Gly 45	CAG Gln	GAA Glu	GCC Ala	144
														ACA Thr		192
														AGG Arg		240
														CTT Leu 95		288
GTT Val	GCA Ala	CCA Pro	GGT Gly 100	TCT Ser	TCG Ser	AAA Lys	ACT Thr	GAA Glu 105	GCA Ala	GAT Asp	GTT Val	GAA Glu	CAA Gln 110	CAA Gln	GCT Ala	336
TTG Leu	ACT Thr	CTC Leu 115	Pro	GAT Asp	CTT Leu	GCA Ala	GAG Glu 120	Gln	TTT Phe	GCC	CCT Pro	CCT Pro 125	GAC	: ATT	GCC	384
		Lev					Val					Lys			CTG Leu	432
GAA Glu 145	C)'s	TC.	ACT Thr	CTA	TAC Tyr 150	Arg	ACA Thr	CAG Gln	AGC Ser	TCC Ser 155	Ser	AAC Asn	CTC	GCA 1 Ala	GAA Glu 160	480
TTA Leu	. CGA ı Arg	CAC	G CTI n Leu	CTI Let 165	ı Asp	TGI Cys	GAT Asp	Thr	CCC Pro	Sei	GTC Val	G GAC L Asp	TTC Lev	G GAZ 1 Glu 175	A ATG 1 Met	528
				Va]					a Phe					ı Lev	G GAC J Asp	576
			n Pro					a Ala					ı Me		r TCT e Ser	624
TT	A GC1	r cc.	A GAJ	A GT	A CA	A AGO	10	C GA	A GA	AT A	TAT	T CA	G CT	TT A	g AAG	672

Leu	Ala 210	Pro	Glu	Val	Gln	Ser 215	Ser	Glu	Glu	Tyr	Ile 220	Gln	Leu	Leu	Lys	
								CCT Pro								720
								AAG Lys								768
								TCT Ser 265								816
								GAT Asp								864
								GAA Glu								912
								AAA Lys								960
								CAA Gln								1008
								GAA Glu 345								1056
								TCT Ser								1104
								AAT Asn								1152
								TCT Ser								1200
								CGG Arg								1248
								CTT Leu 425								1296
_		_	_					AAT Asn								1344

TTA Leu	CAT His 450	GAA Glu	TAT Tyr	AAC Asn	ACT Thr	CAG Gln 455	TTT Phe	CAA Gln	GAA Glu	AAA Lys	AGT Ser 460	CGA Arg	GAA Glu	TAT Tyr	GAT Asp	1392
	TTA Leu															1440
AGG Arg	ACA Thr	GCT Ala	ATT Ile	GAA Glu 485	GCA Ala	TTT Phe	AAT Asn	GAA Glu	ACC Thr 490	ATA Ile	AAA Lys	ATA Ile	TTT Phe	GAA Glu 495	GAA Glu	1488
CAG Gln	TGC Cys	CAG Gln	ACC Thr 500	CAA Gln	GAG Glu	CGG Arg	TAC Tyr	AGC Ser 505	AAA Lys	GAA Glu	TAC Tyr	ATA Ile	GAA Glu 510	AAG Lys	TTT Phe	1536
	CGT Arg															1584
GAT Asp	AAG Lys 530	TTG Leu	AAG Lys	TCT Ser	CGA Arg	ATC Ile 535	AGT Ser	GAA Glu	ATT Ile	ATT	GAC Asp 540	AGT Ser	AGA Arg	AGA Arg	AGA Arg	1632
	GAA Glu					Lys					Tyr					1680
AAA Lys	. CGT . Arg	ATG Met	AAC Asn	AGC Ser 565	Ile	AAA Lys	CCA Pro	GAC Asp	CTT Leu 570	Ile	CAG Gln	CTG Leu	AGA Arg	AAG Lys 575	Thr	1728
AGA Arg	GAC Asp	CAA Glr	TAC Tyr 580	Leu	ATG Met	TGG Trp	TTG	ACT Thr 585	Gln	. AAA . Lys	GGT Gly	GTT Val	CGG Arg 590	Gln	AAG Lys	1776
AAC Lys	TTG Leu	AAC Asr 595	Glu	TGG Trp	TTG	GGC Gly	raa naa 000	ı Glu	AAC Asn	ACI Thr	GAA Glu	GAC Asp 605	Glr	TAT	TCA Ser	1824
CTC	GTC Val	Glu	A GAT L Asp	GAT Asp	GAA Glu	GAT Asp 615	Lei	G CCC	CAT His	CAT His	GAT S ASP 620	r GAC o Glu	G AAC 1 Lys	G ACA	TGG Trp	1872
AAS ASI 625	n Val	r GGZ L Gly	A AGO y Ser	AGC Ser	AAC Asr 630	n Arg	AA Z	C AAA	A GCT	GAA Glu 635	ı Ası	CTC n Lev	TTX	G CGA	A GGG g Gly 640	1920
AA(Ly:	G CGA s Arg	A GA' J Ası	r GG(p Gl)	C ACT Y Thi 645	r Phe	r CTT e Leu	GT(C CGC 1 Arg	GAG GGL GSG	se:	C AG' r Se:	r AAJ r Ly:	A CAG	G GG(n Gly 65!	TGC Y Cys	1968
TA' Ty:	r GCC r Ala	TG a Cy	C TC' s Se:	r Val	A GTY 1 Val	G GTC l Val	G GAG	C GG(p Gl ₂ 66!	y Gl	A GT. ı Va	A AA(l Ly:	G CA' s Hi	T TG' s Cy 67	s Va	C ATA l Ile	2016
AA	C AA	A AC	A GC.	a ac	T GG	C TAT	r GG	C TT	r GC	C GA	G CC	C TA	T AA	C TT	G TAC	2064

Asn	Lys	Thr 675	Ala	Thr	Gly	Tyr	Gly 680	Phe	Ala	Glu	Pro	Tyr 685	Asn	Leu	Tyr	
AGC Ser	TCT Ser 690	CTG Leu	AAA Lys	GAA Glu	CTG Leu	GTG Val 695	CTA Leu	CAT His	TAC Tyr	CAA Gln	CAC His 700	ACC Thr	TCC Ser	CTT Leu	GTG Val	2112
CAG Gln 705	CAC His	AAC Asn	GAC Asp	TCC Ser	CTC Leu 710	AAT Asn	GTC Val	ACA Thr	CTA Leu	GCC Ala 715	TAC Tyr	CCA Pro	GTA Val	TAT Tyr	GCA Ala 720	2160
CAG Gln	CAG Gln	AGG Arg	CGA Arg	CAG Gln 725	GAT Asp	CCA Pro	CCG Pro	GTC Val	GCC Ala 730	ACC Thr	ATG Met	GTG Val	AGC Ser	AAG Lys 735	GGC Gly	2208
GAG Glu	GAG Glu	CTG Leu	TTC Phe 740	ACC Thr	GGG Gly	GTG Val	GTG Val	CCC Pro 745	ATC Ile	CTG Leu	GTC Val	GAG Glu	CTG Leu 750	GAC Asp	GGC Gly	2256
GAC Asp	GTA Val	AAC Asn 755	GGC Gly	CAC His	AAG Lys	TTC Phe	AGC Ser 760	GTG Val	TCC Ser	GGC Gly	GAG Glu	GGC Gly 765	GAG Glu	GGC Gly	GAT Asp	2304
GCC Ala	ACC Thr 770	TAC Tyr	GGC GJY	AAG Lys	CTG Leu	ACC Thr 775	CTG Leu	AAG Lys	TTC Phe	ATC Ile	TGC Cys 780	Thr	ACC Thr	GGC Gly	AAG Lys	2352
CTG Leu 785	Pro	GTG Val	CCC Pro	TGG Trp	CCC Pro 790	ACC Thr	CTC Leu	GTG Val	ACC Thr	ACC Thr 795	Leu	ACC Thr	TAC Tyr	GGC Gly	GTG Val 800	2400
CAG Gln	TGC Cys	TTC Phe	AGC Ser	CGC Arg 805	Tyr	CCC Pro	GAC Asp	CAC	ATG Met 810	Lys	CAG Gln	CAC His	GAC Asp	TTC Phe 815	TTC Phe	2448
AAG Lys	TCC Ser	GCC Ala	ATG Met 820	CCC	GAA Glu	GGC Gly	TAC	GTC Val 825	CAG Gln	GAC Glu	G CGC	ACC Thr	ATC 11e	Phe	TTC Phe	2496
AAG Lys	GAC Asp	GAC Asp 835	Gly	AAC Asn	TAC Tyr	AAG Lys	ACC Thr 840	Arg	GCC Ala	GAC Glu	GTC 1 Val	AAC Lys 845	Ph€	GAC Glu	GGC Gly	2544
GAC Asp	ACC Thr 850	Lei	GTG Val	AAC Asr	CGC Arg	ATC Ile	Glu	CTG Leu	AAC Lys	GG(Gly	ATC 11e 860	e Asp	TTC Phe	AAC E Lys	G GAG Glu	2592
GAC Asp 865	Gly	AA : reA :	ATC	CTC	G GGG 1 G1y 870	His	: AAC : Lys	CTG Lev	GAC Glu	TAC 1 Tyr 87!	c Ast	TA(C AAC C ASI	a AGC n Sei	CAC His 880	2640
AA IzA	GTC n Val	TAT Tyl	r ATC	ATC Met	Ala	GAC Asp	AAC Lys	G CAC	AA0 Lys 890	s Ası	C GG(n Gl	C ATG	C AAG e Lyn	G GT0 s Val 89!	G AAC l Asn 5	2688
TTO	C AAG e Lys	S ATO	C CGC e Arg	g Hi	C AAC s Asr	TATO	GAG Glu	G GAC Asp 909	Gl	C AG y Se	C GTY r Va	G CA	G CT n Le 91	u Al	c GAC a Asp	2736

CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC CCC GTG CTG CCC 2784

His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro 925

GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG AGC AAA GAC CCC AAC Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn 930

GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC GTG ACC GCC GCC GGC GCC GGC GLu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly 955

ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TAA

11e Thr Leu Gly Met Asp Glu Leu Tyr Lys 965

(2) INFORMATION FOR SEQ ID NO:67:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 970 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:67:

Met Ser Ala Glu Gly Tyr Gln Tyr Arg Ala Leu Tyr Asp Tyr Lys Lys 1 5 Glu Arg Glu Glu Asp Ile Asp Leu His Leu Gly Asp Ile Leu Thr Val 3.0 25 20 Asn Lys Gly Ser Leu Val Ala Leu Gly Phe Ser Asp Gly Gln Glu Ala 40 Arg Pro Glu Glu Ile Gly Trp Leu Asn Gly Tyr Asn Glu Thr Thr Gly 60 55 Glu Arg Gly Asp Phe Pro Gly Thr Tyr Val Glu Tyr Ile Gly Arg Lys 75 70 Lys Ile Ser Pro Pro Thr Pro Lys Pro Arg Pro Pro Arg Pro Leu Pro 90 85 Val Ala Pro Gly Ser Ser Lys Thr Glu Ala Asp Val Glu Gln Gln Ala 105 100 Leu Thr Leu Pro Asp Leu Ala Glu Gln Phe Ala Pro Pro Asp Ile Ala 120 125 Pro Pro Leu Leu Ile Lys Leu Val Glu Ala Ile Glu Lys Lys Gly Leu 130 135 140 Glu Cys Ser Thr Leu Tyr Arg Thr Gln Ser Ser Ser Asn Leu Ala Glu 150 155 Leu Arg Gln Leu Leu Asp Cys Asp Thr Pro Ser Val Asp Leu Glu Met 170 165 Ile Asp Val His Val Leu Ala Asp Ala Fhe Lys Arg Tyr Leu Leu Asp 180 190 185 Leu Pro Asn Pro Val Ile Pro Ala Ala Val Tyr Ser Glu Met Ile Ser 2.05 200 195 Leu Ala Pro Glu Val Gln Ser Ser Glu Glu Tyr Ile Gln Leu Leu Lys

	210					215					220				
Lys 225	Leu	Ile	Arg	Ser	Pro 230	Ser	Ile	Pro	His	Gln 235	Tyr	Trp	Leu	Thr	Leu 240
Gln	Tyr	Leu	Leu	Lys 245	His	Phe	Phe	Lys	Leu 250	Ser	Gln	Thr	Ser	Ser 255	Lys
Asn	Leu	Leu	Asn 260	Ala	Arg	Val	Leu	Ser 265	Glu	Ile	Phe	Ser	Pro 270	Met	Leu
Phe	Arg	Phe 275	Ser	Ala	Ala	Ser	Ser 280	qzA	Asn	Thr	Glu	Asn 285	Leu	Ile	Lys
	290					295			Trp		300				
305					310				Pro	315					320
				325					Asn 330					335	
			340					345	Lys				350		
		355					360		Thr			365			
	370					375			Asn		380				
His 385	Arg	qzA	Gly	Lys	Tyr 390	Gly	Phe	Ser	Asp	Pro 395	Leu	Thr	Phe	Ser	Ser 400
	Val	Glu	Leu	11e 405	Asn	His	Tyr	Arg	Asn 410	Glu	Ser	Leu	Ala	Gln 415	Tyr
			420					425	Tyr				430		
		435					440		Ile			445			
	450					455			Glu		460				
465					470				Ser	475					480
				485					Thr 490					495	
	_		500					505	Lys				510		
		515					520		Gln			525			
	530					535			Ile		540				
545					550				Ala	555					560
				565					Leu 570					575	
			580					585	Gln				590		
		595					600		Asn			605			
	610					615			His		620				
625					630				Ala	635					640
				645					Glu 650					655	
			660					665					670		Ile
Asn	Lys	Thr	Ala	Thr	Gly	Tyr	Gly	Phe	Ala	Glu	Pro	Tyr	Asn	Leu	Tyr

680 Ser Ser Leu Lys Glu Leu Val Leu His Tyr Gln His Thr Ser Leu Val 700 690 695 Gln His Asn Asp Ser Leu Asn Val Thr Leu Ala Tyr Pro Val Tyr Ala 715 710 Gln Gln Arg Arg Gln Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly 730 735 725 Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly 745 740 Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Asp 755 760 Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys 775 780 Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val 790 795 Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe 805 810 Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe 820 825 830 Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly 835 840 845 Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu 850 855 860 Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His 865 870 875 Asn Val Tyr Ile Met Ala Asp Lys Glr Lys Asn Gly Ile Lys Val Asn 885 890 895 Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp 905 910 His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro 915 920 925 Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn 930 935 940 Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly 945 950 955 Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 965

(2) INFORMATION FOR SEQ ID NO:68:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1788 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: CDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...1785
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:68:

ATG GGC AAC GCC GCC GCC AAG AAG GGC AGC GAG CAG GAG AGC GTG
Met Gly Asn Ala Ala Ala Ala Lys Lys Gly Ser Glu Gln Glu Ser Val
1 5 10 15

						GCC Ala										96	
						GCC Ala										144	
						TTT Phe 55						_				192	
						GCC Ala							_			240	
						GAG Glu										288	
						TTC Phe										336	
						ATG Met										384	
						CGG Arg 135						_				432	
						ATC Ile										480)
		-				GAC Asp										528	t
						GTG Val				_	_	_			_	57€	j
						TTG Leu									CCC Pro	624	Į
						GGC Gly 215									GCT Ala	672	?
						GAG Glu									TTC Phe 240	720)
GCT	GAC	CAG	CCT	ATC	CAG	ATC	TAT	GAG	AAA	ATC	GTC	TCT	GGG	AAG	GTG	768	3

Ala	Asp	Gln	Pro	Ile 245	Gln	Ile	Tyr		Lys 250	Ile	Val	Ser	Gly	Lys 255	Val	
									TTG Leu							816
									TTT Phe							864
GTC Val	AAT Asn 290	GAC Asp	ATC Ile	AAG Lys	AAC Asn	CAC His 295	AAG Lys	TGG Trp	TTT Phe	GCC Ala	ACG Thr 300	ACT Thr	GAC Asp	TGG Trp	ATT Ile	912
GCC Ala 305	ATC Ile	TAT Tyr	CAG Gln	AGA Arg	AAG Lys 310	GTG Val	GAA Glu	GCT Ala	CCC Pro	TTC Phe 315	ATA Ile	CCA Pro	AAG Lys	TTT Phe	AAA Lys 320	960
									GAC Asp 330							1008
CGG Arg	GTC Val	TCC Ser	ATC Ile 340	Asn	GAG Glu	AAG Lys	TGT Cys	GGC Gly 345	AAG Lys	GAG Glu	TTT Phe	ACT Thr	GAG Glu 350	Phe	GGG	1056
CGC Arg	GCC Ala	ATG Met 355	Ser	AAA Lys	GGA Gly	GAA Glu	GAA Glu 360	Leu	TTC Phe	ACT Thr	GGA Gly	GTT Val 365	Val	CCA Pro	ATT Ile	1104
CTT Leu	GTT Val	Glu	TTA Leu	GAT Asp	GGC Gly	GAT Asp 375	Val	TAA ' naA .	GGG Gly	CA.	A AAA 1 Lys 380	Phe	TCT Ser	GT1	AGT Ser	1152
GG# G1y 385	/ Glu	GGT Gly	GAA Glu	GGT Gly	GAT Asp 390	Ala	ACA Thr	TAC	GGA Gly	AA. Lys 395	. Lei	ACC 1 Thi	CT:	r AA/ Lys	A TTT s Phe 400	1200
ATT Ile	r TGC e Cys	ACT	C ACT	r GGC Gly 405	/ Lys	CTA	CCT Pro	r GT1 o Val	CCA Pro 410	Tr	G CCA Pro	A ACC	CT Lei	r GTG u Val	ACT Thr	1248
AC:	r CTC	ACT Thi	TA:	c Gly	r GT7 / Val	CAA Glr	TGC Cys	TTM S Phe 425	e Ser	AG Arg	A TAG	c cci	A GA' o Asi 43	p Hi	r ATG s Met	1296
AA Ly:	A CAC	G CAT n His 435	s As	TT.	r TT(∋ Phe	C AAC B Lys	G AG S Se 440	r Ala	ATC a Met	CC Pr	C GAI o Gli	A GG u G1; 44	у Ту	T GT r Va	A CAG l Gln	1344
GA.	A AGA u Arg 45	g Th	r AT.	A TT e Ph	r TAC e Ty:	2 AA/ c Lys 45!	s As	T GA(p Asj	c GG(p Gly	G AA Y As	C TAG n Ty. 46	r Ly	G AC s Th	A CG r Ar	T GCT g Ala	1392
GA G1 46	u Va	C AA	G TT s Ph	T GA e Gl	A GG' u Gl; 47	y As	T AC	C CT r Le	T GT u Va	T AA l As 47	n Ar	A AT g Il	C GA e Gl	G TT u Le	A AAA u Lys 480	1440

GGT Gly	ATT Ile	GAT Asp	TTT Phe	AAA Lys 485	GAA Glu	GAT Asp	GGA Gly	AAC Asn	ATT Ile 490	CTT Leu	GGA Gly	CAC His	AAA Lys	ATG Met 495	GAA Glu	1488
					CAT His											1536
AAT Asn	GGC Gly	ATC Ile 515	AAA Lys	GTT Val	AAC Asn	TTC Phe	AAA Lys 520	ATT Ile	AGA Arg	CAC His	AAC Asn	ATT Ile 525	AAA Lys	GAT Asp	GGA Gly	1584
AGC Ser	GTT Val 530	CAA Gln	TTA Leu	GCA Ala	GAC Asp	CAT His 535	TAT Tyr	CAA Gln	CAA Gln	AAT Asn	ACT Thr 540	Pro	ATT	GGC Gly	GAT Asp	1632
GGC Gly 545	CCT Pro	GTC Val	CTT Leu	TTA Leu	CCA Pro 550	GAC Asp	AAC Asn	CAT	TAC Tyr	CTG Leu 555	TCC Ser	ACG Thr	CAA Gln	TCT	GCC Ala 560	1680
CTT Leu	TCC Ser	AAA Lys	GAT Asp	CCC Pro 565	Asn	GAA Glu	AAG Lys	AGA Arg	GAT Asp 570	His	ATG Met	ATC	CTT	CTT Leu 575	GAG Glu	1728
TTT Phe	GTA Val	ACA Thr	GCT Ala 580	Ala	GGG Gly	ATT	ACA Thr	CAT His 585	Gly	ATG Met	GAT Asp	GAA Glu	CTA Leu 590	Тух	AAA Lys	1776
	-	GAG Glu 595														1788

(2) INFORMATION FOR SEQ ID NO:69:

(i) SEQUENCE CHAFACTERISTICS:

(A) LENGTH: 595 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:69:

Met Gly Asn Ala Ala Ala Lys Lys Gly Ser Glu Gln Glu Ser Val 10 1 5 Lys Glu Phe Leu Ala Lys Ala Lys Glu Asp Phe Leu Lys Lys Trp Glu 25 20 Asp Pro Ser Gln Asn Thr Ala Gln Leu Asp Gln Phe Asp Arg Ile Lys 40 35 Thr Leu Gly Thr Gly Ser Phe Gly Arg Val Met Leu Val Lys His Lys 55 60 Glu Ser Gly Asn His Tyr Ala Met Lys Ile Leu Asp Lys Gln Lys Val 75 70 Val Lys Leu Lys Gln Ile Glu His Thr Leu Asn Glu Lys Arg Ile Leu

				85					90					95	
			100		Pro			105					110		
		115			Tyr		120					125			
	130				Arg	135					140				
Arg 145	Phe	Tyr	Ala	Ala	Gln 150	Ile	Val	Leu	Thr	Phe 155	Glu	Tyr	Leu	His	Ser 160
Leu	Asp	Leu	Ile	Tyr 165	Arg	Asp	Leu	Lys	Pro 170	Glu	Asn	Leu	Leu	Ile 175	Asp
Gln	Gln	Gly	Tyr 180		Gln	Val	Thr	Asp 185		Gly	Phe	Ala	Lys 190	Arg	Val
Lys	Gly	Arg 195		Trp	Thr	Leu	Cys 200	Gly	Thr	Pro	Glu	Tyr 205	Leu	Ala	Pro
Glu	Ile 210	Ile	Leu	Ser	Lys	Gly 215	Tyr	Asn	Lys	Ala	Val 220	Asp	Trp	Trp	Ala
225	Gly	Val			Tyr 230					235					240
				245	Gln				250					255	
			260		Phe			265					270		
		275	,		Leu		280					285			
	290)			Asn	295					300				
305					Lys 310					315					320
				325	,				330					335	
			340					345					350)	e Gly
		359	5				360					365			lle
	370)				375					380)			Ser
385)				390	•				395					Phe 400
				405	5				410)				415	
			420)				425	,				430)	Met
		43	5				440)				445	5		l Gln
	45	0				455	5				460)			g Ala
465	5				470)				475	5				480
				48	5				490	C				49	
_			50	0				50	5				51	0	o Lys
		51	5				520)				52	5		p Gly
	53	0				53	5				54	0			y Asp
Gl	y Pr	o Va	1 Le	u Le	u Pro	z Ası	e Ası	n Hi	s Ty:	r Le	ı Se	r Th	r Gl	n Se	r Ala

545 550	555 560
Leu Ser Lys Asp Pro Asn Glu Lys 565	Arg Asp His Met Ile Leu Leu Glu 570 575
Phe Val Thr Ala Ala Gly Ile Thr 580	His Gly Met Asp Glu Leu Tyr Lys 585 590
Pro Gln Glu 595	
(2) INFORMATION FOR SEC	Q ID NO:70:
(i) SEQUENCE CHARACTERIST: (A) LENGTH: 2181 base pa (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	airs
(ii) MOLECULE TYPE: cDNA (ix) FEATURE:	
(A) NAME/KEY: Coding So (B) LOCATION: 12178	equence
(D) OTHER INFORMATION:	
(xi) SEQUENCE DESCRIPTION	: SEQ ID NO:70:
ATG AGC GAC GTG GCT ATT GTG AAG Met Ser Asp Val Ala Ile Val Lys 1 5	GAG GGT TGG CTG CAC AAA CGA GGG 48 Glu Gly Trp Leu His Lys Arg Gly 10 15
GAG TAC ATC AAG ACC TGG CGG CCA Glu Tyr Ile Lys Thr Trp Arg Pro 20	CGC TAC TTC CTC CTC AAG AAT GAT 96 Arg Tyr Phe Leu Leu Lys Asn Asp 25 30
GGC ACC TTC ATT GGC TAC AAG GAG Gly Thr Phe Ile Gly Tyr Lys Glu 35	G CGG CCG CAG GAT GTG GAC CAA CGT 144 1 Arg Pro Gln Asp Val Asp Gln Arg 45
GAG GCT CCC CTC AAC AAC TTC TCT Glu Ala Pro Leu Asn Asn Phe Ser 50	r GTG GCG CAG TGC CAG CTG ATG AAG 192 r Val Ala Gln Cys Gln Leu Met Lys 60
ACG GAG CGG CCC CGG CCC AAC ACC Thr Glu Arg Pro Arg Pro Asn Thr 65	TTC ATC ATC CGC TGC CTG CAG TGG 240 Phe Ile Ile Arg Cys Leu Gln Trp 75 80
ACC ACT GTC ATC GAA CGC ACC TTC Thr Thr Val Ile Glu Arg Thr Phe 85	C CAT GTG GAG ACT CCT GAG GAG CGG 288 E His Val Glu Thr Pro Glu Glu Arg 90 95
GAG GAG TGG ACA ACC GCC ATC CAC Glu Glu Trp Thr Thr Ala Ile Glr 100	G ACT GTG GCT GAC GGC CTC AAG AAG 336 n Thr Val Ala Asp Gly Leu Lys Lys 105 110
CAG GAG GAG GAG ATG GAC TTG Gln Glu Glu Glu Met Asp Pho 115	C CGG TCG GGC TCA CCC AGT GAC AAC 384 e Arg Ser Gly Ser Pro Ser Asp Asn 125
TCA GGG GCT GAA GAG ATG GAG GTV	G TCC CTG GCC AAG CCC AAG CAC CGC 432

Ser	Gly 130	Ala	Glu	Glu	Met	Glu 135	Val	Ser	Leu	Ala	Lys 140	Pro	Lys	His	Arg	
GTG Val 145	ACC Thr	ATG Met	AAC Asn	GAG Glu	TTT Phe 150	GAG Glu	TAC Tyr	CTG Leu	AAG Lys	CTG Leu 155	CTG Leu	GGC Gly	AAG Lys	GGC Gly	ACT Thr 160	480
TTC Phe	GGC Gly	AAG Lys	GTG Val	ATC Ile 165	CTG Leu	GTG Val	AAG Lys	GAG Glu	AAG Lys 170	GCC Ala	ACA Thr	GGC Gly	CGC Arg	TAC Tyr 175	TAC Tyr	528
GCC Ala	ATG Met	AAG Lys	ATC Ile 180	CTC Leu	AAG Lys	AAG Lys	GAA Glu	GTC Val 185	ATC Ile	GTG Val	GCC Ala	AAG Lys	GAC Asp 190	GAG Glu	GTG Val	576
					GAG Glu											624
TTC Phe	CTC Leu 210	ACA Thr	GCC Ala	CTG Leu	AAG Lys	TAC Tyr 215	TCT Ser	TTC Phe	CAG Gln	ACC Thr	CAC His 220	GAC Asp	CGC	CTC Leu	TGC Cys	672
TTT Phe 225	Val	ATG Met	GAG Glu	TAC Tyr	GCC Ala 230	AAC Asn	GGG Gly	GGC Gly	GAG Glu	CTG Leu 235	Phe	TTC Phe	CAC	CTG Leu	TCC Ser 240	720
CGG Arg	GAA Glu	CGT Arg	GTG Val	TTC Phe 245	Ser	GAG Glu	GAC Asp	CGG	GCC Ala 250	Arg	TTC Phe	ТАТ Тух	G17	GCT Ala 255	GAG Glu	768
ATT	GTG Val	TCA Ser	GCC Ala 260	Leu	GAC Asp	TAC Tyr	CTG Leu	CAC His 265	Ser	GAC Glu	AAG Lys	AAC Asn	GTC Val 270	l Val	TAC Tyr	816
CGG Arg	GAC Asp	CTC Lev 275	ı Lys	CTC	G GAG	AAC Asn	CTC Leu 280	Met	CTG Leu	GAC Asp	C AAC D Lys	GAC S Asp 285	Gl	G CAC Y His	ATT Ile	864
AAC Lys	3 ATC 3 Ile 290	Thi	A GAC	TTC Phe	GGG Gly	CTG Leu 295	Суз	AAG Lys	GAC Glu	GGG Gly	OTA E 11 y 300	e Lys	G GAG S Asi	c GG p Gly	r GCC / Ala	912
ACC Th:	e Met	G AAC Lys	G ACC	TT:	T TGC e Cys 310	s Gly	ACA Thi	A CCT	GAC Glu	TAC 1 TY: 31	r Le	g GC0 u Ala	C CC a Pr	C GAG o Gl	G GTG u Val 320	960
CT(Let	G GAG	G GAG	AA C	r GAG n Ası 32!	р Туз	c GGC	CG Arg	r GCA g Ala	A GTO a Val	l As	C TG	G TG p Tr	G GG p Gl	G CT y Le 33	g ggc u Gly 5	1008
GT(Va)	G GTY	C ATG	G TAC t Ty:	r Gl	G ATG	G ATC	TG(G GG' S Gl ₁	y Arg	C CT	G CC u Pr	C TT o Ph	С ТА е Ту 35	r As	C CAG n Gln	1056
GA As	C CA' p Hi	T GA s Gl 35	u Ly	G CT s Le	T TT u Ph	T GAG e Gli	G CT Le 36	u Il	C CT	C AT u Me	G GA t Gl	G GA u G1 36	u Il	C CG .e Ar	C TTC g Phe	1104

									TCC Ser							1152
									GGG Gly							1200
									GGT Gly 410							1248
									AAG Lys							1296
									TTC Phe							1344
									ATG Met							1392
									TAC Tyr							1440
									AGC Ser 490							1488
									CTG Leu							1536
									GAG Glu							1584
									ACC Thr							1632
									TAC Tyr							1680
									GAC Asp 570							1728
									ATC Ile							1776
AAC	TAC	AAG	ACC	CGC	GCC	GAG	GTG	AAG	TTC	GAG	GGC	GAC	ACC	CTG	GTG	1824

Asn	Tyr	Lys 595	Thr	Arg	Ala	Glu	Val 600	Lys	Phe	Glu	Gly	Asp 605	Thr	Leu	Val	
												GAC Asp				1872
CTG Leu 625	GGG Gly	CAC His	AAG Lys	CTG Leu	GAG Glu 630	TAC Tyr	AAC Asn	TAC Tyr	AAC Asn	AGC Ser 635	CAC His	AAC Asn	GTC Val	TAT Tyr	ATC Ile 640	1920
ATG Met	GCC Ala	GAC Asp	AAG Lys	CAG Gln 645	AAG Lys	AAC Asn	GGC Gly	ATC Ile	AAG Lys 650	GTG Val	AAC Asn	TTC Phe	AAG Lys	ATC Ile 655	CGC Arg	1968
												CAC His				2016
												GAC Asp 685				2064
		Thr										Glu			GAT Asp	2112
CAC His 705	ATG Met	GTC Val	CTG Leu	CTG Leu	GAG Glu 710	Phe	GTG Val	ACC Thr	GCC	GCC Ala 715	Gly	ATC	ACT Thr	CTC	GGC Gly 720	2160
			CTG Leu		Lys											2181

(2) INFORMATION FOR SEQ ID NO:71:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 726 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:71:

 Met
 Ser
 Asp
 Val
 Ala
 Ile
 Val
 Lys
 Glu
 Glu
 Gly
 Trp
 Leu
 His
 Lys
 Arg
 Gly

 Glu
 Tyr
 Ile
 Lys
 Thr
 Trp
 Arg
 Pro
 Arg
 Tyr
 Phe
 Leu
 Leu
 Lys
 Asn
 Asp

 Gly
 Thr
 Phe
 Ile
 Gly
 Tyr
 Lys
 Glu
 Arg
 Pro
 Gln
 Asp
 Val
 Asp
 Gln
 Arg

 Glu
 Ala
 Pro
 Leu
 Asn
 Asn
 Asn
 Phe
 Ser
 Val
 Ala
 Gln
 Leu
 Met
 Lys

 Glu
 Ala
 Pro
 Leu
 Asn
 Asn
 Asn
 Phe
 Ser
 Val
 Ala
 Gln
 Leu
 Met
 Lys

 Glu
 Arg
 Pro
 Arg
 Pro
 Asn
 Thr
 Fhe
 Ile
 Ile
 Ile
 Asn
 Cys

65					70					75					80
Thr '				85					90					95	
Glu			100					105					110		
Gln (115					120					125			
	130					135		Ser			140				
Val 145	Thr	Met	Asn	Glu	Phe 150	Glu	Tyr	Leu	Lys	Leu 155	Leu	Gly	Lys	Gly	Thr 160
	Gly	Lys	Val	Ile 165		Val	Lys	Glu	Lys 170	Ala	Thr	Gly	Arg	Тут 175	Tyr
			180					Val 185					190		
		195					200	Val				205			
	210					215		Phe			220				
Phe 225	Val	Met	Glu	Tyr	Ala 230	Asn	Gly	Gly	Glu	Leu 235	Phe	Phe	Hıs	Leu	Ser 240
Arg	Glu	Arg	Val	Phe 245	Ser	Glu	Asp	Arg	Ala 250		Phe	Tyr	Gly	Ala 255	Glu
			260					His 265					270		
		275					280					285			
	290					295		Lys			300				
305					310			Pro		315					320
				325					330	+				335	
			340					345					350	1	Gln
_		355					360	1				365			Phe
	370					375					380				Leu
385					390					395					400
Glu	Ile	Met	Gln	His 405		Phe	Phe	e Ala	Gl ₃ 410		· Val	Trp	Glr	415	: Val
			420					425					430)	Glu
		435	ı				440)				445	ò		e Thr
	450					455	,				460)			Glu
465					470)				475	5				480
				485	5				490)				49	
			500)				505	5				51	0	n Gly
		515	5				520	С				52	5		r Gly
Lys	Leu	Thi	Lev	ı Lys	s Phe	e Ile	э Су:	s Thi	Th:	r Gly	y Lys	s Le	u Pr	o Va	l Pro

	530					535					540					
Trp 545	Pro	Thr	Leu	Val	Thr 550	Thr	Leu	Thr	Tyr	Gly 555	Val	Gln	Cys	Phe	Ser 560	
	Tyr	Pro	Asp	His 565	Met	Lys	Gln	His	Asp 570	Phe	Phe	Lys	Ser	Ala 575	Met	
Pro	Glu	Gly	Тут 580	Val	Gln	Glu	Arg	Thr 585	Ile	Phe	Phe	Lys	Asp 590	Asp	Gly	
Asn	Tyr	Lys 595	Thr	Arg	Ala	Glu	Val 600	Lys	Phe	Glu	Gly	Asp 605	Thr	Leu	Val	
Asn	Arg 610	Ile	Glu	Leu	Lys	Gly 615	Ile	Asp	Phe	Lys	Glu 620	Asp	Gly	Asn	Ile	
625	Gly				630					635					640	
	Ala			645					650					655		
	Asn		660					665					670			
Asn	Thr	Pro 675	Ile	Gly	Asp	Gly	Pro 680		Leu	Leu	Pro	Asp 685	Asn	His	Tyr	
Leu	Ser 690		Gln	Ser	Ala	Leu 695	Ser	Lys	Asp	Pro	Asn 700		Lys	Arg	Asp	
His 705	Met	Val	Leu	Leu	Glu 710		Val	Thr	Ala	Ala 715		Ile	Thr	Leu	Gly 720	
	Asp	Glu	Leu	Tyr 725												
		(2) IV	FORM.	OITA	N FC	R SE	Q ID	NO:	72:						
	(i) S	EOUE	NCE	CHAF	ACTE	RIST	ics:								
	,	(A)	LEN	GTH:	275	1 ba	se p	airs								
					ucle DNES			.e								
		(D)	TOF	OLOG	SY: 1	inea	r									
	-			CULE TURE :	E TYI	E: c	DNA									
								Seque	ence							
					ION: INFO											
		(xi)	SEQ	JENCI	E DES	CRI	PTIO	N: SI	EQ II	ON C	:72:					
ΑT	G GC1	GAC	GT"	r ta	C CC	G GC	C AAC	C GAG	TCC	DAC	G GC	G TC	r ca	G GA	C GTG	4.8
Me 1	t Ala	a Asp	o Val	1 Ty: 5	r Pro	o Ala	a Asi	n Ası	o S∈: 10	r Thi	r Al	a Se:	r GI	n As; 15	p Val	
GC	C AAG	C CG(TTC	C GC	c cg	C AA	A GG	G GC	G CTY	G AG	G CA	G AA	G AA	C GT	G CAT	96
Al	a Asr	n Arg	g Ph 20	e Al	a Ar	g Ly:	s GI	25	a Lei	I AL	Ĉ GI	и гу	30	II va	l His	,
GA	G GTY	G AAJ	A GA	C CA	C AA	TT A	C AT	c GC	C CG	C TT	C TT	C AA	G CA	A CC	C ACC	144
Gl	u Vai	1 Ly: 35	s As	p Hi	s Ly	s Ph	e II 40		a Ar	g Pn	e Pn	е гу 45		n Pi	o Thi	-
TT	C TG	C AG	C CA	C TG	C AC	C GA	C TT	C AT	C TG	G GG	G TI	T GG	G A	A CA	A GGO	192
Ph	е Су 50		r Hi	s Cy	s Th	r As 55		e Il	e Tr	p Gl	y Ph 60		у Гу	's Gl	n Gly	<i>?</i>

					TGC Cys 70											240
					TGT Cys			_								288
					CAC His											336
					TGT Cys											384
					ACC Thr											432
					CTC Leu 150											480
					GCT Ala					_				_	_	528
					AAT Asn											576
					CTG Leu											624
					ACC Thr								_			672
				-	AAA Lys 230								_	_	CTG Leu 240	720
					GAC Asp											768
														Ala	AGT Ser	816
													Tyr		GTG Val	864
CCC	ATT	CCA	GAA	GGA	GAT	GAA	GAA	GGC	AAC	ATG	GAA	CTC	AGG	CAG	AAG	912

Pro	Ile 290	Pro	Glu	Gly	Asp	Glu (295	Glu	Gly	Asn	Met	Glu 300	Leu	Arg	Gln	Lys	
					CTA Leu 310											960
TCA Ser	GAA Glu	GAC Asp	AGA Arg	AAG Lys 325	CAA Gln	CCA Pro	TCC Ser	AAC Asn	AAC Asn 330	CTG Leu	GAC Asp	AGA Arg	GTG Val	AAA Lys 335	CTC Leu	1008
ACA Thr	GAC Asp	TTC Phe	AAC Asn 340	TTC Phe	CTC Leu	ATG Met	GTG Val	CTG Leu 345	GGG Gly	AAG Lys	GGG Gly	AGT Ser	TTT Phe 350	GGG Gly	AAG Lys	1056
GTG Val	ATG Met	CTT Leu 355	GCT Ala	GAC Asp	AGG Arg	AAG Lys	GGA Gly 360	ACG Thr	GAG Glu	GAA Glu	CTG Leu	TAC Tyr 365	GCC Ala	ATC Ile	AAG Lys	1104
ATC Ile	CTG Leu 370	AAG Lys	AAG Lys	GAC Asp	GTG Val	GTG Val 375	ATC Ile	CAG Gln	GAC Asp	GAC Asp	GAC Asp 380	GTG Val	GAG Glu	TGC Cys	ACC Thr	1152
ATG Met 385	Val	GAG Glu	AAG Lys	CGC	GTG Val 390	CTG Leu	GCC Ala	CTG Leu	CTG Leu	GAC Asp 395	Lys	CCG Pro	CCA Pro	TTT Phe	CTG Leu 400	1200
ACA Thr	CAG Gln	CTG Leu	CAC His	TCC Ser 405	Cys	TTC Phe	CAG Gln	ACA Thr	GTG Val 410	Asp	CGG Arg	CTG Leu	TAC Tyr	TTC Phe 415		1248
				Asn					Met					Glr	GTC Val	1296
Gly	AAA Lys	TTT Phe 435	Lys	GAC Glu	CCA Pro	CAA Gln	GCA Ala 440	val	TTC Phe	TAC Tyr	C GCA	A GCC A Ala 445	Glu	ATC	TCC Ser	1344
ATC Ile	GGA Gly 450	/ Lei	TTC 1 Phe	TTC Phe	CTT Leu	CAT His	Lys	A AGA S Arg	Gly GGC	AT(e Ile 46	e Tyr	AGC Arg	G GA'	r CTG Leu	1392
AA0 Lys 465	. Lev	AAG ASI	C AAT 1 Asr	GTY Val	ATG Met 470	Leu	AAC Asr	TCA Ser	GAZ Glu	47	y Hi	C ATO	C AAA e Lys	A ATO	GCC e Ala 480	1440
GA(As _l	TTX Phe	G GGG G Gly	G ATK y Met	TG(Cy: 48!	s Lys	GAA Glu	A CAG	C ATO	3 ATC : Met 490	t As	T GG p G1	A GT y Va	C ACC	G AC r Th 49	C AGG r Arg 5	1498
AC Th	C TTY r Phe	C TG	C GGZ s Gly 50	y Th	r CCC r Pro	G GAC	TAC Ty:	C AT r Ile 509	e Ala	C CC a Pr	A GA o Gl	G AT u Il	A AT e Il 51	e Al	T TAC a Tyr	1536
CA Gl	G CCC	G TA O Ty 51	r Gl	G AA y Ly	G TC' s Sei	r GTA	A GA l As 52	p Tŋ	G TG	G GC p Al	G TA a Ty	C GG T G1 52	y Va	G CI 1 Le	G CTG u Leu	1584

											GGT Gly 540					1632
											TCC Ser					1680
											CTT Leu				_	1728
											GAG Glu			_		1776
											AAA Lys					1824
											GGC Gly 620					1872
											GTC Val				CCA Pro 640	1920
											GAT Asp					1968
											TTG Leu			Ala	GTA Val	2016
															CCA Pro	2064
		Val					Asp					Lys			GTT Val	2112
											Lys				AAA Lys 720	2160
										Pro					GTC Val	2208
									Phe					Asp	CAT His	2256
ATG	AAA	CAG	CAT	GAC	TTT	TTC	AAG	AGT	GCC	OTA :	CCC	GAA	GGI	rat :	GTA	2304

Met	Lys	Gln 755	His	Asp	Phe	Phe	Lys 760	Ser	Ala	Met	Pro	Glu 765	Gly	Tyr	Val	
CAG Gln	GAA Glu 770	AGA Arg	ACT Thr	ATA Ile	TTT Phe	TAC Tyr 775	AAA Lys	GAT Asp	GAC Asp	GGG Gly	AAC Asn 780	TAC Tyr	AAG Lys	ACA Thr	CGT Arg	2352
GCT Ala 785	GAA Glu	GTC Val	AAG Lys	TTT Phe	GAA Glu 790	GGT Gly	GAT Asp	ACC Thr	CTT Leu	GTT Val 795	AAT Asn	AGA Arg	ATC Ile	GAG Glu	TTA Leu 800	2400
AAA Lys	GGT Gly	ATT	GAT Asp	TTT Phe 805	AAA Lys	GAA Glu	GAT Asp	GGA Gly	AAC Asn 810	ATT Ile	CTT Leu	GGA Gly	CAC His	AAA Lys 815	ATG Met	2448
GAA Glu	TAC Tyr	AAT Asn	ТАТ Туг 820	AAC Asn	TCA Ser	CAT His	AAT Asn	GTA Val 825	TAC Tyr	ATC Ile	ATG Met	GCA Ala	GAC Asp 830	AAA Lys	CCA Pro	2496
AAG Lys	AAT Asn	GGC Gly 835	ATC Ile	AAA Lys	GTT Val	AAC Asn	TTC Phe 840	AAA Lys	ATT	AGA Arg	CAC His	AAC Asn 845	ATT	AAA Lys	GAT Asp	2544
GGA Gly	AGC Ser 850	Val	CAA Gln	TTA Leu	GCA Ala	GAC Asp 855	His	TAT Tyr	CAA Gln	CAA Gln	AAT Asn 860	Thr	CCA Pro	ATT Ile	GGC Gly	2592
GAT Asp 865	Gly	CCT Pro	GTC Val	CTT Leu	TTA Leu 870	Pro	GAC Asp	AAC Asr	CAT His	TAC Tyr 875	Leu	TCC Ser	ACG Thr	CA#	TCT Ser 880	2640
GCC	CTI Lev	TCC Ser	AAA Lys	GAT Asp 885	Pro	AAC Asr	GAA Glu	AAC Lys	AGA Arg 890	Asp	CAC His	ATC Met	ATC	CT: Lev 895	CTT Leu	2688
GAC Glu	TTI Phe	r GTA e Val	ACA Thr	Ala	GCT Ala	Gl ⁷	ATT	ACA Thi	His	GI)	C ATY Met	G GAT	GAA Glu 910	ı Le	A TAC 1 Tyr	2736
		CAC Glr 915	ı Glu		Α.											2751

- (2) INFORMATION FOR SEQ ID NO:73:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 916 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:73:

Met Ala Asp Val Tyr Pro Ala Asn Asp Ser Thr Ala Ser Gln Asp Val

1				5					10					15	
Ala			20	Ala	Arg			25					30		
Glu		Lys 35	Asp	His	Lys	Phe	Ile 40	Ala	Arg	Phe	Phe	Lys 45	Gln	Pro	Thr
Phe	Cys 50	Ser	His	Cys	Thr	Asp 55	Phe	Ile	Trp	Gly	Phe 60	Gly	Lys	Gln	Gly
Phe 65	Gln	Cys	Gln	Val	Cys 70	Cys	Phe	Val	Val	His 75	Lys	Arg	Суѕ	His	Glu 80
Phe				85	Cys				90					95	
			100		His			105					110		
		115			Cys		120					125			
	130				Thr	135					140				
145					Leu 150					155					160
				165	Ala				170					175	
			180		Asn			185					190		
		195	,		Leu		200					205			
	210				Thr	215					220				
225					230					235	·				Leu 240
				245	Asp				250)				255	1
			260)				265	,				270)	Ser
		275	5				280)				285	•		val
	290)				295	5				300)			Lys
305	;				310)				315	5				20 320
Ser	Glu			325	5				330)				333	
			34	0				345	5				35	0	y Lys
		35	5				360)				36	5		e Lys
	370	0				37	5				38	0			s Thr
Met 385		l Gl	u Ly	s Ar	g Va. 390		u Ale	a Le	u Le	u As 39	р гу 5	S PI	O PI	O FII	e Leu 400
Th	r Glı			40	r Cy: 5	s Ph			41	0				41	
			42	1 As	n Gl			42	5				4.5	U	n Val
		43	ie Ly	s Gl			44	0				44	.5		e Ser
	4.5	0				45	5				46	0			p Leu
Ly	s Le	u As	sn As	n Va	l Me	t Le	u As	n Se	r Gl	u Gl	у Ні	s Il	e L	's Il	e Ala

465					470					475					480
Asp				485					490					495	
			Gly 500					505					510		
		515	Gly				520					525			
	530		Leu			535					540				
545			Gln		550					555					560
			Glu	565					570					575	
			Arg 580					585					590		
		595	Phe				600					605			
	610		Pro			615					620				
625			Lys		630					635					640
			Val	645					650					655	
			Asn 660					665					670		
		675	Met Glu				680					685			
	690		Gly			695					700				
705					710					715					720
			Thr	725					730					735	
			740					745					750		His
		755					760					765			Val
	770					775					780				Arg Leu
785					790					795					800
				805					810)				815	
			820					825					830)	Pro
		835	,				840)				845			Asp
_	850					855	5				860)			Gly
Asp 865	Gly	Pro	Val	rea	870) ASE) AST	i His	875		ı ser	1111	. G11	880
				885	,				890)				895	
Glu	Ph∈	· Val	1 Thr 900		Ala	Gly	/ Ile	905		Gly	/ Met	. Asp	910		ı Tyr
Lys	Pro	915	n Glu	1											

(2) INFORMATION FOR SEQ ID NO:74:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2157 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...2154
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:74:

	()	.1) =	EQUE	71/012	DLSC	1/11/1	1014.	224			•					
ATG Met 1	TCG Ser	TCC Ser	ATC Ile	TTG Leu 5	CCA Pro	TTC Phe	ACG Thr	CCG Pro	CCA Pro 10	GTT Val	GTG Val	AAG Lys	AGA Arg	CTG Leu 15	CTG Leu	48
GGA Gly	TGG Trp	AAG Lys	AAG Lys 20	TCA Ser	GCT Ala	GGT Gly	GGG Gly	TCT Ser 25	GGA Gly	GGA Gly	GCA Ala	GGC Gly	GGA Gly 30	GGA Gly	GAG Glu	96
														AGT Ser		144
GTG Val	AAG Lys 50	AAG Lys	CTA Leu	AAG Lys	AAA Lys	ACA Thr 55	GGA Gly	CGA Arg	TTA Leu	GAT Asp	GAG Glu 60	CTT Leu	GAG Glu	AAA Lys	GCC Ala	192
ATC Ile 65	ACC Thr	ACT Thr	CAA Gln	AAC Asn	TGT Cys 70	AAT Asn	ACT Thr	AAA Lys	TGT Cys	GTT Val 75	ACC Thr	ATA Ile	CCA Pro	AGC Ser	ACT Thr 80	240
TGC Cys	TCT Ser	GAA Glu	ATT Ile	TGG Trp 85	GGA Gly	CTG Leu	AGT Ser	ACA Thr	CCA Pro 90	AAT Asn	ACG Thr	ATA Ile	GAT Asp	CAG Gln 95	TGG Trp	288
GAT Asp	ACA Thr	ACA Thr	GGC Gly 100	CTT Leu	TAC Tyr	AGC Ser	TTC Phe	TCT Ser 105	GAA Glu	CAA Gln	ACC Thr	AGG Arg	TCT Ser 110	CTT Leu	GAT Asp	336
GGT Gly	CGT Arg	CTC Leu 115	Gln	GTA Val	TCC Ser	CAT His	CGA Arg 120	AAA Lys	GGA Gly	TTG	CCA Pro	CAT His 125	Val	`ATA	TAT Tyr	384
TGC Cys	CGA Arg 130	Leu	TGG Trp	CGC Arg	TGG Trp	CCT Pro 135	Asp	CTT Leu	CAC	AGT Ser	CAT His	His	GAA Glu	CTC	AAG Lys	432
GCA Ala 145	Ile	GAA Glu	AAC Asn	TGC Cys	GAA Glu 150	Tyr	GCT Ala	TTI Phe	AAT Asn	CTI Leu 155	ı Lys	A AAC S Lys	GAT Asp	GAA Glu	GTA Val 160	480
TGT	GTA	. AAC	CCT	TAC	CAC	TAT	CAG	AGA	GTI	GAC	ACA	A CCA	A GT	r TTC	CCT	528

Cys	Val	Asn	Pro	Tyr 165	His	Tyr	Gln	Arg	Val 170	Glu	Thr	Pro	Val	Leu 175	Pro	
CCA Pro	GTA Val	TTA Leu	GTG Val 180	CCC Pro	CGA Arg	CAC His	ACC Thr	GAG Glu 185	ATC Ile	CTA Leu	ACA Thr	GAA Glu	CTT Leu 190	CCG Pro	CCT Pro	576
								CCA Pro								624
GGA Gly	ATT Ile 210	GAG Glu	CCA Pro	CAG Gln	AGT Ser	AAT Asn 215	тат Туг	ATT Ile	CCA Pro	GAA Glu	ACG Thr 220	CCA Pro	CCT Pro	CCT Pro	GGA Gly	672
TAT Tyr 225	ATC Ile	AGT Ser	GAA Glu	GAT Asp	GGA Gly 230	GAA Glu	ACA Thr	AGT Ser	GAC Asp	CAA Gln 235	CAG Gln	TTG Leu	AAT Asn	CAA Gln	AGT Ser 240	720
ATG Met	GAC Asp	ACA Thr	GGC Gly	TCT Ser 245	CCA Pro	GCA Ala	GAA Glu	CTA Leu	TCT Ser 250	CCT Pro	ACT Thr	ACT Thr	CTT Leu	TCC Ser 255	CCT Pro	768
GTT Val	AAT Asn	CAT	AGC Ser 260	TTG Leu	GAT Asp	TTA Leu	CAG Gln	CCA Pro 265	GTT Val	ACT Thr	TAC Tyr	TCA Ser	GAA Glu 270	Pro	GCA Ala	816
TTI Phe	TGG Trp	TGT Cys 275	Ser	ATA Ile	GCA Ala	тат Туг	ТАТ Туг 280	Glu	TTA Leu	TAA naA	CAG Gln	AGG Arg 285	Val	GGA Gly	GAA Glu	864
ACC Thr	TTC Phe 290	His	GCA Ala	TCA Ser	CAG Gln	CCC Pro 295	Ser	CTC Leu	ACT Thr	GTA Val	GAT Asp 300	Gly	TTI Phe	ACA Thr	GAC Asp	912
CCF Pro 305	Ser	AAT Asr	TCA Ser	GAG Glu	AGG Arg 310	Phe	TGC Cys	TTA Leu	GGT Gly	TTA Leu 315	Leu	TCC Ser	AA1 Asr	r GTT n Val	AAC Asn 320	960
CG/ Arg	A AAT J Asr	GCC Ala	ACC Thr	GTA Val	Glu	ATG Met	ACA Thi	A AGA c Arg	AGG Arg 330	His	T ATA	GGA Gly	A AGA	335 335	A GTG / Val	1008
CGG	TTA	A ТАС 1 ТУ1	TAC Ty:	: Ile	GGT Gly	GGC Gly	GAZ Glv	A GTT u Val 345	Phe	GCT Ala	r GAC a Glu	TG(CTI Let 35	ı Se:	r GAT r Asp	1056
			e Phe					o Asr					д Ту		TGG Y Trp	1104
CA Hi	c cc s Pro 370	o Ala	A AC	A GTO	G TGT L Cys	r AA 5 Lys 379	s Il	T CC? e Pro	A CC	A GG(o Gl:	TG' Y Cy: 38	s As:	r CT n Le	G AA u Ly	G ATC s Ile	1152
TT Ph 38	∈ As	C AA n As:	C CA	G GAI n Gli	A TT 1 Pho 39	e Ala	r GC a Al	T CT a Lei	r CTY	G GC u Al 39	a Gl	G TC n Se	T GT r Va	T AA 1 As	T CAG n Gln 400	1200

											TGC Cys					1248
											AGG Arg					1296
											GGA Gly					1344
											TCA Ser 460					1392
AGC Ser 465	ATG Met	TCA Ser	TGG Trp	GTA Val	CCG Pro 470	CGG Arg	GCC Ala	CGG Arg	GAT Asp	CCA Pro 475	CCG Pro	GTC Val	GCC Ala	ACC Thr	ATG Met 480	1440
											GTG Val					1488
GAG Glu	CTG Leu	GAC Asp	GGC Gly 500	Asp	GTA Val	AAC Asn	GGC Gly	CAC His 505	AAG Lys	TTC Phe	AGC Ser	GTG Val	TCC Ser 510	Gly	GAG Glu	1536
GGC Gly	GAG Glu	GGC Gly 515	Asp	GCC Ala	ACC Thr	TAC Tyr	GGC Gly 520	Lys	CTG Leu	ACC Thr	CTG Leu	AAG Lys 525	TTC	ATC Ile	TGC Cys	1584
		Gly					Pro				CTC Leu 540	Val			CTG Leu	1632
ACC Thr 545	Tyr	GGC	GTG Val	CAG Gln	TGC Cys 550	Phe	AGC Ser	CGC Arg	TAC Tyr	CCC Pro 555	Asp	CAC His	ATC Met	AAG Lys	Gln 560	1680
CAC His	GAC Asp	TTC Phe	TTC Phe	Lys	TCC	Ala	: ATC	CCC Pro	GAA Glu 570	Gly	TAC	GTC Val	Glr	GAC 1 Glu 575	Arg	1728
ACC Thr	: ATC	TTC Phe	TTC Phe 580	Lys	GAC Asp	GAC Asp	Gly	AAC Asr 585	туг	AAC Lys	G ACC	CGC Arg	GC0 Ala 590	a Glu	GTG Val	1776
AAG Lys	TTC Phe	GAC Glu 599	ı Gl	GAC Asp	ACC Thr	CTC Lev	GT(1 Val 600	l Asr	CGC Arg	TATO	C GAC e Glu	CTC Lev 605	Ly:	G GG(S Gl)	ATC / Ile	1824
GAC Asr	TTC Phe 610	≥ Lys	G GAC	GAC 1 Asp	GG(AAC Asr 615	ı Ile	CTC e Lev	GGC Gly	G CAG	C AAC s Ly: 620	s Leu	G GAG	G TAC	AAC Asn	1872
TAC	AA C	AG(CAC	DAA C	GTO	TA?	TA T	OTA C	G GCC	C GA	C AAG	G CAC	AA E	G AA	C GGC	1920

Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly ATC AAG GTG AAC TTC AAG ATC CGC CAC AAC ATC GAG GAC GGC AGC GTG Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val CAG CTC GCC GAC CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC CCC 2016 Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro 660 GTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG AGC 2064 Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC GTG 2112 Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val 690 ACC GCC GCC GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TAA 2157 Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 710

(2) INFORMATION FOR SEQ ID NO:75:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 718 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:75:

Met Ser Ser Ile Leu Pro Phe Thr Pro Pro Val Val Lys Arg Leu Leu 10 1 Gly Trp Lys Lys Ser Ala Gly Gly Ser Gly Gly Ala Gly Gly Glu 25 Gln Asn Gly Gln Glu Glu Lys Trp Cys Glu Lys Ala Val Lys Ser Leu 40 Val Lys Lys Leu Lys Lys Thr Gly Arg Leu Asp Glu Leu Glu Lys Ala 55 Ile Thr Thr Gln Asn Cys Asn Thr Lys Cys Val Thr Ile Pro Ser Thr 75 70 Cys Ser Glu Ile Trp Gly Leu Ser Thr Pro Asn Thr Ile Asp Gln Trp 90 85 Asp Thr Thr Gly Leu Tyr Ser Phe Ser Glu Gln Thr Arg Ser Leu Asp 100 105 110 Gly Arg Leu Gln Val Ser His Arg Lys Gly Leu Pro His Val Ile Tyr 115 120 125 Cys Arg Leu Trp Arg Trp Pro Asp Leu His Ser His His Glu Leu Lys 135 140 Ala Ile Glu Asn Cys Glu Tyr Ala Phe Asn Leu Lys Lys Asp Glu Val 155 150 Cys Val Asn Pro Tyr His Tyr Gln Arg Val Glu Thr Pro Val Leu Pro

				165					170					175	
Pro	Val	Leu	Val 180	Pro	Arg	His	Thr	Glu 185	Ile	Leu	Thr	Glu	Leu 190	Pro	Pro
Leu	Asp	Asp 195	Tyr	Thr	His	Ser	Ile 200	Pro	Glu	Asn	Thr	Asn 205	Phe	Pro	Ala
Gly	11e 210	Glu	Pro	Gln	Ser	Asn 215	Tyr	Ile	Pro	Glu	Thr 220	Pro	Pro	Pro	Gly
225				-	230		Thr			235					240
				245			Glu		250					255	
			260				Gln	265					270		
		275					Tyr 280					285			
	290					295	Ser				300				
305					310		Cys			315					320
				325			Thr Glu		330					335	
			340				Pro	345					350		
		355					360 Ile		_			365			
	370					375	Ala				380				
385					390					395					400
				405			Leu		410					415	
			420	_	_		Ala	425					430		
		435					Leu 440 Met					445			
	450	_				455	Ala				460				
465					470					475					480
				485			Phe		490					495	
			500					505					510		Glu
		515					520					525			Cys
	530					535					540				Leu
7nr 545		GIY	vai	GIN	550		Ser	Arg	ıyı	555	Asp	nis	rie c	Lys	Gln 560
		Phe	Phe	Lys 565			Met	Pro	Glu 570		Tyr	Val	Gln	Glu 575	Arg
			580					585					590		Val
		595					600					605			Ile
•	610	_		_	_	615					620)			Asn
Tyr	Asn	Ser	His	Asn	Val	Tyr	Ile	Met	Ala	Asp	Lys	Gln	Lys	Asn	Gly

625 630 635 640	
Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val 645 650 655	
Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro	
Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser	
675 680 685 Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val	
690 695 700	
Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 705 710 715	
(2) INFORMATION FOR SEQ ID NO:76:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 2397 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single (D) TOPOLOGY: linear	
A CONTROL OF THE PROPERTY OF T	
(ii) MOLECULE TYPE: cDNA (ix) FEATURE:	
(A) NAME/KEY: Coding Sequence	
(B) LOCATION: 12394	
(D) OTHER INFORMATION:	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:76:	
ATG GAC AAT ATG TOT ATT ACG AAT ACA COA ACA AGT AAT GAT GCC TGT	48
Met Asp Asn Met Ser Ile Thr Asn Thr Pro Thr Ser Asn Asp Ala Cys 1 5 10 15	
CTG AGC ATT GTG CAT AGT TTG ATG TGC CAT AGA CAA GGT GGA GAG AGT Leu Ser Ile Val His Ser Leu Met Cys His Arg Gln Gly Gly Glu Ser	96
20 25 30	
GAA ACA TTT GCA AAA AGA GCA ATT GAA AGT TTG GTA AAG AAG CTG AAG	144
GAA ACA TIT GCA AAA AGA GCA AIT GAA AGI IIG GIA AAG AAG CIG AAG Glu Thr Phe Ala Lys Arg Ala Ile Glu Ser Leu Val Lys Lys Leu Lys	
35 40 45	
GAG AAA AAA GAT GAA TTG GAT TCT TTA ATA ACA GCT ATA ACT ACA AAT	192
Glu Lys Lys Asp Glu Leu Asp Ser Leu Ile Thr Ala Ile Thr Thr Asn	
50 55 60	
GGA GCT CAT CCT AGT AAA TGT GTT ACC ATA CAG AGA ACA TTG GAT GGG	240
Gly Ala His Pro Ser Lys Cys Val Thr Ile Gln Arg Thr Leu Asp Gly 65 70 75 80	
65	
AGG CTT CAG GTG GCT GGT CGG AAA GGA TTT CCT CAT GTG ATC TAT GCC	288
Arg Leu Gln Val Ala Gly Arg Lys Gly Phe Pro His Val Ile Tyr Ala 85 90 95	
	226
CGT CTC TGG AGG TGG CCT GAT CTT CAC AAA AAT GAA CTA AAA CAT GTT Arg Leu Trp Arg Trp Pro Asp Leu His Lys Asn Glu Leu Lys His Val	336
arg Leu Trp Arg Trp Pro Asp Leu Ars Lys Ash Gra Leu Lys Ars 110	
AAA TAT TGT CAG TAT GCG TTT GAC TTA AAA TGT GAT AGT GTC TGT GTG	384

Lys	Tyr	Cys 115	Gln	Tyr	Ala		Asp 120	Leu	Lys	Cys	Asp	Ser 125	Val	Суѕ	Val	
AAT Asn	CCA Pro 130	TAT Tyr	CAC His	TAC Tyr	GAA Glu	CGA Arg 135	GTT Val	GTA Val	TCA Ser	CCT Pro	GGA Gly 140	ATT Ile	GAT Asp	CTC Leu	TCA Ser	432
GGA Gly 145	TTA Leu	ACA Thr	CTG Leu	CAG Gln	AGT Ser 150	AAT Asn	GCT Ala	CCA Pro	TCA Ser	AGT Ser 155	ATG Met	ATG Met	GTG Val	AAG Lys	GAT Asp 160	480
GAA Glu	ТАТ Туг	GTG Val	CAT His	GAC Asp 165	TTT Phe	GAG Glu	GGA Gly	CAG Gln	CCA Pro 170	TCG Ser	TTG Leu	TCC Ser	ACT Thr	GAA Glu 175	GGA Gly	528
CAT His	TCA Ser	ATT Ile	CAA Gln 180	ACC Thr	ATC Ile	CAG Gln	CAT His	CCA Pro 185	CCA Pro	AGT Ser	AAT Asn	CGT Arg	GCA Ala 190	Ser	ACA Thr	576
GAG Glu	ACA Thr	TAC Tyr 195	Ser	ACC Thr	CCA Pro	GCT Ala	CTG Leu 200	Leu	GCC Ala	CC# Pro	TCI Ser	GAG Glu 205	Ser	AAT Asn	GCT Ala	624
ACC Thr	AGC Ser 210	Thr	GCC Ala	AAC Asn	TTT Phe	CCC Pro 215	AAC Asn	ATT	CCT Pro	GT(G GCT L Ala 220	a Ser	ACA Thr	A AGT	CAG Gln	672
CCT Pro	Ala	: AGT Ser	TATA	CTG Leu	GGG Gly 230	Gly	AGC Ser	CAT His	AGT Ser	GAJ Glu 23!	ı Gly	A CTO	TTC	G CAC	ATA lle 240	720
GC? Ala	A TCA	. G17	G CCT	CAC Glr 245	n Pro	. GGA . Gly	CAC Glr	G CAG	G CAC 1 Glr 250	n As	T GG n Gl	A TT' y Phe	r AC' e Th:	r GG' r Gly 25!	r CAG y Gln 5	76 3
CC/ Pro	A GC' o Ala	r ACT a Thi	г тас тут 260	His	r CAT s His	`AAC Asn	: AG(: Se:	C ACT c Thr 265	Thi	C AC	C TG r Tr	G AC p Th	T GG. r Gl: 27	y Se	r AGG r Arg	816
AC'	T GC.	A CCA a Pro 27!	э Туг	ACA Thi	A CCI	TAA T	TTV Lev 28	u Pro	r CAG	C CA s Hi	C CA s Gl	A AA n As 28	n Gl	С СА У Ні	T CTT s Leu	864
CA Gl	G CA n Hi 29	s Hi	C CCC	G CC'	T ATO	G CCC E Pro 295	o Pr	C CA'	r CC s Pr	c GG o Gl	SA CA y Hi 30	s Ty	C TG T Tr	G CC P Pr	T GTT o Val	912
СА Ні 30	s As	T GA n Gl	G CT u Le	r GC. u Al	A TTO a Pho 31	e Gl	G CC n Pr	T CC	C AT	T TO e Se 31	er As	AT CA	T CC	T GC	CT CCT a Pro 320	960
GA Gl	G TA .u Ty	T TG T Tr	G TG p Cy	T TC s Se 32	r Il	T GC' e Al	т та а ту	C TT T Ph	T GA e G1 33	u Me	rg GÆ et As	AT GT sp Va	TT CA	AG GT In Va 33	TA GGA al Gly 85	1008
G)	AG AC Lu Th	CA TI	T AA e Ly 34	s Va	T CC	T TC o Se	A AC	GC TG er Cy 34	's Pr	T A'	TT G' le V	TT AG	ar V	MT GA al As 50	AT GGA sp Gly	1056

TAC Tyr	GTG Val	GAC Asp 355	CCT Pro	TCT Ser	GGA (Gly	GAT Asp 360	CGC Arg	TTT Phe	TGT Cys	TTG Leu	GGT Gly 365	CAA Gln	CTC Leu	TCC Ser	1104
AAT Asn	GTC Val 370	CAC His	AGG Arg	ACA Thr	GAA Glu	GCC Ala 375	ATT Ile	GAG Glu	AGA Arg	GCA Ala	AGG Arg 380	TTG Leu	CAC His	ATA Ile	GGC Gly	1152
AAA Lys 385	GGT Gly	GTG Val	CAG Gln	TTG Leu	GAA Glu 390	TGT Cys	AAA Lys	GGT Gly	GAA Glu	GGT Gly 395	GAT Asp	GTT Val	TGG Trp	GTC Val	AGG Arg 400	1200
TGC Cys	CTT Leu	AGT Ser	GAC Asp	CAC His 405	GCG Ala	GTC Val	TTT Phe	GTA Val	CAG Gln 410	AGT Ser	TAC Tyr	TAC Tyr	TTA Leu	GAC Asp 415	AGA Arg	1248
GAA Glu	GCT Ala	GGG Gly	CGT Arg 420	GCA Ala	CCT Pro	GGA Gly	GAT Asp	GCT Ala 425	GTT Val	CAT His	AAG Lys	ATC Ile	TAC Tyr 430	CCA Pro	AGT Ser	1296
GCA Ala	ТАТ Туг	ATA Ile 435	Lys	GTC Val	TTT Phe	GAT Asp	TTG Leu 440	CGT Arg	CAG Gln	TGT Cys	CAT	CGA Arg 445	CAG Gln	ATG Met	CAG Gln	1344
CAG Gln	CAG Gln 450	Ala	GCT Ala	ACT Thr	GCA Ala	CAA Gln 455	Ala	GCA	GCA Ala	GCT Ala	GCC Ala 460	Gln	GCA Ala	GCA Ala	GCC Ala	1392
GTG Val 465	. Ala	GGA Gly	AAC Asn	ATC	CCT Pro 470	Gly	CCA Pro	GGA Gly	TCA Ser	GTA Val 475	. G13	r GGA / Gly	ATA / Ile	GCT Ala	CCA Pro 480	1440
GCT Ala	ATC	AG1 Sei	CTC	TCA Ser 485	Ala	GCT Ala	GCT Ala	r GGA a Gly	ATT 116 490	e Gl	r GT / Val	r GAT l Asp	r GAC o Asp	CTT Lev 495	CGT Arg	1488
CG(TTA J Lev	TG(E ATA	e Leu	AGG Arg	ATC Met	AG: Sei	r TTT r Phe 505	e Val	3 AAi l Lys	A GGG	C TG(y Tr _I	G GG7 D Gly 510	/ Pro	G GAT	1536
TAC Tyr	C CCF	A AG2 5 Arg 51	g Glr	3 AGC n Sei	C ATC	AA. Lys	52	u Th	A CCI	T TGG	C TG s Tr	G AT' p Ile 52!	e Gl	A AT'	r CAC e His	1584
TT. Le	A CAG u Hi: 53	s Ar	g GC0 g Ala	C CTO	C CAC	G CTY n Lev 53!	ı Le	A GA(u As)	C GAN p Gl	A GT. u Va	A CT l Le 54	u Hi	T AC	C AT r Me	G CCG t Pro	1632
AT 11 54	e Al	A GA a As	C CC. p Pr	A CAI	A CC' n Pro 550	o Lei	A GA u As	C TG p Tr	G GA' p As	T CC p Pr 55	o Pr	G GT O Va	C GC 1 Al	C AC a Th	C ATG r Met 560	1680
GT Va	G AG 1 Se	C AA r Ly	G GG s Gl	C GA y Gl 56	u Gl	G CT	G TT u Ph	C AC le Th	c GG r Gl 57	y Vā	G GI 1 Va	G CC	C AT	C CT e Le 57	G GTC Eu Val	1728
GA	G CT	G GA	.C GG	C GA	C GT	AA A	c GG	C CA	CAA	G TI	C AC	GC GI	TC TC	C GG	C GAG	1776

Glu	Leu	Asp	Gly 580	Asp	Val	Asn	Gly	His 585	Lys	Phe	Ser	Val	Ser 590	Gly	Glu	
							_		CTG Leu				_	_		1824
									CCC Pro							1872
									TAC Tyr							1920
									GAA Glu 650							1968
									TAC Tyr							2016
									CGC Arg							2064
									GGG Gly							2112
									GCC Ala						_	2160
									AAC Asn 730							2208
									ACC Thr							2256
									AGC Ser				Ala		AGC Ser	2304
												Leu			GTG Val	2352
									GAC Asp		Leu					2397

(2) INFORMATION FOR SEQ ID NO:77:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 798 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGRENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:77:

Met Asp Asn Met Ser Ile Thr Asn Thr Pro Thr Ser Asn Asp Ala Cys 5 10 Leu Ser Ile Val His Ser Leu Met Cys His Arg Gln Gly Glu Ser 25 20 Glu Thr Phe Ala Lys Arg Ala Ile Glu Ser Leu Val Lys Lys Leu Lys 40 Glu Lys Lys Asp Glu Leu Asp Ser Leu Ile Thr Ala Ile Thr Thr Asn 55 Gly Ala His Pro Ser Lys Cys Val Thr Ile Gln Arg Thr Leu Asp Gly 75 70 Arg Leu Gln Val Ala Gly Arg Lys Gly Phe Pro His Val Ile Tyr Ala 90 85 Arg Leu Trp Arg Trp Pro Asp Leu His Lys Asn Glu Leu Lys His Val 105 110 100 Lys Tyr Cys Gln Tyr Ala Phe Asp Leu Lys Cys Asp Ser Val Cys Val 120 125 115 Asn Pro Tyr His Tyr Glu Arg Val Val Ser Pro Gly Ile Asp Leu Ser 140 135 Gly Leu Thr Leu Gln Ser Asn Ala Pro Ser Ser Met Met Val Lys Asp 155 150 Glu Tyr Val His Asp Phe Glu Gly Gln Pro Ser Leu Ser Thr Glu Gly 170 His Ser Ile Gln Thr Ile Gln His Pro Pro Ser Asn Arg Ala Ser Thr 185 Glu Thr Tyr Ser Thr Pro Ala Leu Leu Ala Pro Ser Glu Ser Asn Ala 200 205 Thr Ser Thr Ala Asn Phe Pro Asn Ile Pro Val Ala Ser Thr Ser Gln 215 220 Pro Ala Ser Ile Leu Gly Gly Ser His Ser Glu Gly Leu Leu Gln Ile 225 230 235 Ala Ser Gly Pro Gln Pro Gly Gln Gln Asn Gly Fhe Thr Gly Gln 250 245 Pro Ala Thr Tyr His His Asn Ser Thr Thr Thr Trp Thr Gly Ser Arg 265 260 Thr Ala Pro Tyr Thr Pro Asn Leu Pro His His Gln Asn Gly His Leu 280 Gln His His Pro Pro Met Pro Pro His Pro Gly His Tyr Trp Pro Val 300 295 His Asn Glu Leu Ala Phe Gln Pro Pro Ile Ser Asn His Pro Ala Pro 315 310 Glu Tyr Trp Cys Ser Ile Ala Tyr Phe Glu Met Asp Val Gln Val Gly 330 325 Glu Thr Phe Lys Val Pro Ser Ser Cys Pro Ile Val Thr Val Asp Gly 345 340 Tyr Val Asp Pro Ser Gly Gly Asp Arg Phe Cys Leu Gly Gln Leu Ser 360 365

Asn Val His Arg Thr Glu Ala Ile Glu Arg Ala Arg Leu His Ile Gly

375 Lys Gly Val Gln Leu Glu Cys Lys Gly Glu Gly Asp Val Trp Val Arg 390 395 Cys Leu Ser Asp His Ala Val Phe Val Gln Ser Tyr Tyr Leu Asp Arg 405 410 Glu Ala Gly Arg Ala Pro Gly Asp Ala Val His Lys Ile Tyr Pro Ser 425 420 Ala Tyr Ile Lys Val Phe Asp Leu Arg Gln Cys His Arg Gln Met Gln 440 435 Gln Gln Ala Ala Thr Ala Gln Ala Ala Ala Ala Gln Ala Ala Ala 455 460 Val Ala Gly Asn Ile Pro Gly Pro Gly Ser Val Gly Gly Ile Ala Pro 470 475 Ala Ile Ser Leu Ser Ala Ala Ala Gly Ile Gly Val Asp Asp Leu Arg 490 485 Arg Leu Cys Ile Leu Arg Met Ser Phe Val Lys Gly Trp Gly Pro Asp 505 500 Tyr Pro Arg Gln Ser Ile Lys Glu Thr Pro Cys Trp Ile Glu Ile His 520 515 Leu His Arg Ala Leu Gln Leu Leu Asp Glu Val Leu His Thr Met Pro 535 540 Ile Ala Asp Pro Gln Pro Leu Asp Trp Asp Pro Pro Val Ala Thr Met 550 555 Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val 565 570 575 Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu 580 585 590 Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys 595 600 605 Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu 620 615 Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln 630 635 His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg 645 650 Thr Ile Phe Fhe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val 660 665 Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile 680 685 Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn 700 695 Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly 705 710 715 Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val 730 725 Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Fro Ile Gly Asp Gly Pro 745 Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser 760 Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val 775 Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys

(2) INFORMATION FOR SEQ ID NO:78:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3138 base pairs

	((C) S	TRAN		ESS:	aci sin										
	(ii) MOLECULE TYPE: cDNA (ix) FEATURE:															
	(A) NAME/KEY: Coding Sequence(B) LOCATION: 13135(D) OTHER INFORMATION:															
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:78: ATG GCG GGC TGG ATC CAG GCC CAG CAG CTG CAG GGA GAC GCG CTG CGC																
ATG Met	GCG Ala	GGC Gly	TGG Trp	ATC Ile 5	CAG (GCC (Ala (CAG Gln	Gln	CTG Leu 10	CAG Gln	GGA (GAC Asp	GCG Ala	CTG Leu 15	CGC Arg	48
CAG Gln	ATG Met	CAG Gln	GTG Val 20	CTG Leu	TAC Tyr	GGC Gly	CAG Gln	CAC His 25	TTC Phe	CCC Pro	ATC Ile	GAG Glu	GTC Val 30	CGG Arg	CAC His	96
TAC Tyr	TTG Leu	GCC Ala 35	CAG Gln	TGG Trp	ATT Ile	GAG Glu	AGC Ser 40	CAG Gln	CCA Pro	TGG Trp	GAT Asp	GCC Ala 45	ATT Ile	GAC Asp	TTG Leu	144
GAC Asp	AAT Asn 50	CCC Pro	CAG Gln	GAC Asp	AGA Arg	GCC Ala 55	CAA Gln	GCC Ala	ACC Thr	CAG Gln	CTC Leu 60	CTG Leu	GAG Glu	GGC Gly	CTG Leu	192
GTG Val 65	CAG Gln	GAG Glu	CTG Leu	CAG Gln	AAG Lys 70	AAG Lys	GCG Ala	GAG Glu	CAC His	CAG Gln 75	GTG Val	GGG Gly	GAA Glu	GAT Asp	GGG Gly 80	240
TTT Pì:e	TTA Leu	CTG Leu	AAG Lys	ATC Ile 85	AAG Lys	CTG Leu	GGG Gly	CAC His	TAC Tyr 90	GCC Ala	ACG Thr	CAG Gln	CTC	CAG Gln 95	AAA Lys	288
ACA Thr	ТАТ Тух	GAC Asp	CGC Arg 100	Cys	CCC Pro	CTG Leu	GAG Glu	CTG Leu 105	Val	CGC Arg	TGC Cys	ATC Ile	CGC Arg	His	ATT Ile	336
CTG Leu	TAC Tyr	AAT Asn 115	Glu	. CAG . Gln	AGG Arg	CTG Leu	GTC Val 120	Arg	GAA Glu	A GCC	AAC Asn	AAT Asr 125) CA	C AGC S Ser	TCT Ser	384
CCG Pro	GCT Ala	Gly	ATC	: CTG	GTT Val	GAC Asp	Ala	ATG Met	TCC Ser	CA(AAG Lys 140	His	CT s Le	r CAC u Glr	G ATC	432
AAC	CAC	ACA	TTT	GAC	GAC	CTC	G CGF	A CTC	GTO	C AC	G CAC	GA(C AC.	A GAG	TAA E	480

Asn Gln Thr Phe Glu Glu Leu Arg Leu Val Thr Gln Asp Thr Glu Asn

GAG CTG AAG AAA CTG CAG CAG ACT CAG GAG TAC TTC ATC ATC CAG TAC

Glu Leu Lys Lys Leu Gln Gln Thr Gln Glu Tyr Phe Ile Ile Gln Tyr

CAG GAG AGC CTG AGG ATC CAA GCT CAG TTT GCC CAG CTG GCC CAG CTG

Gln Glu S	er Leu 180	Arg Ile	Gln	Ala	Gln 185	Phe	Ala	Gln	Leu	Ala 190	Gln	Leu	
AGC CCC C Ser Pro G													624
GTG TCT C Val Ser L 210													672
TAC CGC G Tyr Arg V 225													720
CGG AAG C Arg Lys G			_										768
CGG CGG C Arg Arg G													816
GAC GTG C Asp Val L													864
AAC CGG C Asn Arg G 290													912
ATC CCC G Ile Pro G 305													960
ACG GAC A Thr Asp I										_			1008
CAG CCT C													1056
CGC CTG C Arg Leu L 3													1104
GTG AAG G Val Lys A 370													1152
AAT GAG A Asn Glu A 385													1200
TGC GTG A													1248

AGG AAC AT Arg Asn Me	rG TCA et Ser 420	CTG AAG Leu Lys	AGG ATG	C AAG (e Lys 2 425	CGT G(Arg A)	CT GAC la Asp	CGG CG Arg Ar 43	g Gly	GCA Ala	1296
GAG TCC GT Glu Ser Va	rg ACA al Thr 35	GAG GAG Glu Glu	AAG TTO Lys Pho	e Thr	GTC C' Val L	TG TYT eu Phe	GAG TO Glu Se 445	CT CAG er Gln	TTC Phe	1344
AGT GTT G Ser Val G 450	GC AGC ly Ser	AAT GAG Asn Glu	CTT GT Leu Va 455	G TTC l Phe	CAG G Gln V	TG AAG al Lys 460	ACT C	rg TCC eu Ser	CTA Leu	1392
CCT GTG G Pro Val V 465	TT GTC	ATC GTC Ile Val 470	His Gl	C AGC y Ser	Gln A	AC CAC Sp His	AAT G Asn A	CC ACG la Thr	GCT Ala 480	1440
ACT GTG C	TG TGG eu Trp	GAC AAT Asp Asr 485	GCC TT Ala Ph	TT GCT ne Ala	GAG C Glu F 490	cc GGC Pro Gly	AGG G Arg V	TG CCA al Pro 495	Phe	1488
GCC GTG C Ala Val F	CT GAC Pro Asp 500	AAA GTO	CTG TO	G CCG OP Pro 505	CAG (CTG TGT Leu Cys	Glu A	CG CTC la Leu 10	AAC Asn	1536
ATG AAA 1 Met Lys I	MTC AAG Phe Lys 515	GCC GAA	ı Val G	AG AGC ln Ser 20	AAC (Asn)	CGG GG(Arg Gl)	CTG A Leu T 525	CC AAC	G GAG Glu	1584
AAC CTC (Asn Leu V	GTG TTC Val Phe	CTG GCC	G CAG A a Gln L 535	AA CTG ys Leu	TTC I	AAC AAG Asn Asi 54	n Ser S	AGC AGC Ser Sei	C CAC C His	1632
CTG GAG (Leu Glu) 545	GAC TAC Asp Tyr	AGT GG Ser Gl; 55	y Leu S	CC GTG er Val	Ser	TGG TC Trp Se 555	C CAG '	Phe Asi	n Arg 560	1680
GAG AAC Glu Asn	TTG CCC Leu Pro	GGC TG Gly Tr 565	G AAC T p Asn T	AC ACC	TTC Phe 570	TGG CA Trp Gl	G TGG ' n Trp	TTT GA Phe As 57	p Gly	1728
GTG ATG Val Met	GAG GTX Glu Va: 580	l Leu Ly	G AAG C s Lys H	AC CAC is His 585	Lys	CCC CA Pro Hi	C TGG s Trp	AAT GA Asn As 590	T GGG p Gly	1776
GCC ATC Ala Ile	CTA GG' Leu Gl _! 595	T TTT GI y Phe Va	l Asn I	AAG CAA Jys Gli 300	A CAG	GCC CA Ala Hi	C GAC s Asp 605	CTG CT Leu Le	C ATC	1824
AAC AAG Asn Lys 610	CCC GA	C GGG AC p Gly Th	CC TTC T nr Phe I 615	rTG TT Leu Le	G CGC u Arg	Phe Se	GT GAC er Asp 20	TCA GA	A ATC Lu Ile	1872
GGG GGC Gly Gly 625	ATC AC	r Ile A	CC TGG A La Trp 1 30	AAG TT Lys Ph	T GAC e Asp	TCC CC Ser P: 635	CG GAA ro Glu	CGC AM	AC CTG sn Leu 640	1920
TGG AAC	CTG AA	A CCA T	rc acc	ACG CG	g gat	TTC T	OTA OO	AGG TY	CC CTG	1968

Trp	Asn	Leu	Lys	Pro 645	Phe	Thr	Thr	Arg	Asp 650	Phe	Ser	Ile	Arg	Ser 655	Leu	
					GAC Asp											2016
					GTC Val											2064
					ТАТ Туг											2112
					TCT Ser 710											2160
					TCC Ser											2208
					CCT Pro											2256
					ATG Met						_					2304
					AGT Ser											2352
					AGA Arg 790						_					2400
			_	_	ACC Thr		_			_	_	_				2448
					CTG Leu											2496
					GGC Gly											2544
					ATC Ile											2592
					ACC Thr 870											2640

TAC Tyr	CCC Pro	GAC Asp	CAC His	ATG Met 885	AAG Lys	CAG Gln	CAC His	GAC Asp	TTC Phe 890	TTC Phe	AAG Lys	TCC Ser	GCC Ala	ATG Met 895	CCC Pro	2688
GAA Glu	GGC Gly	TAC Tyr	GTC Val 900	CAG Gln	GAG Glu	CGC Arg	ACC Thr	ATC Ile 905	TTC Phe	TTC Phe	AAG Lys	GAC Asp	GAC Asp 910	GGC Gly	AAC Asn	2736
TAC Tyr	AAG Lys	ACC Thr 915	CGC Arg	GCC Ala	GAG Glu	GTG Val	AAG Lys 920	TTC Phe	GAG Glu	GGC Gly	GAC Asp	ACC Thr 925	CTG Leu	GTG Val	AAC Asn	2784
CGC Arg	ATC Ile 930	GAG Glu	CTG Leu	AAG Lys	GGC Gly	ATC Ile 935	GAC Asp	TTC Phe	AAG Lys	GAG Glu	GAC Asp 940	GGC Gly	AAC Asn	ATC Ile	CTG Leu	2832
GGG Gly 945	CAC His	AAG Lys	CTG Leu	GAG Glu	TAC Tyr 950	AAC Asn	TAC Tyr	AAC Asn	AGC Ser	CAC His 955	AAC Asn	GTC Val	тат туг	ATC Ile	ATG Met 960	2880
GCC Ala	GAC Asp	AAG Lys	CAG Gln	AAG Lys 965	AAC Asn	GGC Gly	ATC Ile	AAG Lys	GTG Val 970	AAC Asn	TTC Phe	AAG Lys	ATC	CGC Arg 975	His	2928
AAC Asn	ATC Ile	GAG Glu	GAC Asp 980	Gly	AGC Ser	GTG Val	CAG Gln	CTC Leu 985	Ala	GAC Asp	CAC	TAC Tyr	CAC Glr 990	Gln	AAC Asn	2976
ACC Thr	CCC	ATC Ile 995	Gly	GAC Asp	GGC Gly	CCC Prc	GTG Val	Leu	CTG Leu	CCC Pro	GAC Asp	AAC Asn 1005	His	TAC TYT	CTG Leu	3024
AGC Ser	Thr	Gln	TCC Ser	GCC Ala	CTG Leu	AGC Ser 1015	Lys	GAC Asp	CCC Prc	AAC Asn	GAC Glu 1020	Lys	G CGC	GAT J Asp	CAC His	3072
ATG Met 1025	. Val	CTC Lev	CTG	GAG Glu	TTC Phe	· Val	ACC l Thr	GCC Ala	GCC Ala	GGG Gly 1035	, Ile	C ACT	r CTY	u Gly	ATG Met 1040	3120
			TAC Tyr			A.										3138

(2) INFORMATION FOR SEQ ID NO:79:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1045 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:79:

Met 1	Ala	Gly	Trp	Ile 5	Gln	Ala	Gln	Gln	Leu 10	Gln	Gly	Asp	Ala	Leu 15	Arg
Gln	Met	Gln	Val 20	Leu	Tyr	Gly	Gln	His 25	Phe	Pro	Ile	Glu	Val 30	Arg	His
Tyr	Leu	Ala 35	Gln	Trp	Ile	Glu	Ser 40	Gln	Pro	Trp	Asp	Ala 45	Ile	Asp	Leu
Asp	Asn 50	Pro	Gln	Asp	Arg	Ala 55	Gln	Ala	Thr	Gln	Leu 60	Leu	Glu	Gly	Leu
Val 65	Gln	Glu	Leu	Gln	Lys 70	Lys	Ala	Glu	His	Gln 75	Val	Gly	Glu	Asp	Gly 80
Phe	Leu	Leu	Lys	Ile 85	Lys	Leu	Gly	His	Tyr 90	Ala	Thr	Gln	Leu	Gln 95	Lys
Thr	Tyr	Asp	Arg 100	Cys	Pro	Leu	Glu	Leu 105	Val	Arg	Суѕ	Ile	Arg 110	His	Ile
	-	115			Arg		120					125			
	130				Val	135					140				
	Gln	Thr	Phe	Glu	Glu	Leu	Arg	Leu	Val		Gln	Asp	Thr	Glu	
145	T	7	1	T 011	150	Cln	mb ×	Cln	C1	155	Pho	Tlo	TIO	Cln	160
		-	_	165	Gln				170	_				175	
			180		Ile			185					190		
		195		•	Leu		200					205		_	
	210				Trp	215					220				
225					Ala 230					235					240
_	-			245	Ile			_	250					255	
_	_		260			_		265					270		Leu Gln
		275					280					285			Pro
	290					295					300				Ile
305		-			310					315					320
Thr	Asp	Ile	Ile	Ser 325	Ala	Leu	Val	Thr	Ser 330	Thr	Phe	Ile	Ile	335	Lys
			340		Leu			345					350		
Arg	Leu	Leu 355	Val	Gly	Gly	Lys	Leu 360	Asn	Val	His	Met	Asn 365	Pro	Pro	Gln
	370					375					380				Lys
385					390					395					Cys 400
				405					410					415	
			420					425					430		Ala
Glu	Ser	Val 435		Glu	Glu	Lys	Phe 440		Val	Leu	Phe	Glu 445		Gln	Phe
Ser	Val	Gly	Ser	Asn	Glu	Leu	Val	Phe	Gln	Val	Lys	Thr	Leu	Ser	Leu

	450					455					460					
Pro	Val	Val	Val	Ile	Val	His	Gly	Ser	Gln	Asp	His	Asn	Ala	Thi	r A	la
165					470					475					4	30
Thr				485					490		Gly			49	5	
			500					505			Суѕ		510			
		515	Lys				520				Gly	525				
	530	Val				535					Asn 540					
Leu 545	Glu	Asp	Tyr	Ser	Gly 550	Leu	Ser	Val	Ser	Trp 555	Ser	Gln	Ph∈	As	n A	rg 60
Glu				565					570)	Gln			5/	5	
			580	Leu	Lys			585			His		590)		
		595	,				600				a His	605)			
	610	ı				615					e Sei 620)				
Gly 625		, Ile	• Thi	Ile	Ala 630		Lys	Phe	e Asp	Se 63	r Pro 5	o Glu	ı Ar	g As	sn 1	Leu 540
Trp	Asr			64	o Phe	Thr			65	0	e Se			6	55	
			66)				665	5		e Ty		6/	U		
		67	5				680)			r Th	68	5			
	691	n				699	5				е Lу 70	0				
705					710)				71						120
Met	As			72	5				73	0	ro Gl			,	35	
			74	0				74	5		sp Gl		75	0		
		75	5				76	0			is Vā	76	5			
	77	0				77	5				eu Se 78	30				
70	5				79	0				7	rp Vä 95					800
As	p Pr			80)5				83	10					812	Thr
			83	20				82	25				8	30		His
		83	3.5				84	0				8	45			Lys
	ρg	50				85	55				8	60				Trp
Pr	o Tì	ur L	eu V	al T	nr Th	ır Le	eu Th	ir T	yr G	ly V	al G	ln C	ys P	he	Ser	Arg
86	5				87	70				8	75					880
Τ'n	r P	ro A	sp H		et L <u>y</u> 85	s G.	ın H	ıs A	sp P R	ne F 90	TIE D	ys S	CI P	110	895	Pro
G1	.u G	ly T		al G 00	ln G	Lu A:	rg Tì	nr I 9			he L	ys A	ag A	sp 10		Asn
T	r L	ys T	hr A	rg A	la G	lu V	al L			lu C	Sly A	r ds	hr I	.eu	Va]	. Asn

		915					920					925				
	930					935	Asp				940					
-	His	Lys	Leu	Glu	Tyr 950	Asn	Tyr	Asn	Ser	His 955	Asn	Val	Tyr	Ile	Met 960	
945 Ala	Asp	Lys	Gln	Lys 965		Gly	Ile	Lys	Val 970		Phe	Lys	Ile	Arg 975		
Asn	Ile	Glu	Asp 980		Ser	Val	Gln	Leu 985		Asp	His	Tyr	Gln 990	-	Asn	
Thr	Pro	11e 995		Asp	Gly		Val	Leu	Leu	Pro		Asn 1005		Tyr	Leu	
	Thr 1010		Ser	Ala			Lys		Pro				Arg	Asp	His	
		Leu	Leu	Glu			Thr	Ala	Ala			Thr	Leu			
025	-	_			1030					1035					1040	
Asp	Glu	Leu	_	Lys 1045												
		(2) IN	FORM	ATIO	n Fo	R SE	Q ID	NO:	80:						
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		(D)	TOP	OLOG	Y: 1	inea	r									
	(xi)	SEQU	ENCE	DES	CRIF	MOIT	I: SE	Q II) NO:	80:					
TGG	GATO	CTC	AGGC	CGTC	CT G	CTGC	CCG									28
		(2	?) IN	FORM	IATIC	N FC	R SE	Q II	NO:	81:						
	(i) S	SEQUE	NCE	CHAR	ACTE	ERIST	ICS:								
							pai	rs								
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		(-,														
	ı	(xi)	SEQU	JENCI	E DES	SCRI	OITS	J: SI	EQ II	ON C	:81:					
GTC	TCGA	AGGG	AGCA	ATGG	GCA (CTT	GCG									27
		(2	2) I!	VFORI	OITAN	ON FO	OR SI	EQ II	ОИС	:82:						
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		, 2	, 10.				_									
		(xi)	SEÇ	UENC:	E DE	SCRI	PTIO	N: S	EQ I	D NO	:82:					

(2) INFORMATION FOR SEQ ID NO:83:

TGGGATCCGA GAAGTCTATA TCCCATC

27

(1) SEQUENCE CHARACTERISTICS.	
(A) LENGTH: 28 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:83:	
TGGGATCCTT AGAAGTCTAT ATCCCATC	28
(2) INFORMATION FOR SEQ ID NO:84:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 28 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:84:	
GTCTCGAGCC ATGAACGCCC CCGAGCGG	28
GICICOAGCC AIGAACGCCC CCGAGGGG	
(2) INFORMATION FOR SEQ ID NO:85:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 30 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(b) TOPOLAGY: Timedi	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:85:	
GTGAATTCTC GTCTGATTTC TGGCAGGAGG	30
(2) INFORMATION FOR SEQ ID NO:86:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 30 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:86:	
OCCUPATION DESCRIPTION DE CONCUENT DE CONCUENTA DE CONCUE	30
GTGAATTCTT TACGTCTGAT TTCTGGCAGG	•
(2) INFORMATION FOR SEQ ID NO:87:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 34 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	

34

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:87:

GTCTCGAGCC ATGGACGAAC TGTTCCCCCT CATC

(2) INFORMATION FOR SEQ ID NO:88:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 31 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:88:	
GTGGATCCAA GGAGCTGATC TGACTCAGCA G	31
(2) INFORMATION FOR SEQ ID NO:89:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 32 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:89:	
GTGGATCCTT AGGAGCTGAT CTGACTCAGC AG	32
(2) INFORMATION FOR SEQ ID NO:90:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 32 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:90:	
CCTCCTAAGC TTATCATGGA CCATTATGAT TC	32
(2) INFORMATION FOR SEQ ID NO:91:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 33 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:91:	
	33
CCTCCTGGAT CCCTGCGCAG GATGATGGTC CAG	

(2) INFORMATION FOR SEQ ID NO:92:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 45 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:92:	
GGATGGAAGC TTCAATGGCT GCCATCCGGA AGAAACTGGT GATTG	45
(2) INFORMATION FOR SEQ ID NO:93:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 45 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:93:	
GGATGGGGAT CCTCACAAGA CAAGGCAACC AGATTTTTTC TTCCC	45
(2) INFORMATION FOR SEQ ID NO:94:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 29 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:94:	
GGGAAGCTTC CATGAGCGAG ACGGTCATC	29
(2) INFORMATION FOR SEQ ID NO:95:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 28 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:95:	
CCCGGATCCT CAGGGAGAAC CCCGCTTC	28
(2) INFORMATION FOR SEQ ID NO:96:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 30 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single	

(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:96:	
GTGAATTCGA CCATGGAGCG GCCCCCGGGG	30
(2) INFORMATION FOR SEQ ID NO:97:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 27 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:97:	
GTGGTACCCA TTCTGTTAAC CAACTCC	27
(2) INFORMATION FOR SEQ ID NO:98:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 28 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:98:	
GTGGTACCTC ATTCTGTTAA CCAACTCC	28
(2) INFORMATION FOR SEQ ID NO:99:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 28 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:99:	
GTCTCGAGAG ATGCTGTCCC GTGGGTGG	28
(2) INFORMATION FOR SEQ ID NO:100:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 27 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:100:	
The state of the s	27

GTGAATTCGC TTCCTCTTGA GGGAACC

(2) INFORMATION FOR SEQ 10 NO.101.	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 27 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:101:	
GTGAATTCAC TTCCTCTTGA GGGAACC	27
(2) INFORMATION FOR SEQ ID NO:102:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 29 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:102:	
GTCTCGAGCC ATGGAGAACT TCCAAAAGG	29
(2) INFORMATION FOR SEQ ID NO:103:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 28 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:103:	
GTGGATCCCA GAGTCGAAGA TGGGGTAC	28
(2) INFORMATION FOR SEQ ID NO:104:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 29 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:104:	
GTGGATCCTC AGAGTCGAAG ATGGGGTAC	29
(2) INFORMATION FOR SEQ ID NO:105:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 30 base pairs(B) TYPE: nucleic acid	

(C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:105:	
GTGAATTCGG CGATGCCAGA CCCCGCGGCG	30
(2) INFORMATION FOR SEQ ID NO:106:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 32 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:106:	
GTGGATCCCA GGCACAGGCA GCCTCAGCCT TC	32
(2) INFORMATION FOR SEQ ID NO:107:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 33 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:107:	
GTGGATCCTC AGGCACAGGC AGCCTCAGCC TTC	33
(2) INFORMATION FOR SEQ ID NO:108:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 2616 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA (ix) FEATURE:	
(A) NAME/KEY: Coding Sequence(B) LOCATION: 12613(D) OTHER INFORMATION:	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:108:	
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 1 5 10 15	48
GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly	96

GAG Glu	GGC Gly	GAG Glu 35	GGC Gly	GAT Asp	GCC . Ala	Thr	TAC Tyr 40	GJY GGC	AAG Lys	CTG Leu	ACC Thr	CTG Leu 45	AAG Lys	TTC Phe	ATC Ile	144
TGC Cys	ACC Thr 50	ACC Thr	GGC Gly	AAG Lys	CTG Leu	CCC Pro 55	GTG Val	CCC Pro	TGG Trp	CCC Pro	ACC Thr 60	CTC Leu	GTG Val	ACC Thr	ACC Thr	192
CTG Leu 65	ACC Thr	TAC Tyr	GGC Gly	GTG Val	CAG Gln 70	TGC Cys	TTC Phe	AGC Ser	CGC Arg	TAC Tyr 75	CCC	GAC Asp	CAC His	ATG Met	AAG Lys 80	240
CAG Gln	CAC His	GAC Asp	TTC Phe	TTC Phe 85	AAG Lys	TCC Ser	GCC Ala	ATG Met	CCC Pro 90	GAA Glu	GGC Gly	TAC Tyr	GTC Val	CAG Gln 95	GAG Glu	288
CGC Arg	ACC Thr	ATC	TTC Phe	Phe	AAG Lys	GAC Asp	GAC Asp	GGC Gly 105	AAC Asn	TAC Tyr	AAG Lys	ACC Thr	CGC Arg 110	GCC Ala	GAG Glu	336
GTG Val	AAG Lys	TTC Phe	e Glu	GGC Gly	GAC Asp	ACC Thr	CTG Leu 120		AAC Asn	CGC Arg	: ATC	GAG Glu 125	Leu	AAG Lys	GGC Gly	384
ATC Ile	GAC Asp 130	Phe	C AAC	G GAG	GAC Asp	GGC Gly 135	AAC Asr	: ATC	CTG Leu	GGC Gly	CAC His	: Lys	CTC Lev	GAG Glu	TAC Tyr	432
AA(Asi 14!	ı Tyr	C AAC	C AGO	C CAC	AAC Asn 150	Val	TAT Ty:	r ATC	: ATC	GCC : Ala 159	a Ası	Z AAC D Lys	G CAC	AAC Lys	AAC Asn 160	480
GG(C ATV	C AAG e Ly	G GTY s Va	G AA0 1 Asi 16!	n Phe	: AAG : Lys	TA ;	e Arc	CAC His	s Ası	n Il	C GAG	G GAG L Asj	GG(G) Gly 17	C AGC y Ser 5	528
GT Va	G CA	G CT n Le	C GC u Al 18	a As	C CAC	TAC	CAC	G CAC n Glr 189	n Ası	n Th	C CC r Pr	C AT	C GG e Gl; 19	y As	c GGC p Gly	576
CC Pr	C GT o Va	G CT 1 Le 19	u Le	G CC	C GAG o Asi	AA(Asi	C CA n Hi 20	s Ty:	C CT r Le	G AG u Se	C AC	C CA r Gl 20	n Se	c GC r Al	C CTG a Leu	624
AC Se	C AA er Ly 21	s As	C CC	C AA	C GA(3 AA0 u Ly: 21	s Ar	C GA	T CA p Hi	C AT s Me	G GT et Va 22	ıl L∈	G CT u Le	G GA u Gl	G TTC u Phe	672
GT Va 22	al Th	C GC ir Al	CC GC la Al	C GG la Gl	G ATO y Il 23	e Th	T CI r Le	rc GG eu Gl	С АТ У Ме	G GA et As 23	sp G]	AG CI lu Le	YG TA	C AA	G TCC 's Ser 240	720
G(G]	GA CT	rc Ad eu Ai	GA TO	CT CC er Ar 24	g Al	T CA a Gl	A GC n Al	CT TC La Se	G AA er As 25	n Se	CG G(er A)	CG A	rg co et Pi	co As	AC CCC sp Pro	768
G	CG G(cg ci	AC C	rg co	C TT	C TI	C T	AC GG	C AC	GC A'	rc r	CG C	GT G	CC G	AG GCC	816

Ala	Ala	His	Leu 260	Pro	Phe	Phe	Tyr	Gly 265	Ser	Ile	Ser	Arg	Ala 270	Glu	Ala	
											GGG Gly					864
											CTG Leu 300					912
											CAG Gln					960
											CCG Pro					1008
											TGC Cys					1056
											CCG Pro					1104
											CGC Arg 380					1152
	Glu										AGC Ser					1200
															TAC Tyr	1248
									Glu					Ser	GGG Gly	1296
			Asp					Leu					Glu		GGC Gly	1344
ACA Thr	TAC Tyr 450	Ala	CTG Leu	TCC	CTC	ATC Ile 455	Tyr	GGG Gly	AAG Lys	ACC Thr	GTG Val 460	Tyr	CAC His	TAC	CTC Leu	1392
ATC 11e 465	Ser	CAA Gln	GAC Asp	AAG Lys	GCG Ala 470	Gly	AAC Lys	TAC Tyr	TGC Cys	: ATT : Ile 475	Pro	GAC Glu	GGC GGC	ACC Thr	AAG Lys 480	1440
					Gln					Leu					GAC ASP	1488

														GCC Ala		1536
AAC Asn	GCC Ala	TCA Ser 515	GGG Gly	GCT Ala	GCT Ala	GCT Ala	CCC Pro 520	ACA Thr	CTC Leu	CCA Pro	GCC Ala	CAC His 525	CCA Pro	TCC Ser	ACG Thr	1584
TTG Leu	ACT Thr 530	CAT His	CCT Pro	CAG Gln	AGA Arg	CGA Arg 535	ATC Ile	GAC Asp	ACC Thr	CTC Leu	AAC Asn 540	TCA Ser	GAT Asp	GGA Gly	TAC Tyr	1632
ACC Thr 545	CCT Pro	GAG Glu	CCA Pro	GCA Ala	CGC Arg 550	ATA Ile	ACG Thr	TCC Ser	CCA Pro	GAC Asp 555	AAA Lys	CCG Pro	CGG Arg	CCG Pro	ATG Met 560	1680
CCC Pro	ATG Met	GAC Asp	ACG Thr	AGC Ser 565	GTG Val	TAT Tyr	GAG Glu	AGC Ser	CCC Pro 570	TAC Tyr	AGC Ser	GAC Asp	CCA Pro	GAG Glu 575	GAG Glu	1728
														ATA Ile		1776
GAC Asp	ATT	GAA Glu 595	Leu	GGC Gly	TGC Cys	GGC Gly	AAC Asn 600	TTT Phe	GGC Gly	TCA Ser	GTG Val	CGC Arg 605	CAG Gln	GGC Gly	GTG Val	1824
TAC Tyr	CGC Arg 610	ATG Met	CGC Arg	AAG Lys	AAG Lys	CAG Gln 615	ATC Ile	GAC Asp	GTG Val	GCC Ala	ATC Ile 620	Lys	GTG Val	CTG Leu	AAG Lys	1872
CAG Gln 625	Gly	ACG	GAG Glu	AAG Lys	GCA Ala 630	GAC Asp	ACG Thr	GAA Glu	GAG Glu	ATG Met 635	Met	CGC	GAG	GCG Ala	CAG Gln 640	1920
ATC Ile	ATG Met	CAC	CAG Gln	CTG Leu 645	Asp	AAC Asn	CCC Pro	TAC Tyr	Ile 650	Val	CGG Arg	CTC Lev	ATT	GGC Gly 655	Val	1968
TGC Cys	CAG Gln	GCC Ala	GAG Glu 660	Ala	CTC Leu	ATG Met	Leu	GTC Val	Met	GAC Glu	ATC	GCT Ala	GGC 1 Gly 670	/ Gly	GGG Gly	2016
CCC	CTG	CAC His	Lys	TTC Phe	CTG	GTC Val	680 680	/ Lys	AGG Arg	GAC Glu	G GAC	ATC 1 116 685	Pro	r GTC o Val	AGC Ser	2064
AA1 Asr	r GTC n Val 690	Ala	GAG Glu	CTC Lev	CTC Lev	CAC His 695	Glr	G GTC	TCC Ser	ATY Me	G GGC E Gly 700	/ Mel	AA(G TAC	CTG Leu	2112
GA0 Gl: 70!	ı Glı	AA(3 AAC s Asr	TT:	GTC Val	His	C CG	T GAC	CTC Lev	G GC0 1 Ala 71	a Ala	C CGG	C AAG g Asi	C GT(n Val	C CTG Leu 720	2160
CIX	G GT	r aa	c cc	G CAG	OAT C	G GC	DAA C	YA E	C AGG	G GA	C TT	r gg	C CT	C TC	AAA C	2208

Leu	Val	Asn	Arg	His 725	Tyr	Ala	Lys	Ile	Ser 730	Asp	Phe	Gly	Leu	Ser 735	Lys	
														GGG Gly		2256
TGG Trp	CCG Pro	CTC Leu 755	AAG Lys	TGG Trp	TAC Tyr	GCA Ala	CCC Pro 760	GAA Glu	TGC Cys	ATC Ile	AAC Asn	TTC Phe 765	CGC Arg	AAG Lys	TTC Phe	2304
TCC Ser	AGC Ser 770	CGC Arg	AGC Ser	GAT Asp	GTC Val	TGG Trp 775	AGC Ser	TAT Tyr	GGG Gly	GTC Val	ACC Thr 780	ATG Met	TGG Trp	GAG Glu	GCC Ala	2352
														GAG Glu		2400
ATG Met	GCC Ala	TTC Phe	ATC Ile	GAG G1u 805	Gln	GGC Gly	AAG Lys	CGG Arg	ATG Met 810	GAG Glu	TGC Cys	CCA Pro	CCA Pro	GAG Glu 815	TGT Cys	2448
CCA Pro	CCC Pro	GAA Glu	CTG Leu 820	Tyr	GCA Ala	CTC Leu	ATG Met	AGT Ser 825	Asp	TGC Cys	TGC Trp	ATC Ile	TAC Tyr 830	Lys	TGG Trp	2496
GAG Glu	GAT Asp	CGC Arg 835	Pro	GAC Asp	TTC Phe	CTG	ACC Thr 840	Val	GAG Glu	CAG Gln	CGC Arg	ATG Met 845	Arg	A GCC J Ala	TGT Cys	2544
TAC Tyr	TAC Tyr 850	Ser	CTC	GCC Ala	: AGC	AAG Lys 855	Val	GAA Glu	Gly GGG	CCC Pro	CC! Pro 860	Gl}	AGC Sei	C ACA	A CAG	2592
	Ala				TGT Cys 870	: Ala	TGA	.							·	2616

(2) INFORMATION FOR SEQ ID NO:109:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 871 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:109:

Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 115 120 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 130 135 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Ala Met Pro Asp Pro Ala Ala His Leu Pro Phe Phe Tyr Gly Ser Ile Ser Arg Ala Glu Ala Glu Glu His Leu Lys Leu Ala Gly Met Ala Asp Gly Leu Phe Leu Leu Arg Gln Cys Leu Arg Ser Leu Gly Gly Tyr Val Leu Ser Leu Val His Asp Val Arg Phe His His Phe Pro Ile Glu Arg Gln Leu Asn Gly Thr Tyr Ala Ile Ala Gly Gly Lys Ala His Cys Gly Pro Ala Glu Leu Cys Glu Phe Tyr Ser Arg Asp Pro Asp Gly Leu Pro Cys Asn Leu Arg Lys Pro Cys Asn Arg Pro Ser Gly Leu Glu Pro Gln Pro Gly Val Phe Asp Cys Leu Arg Asp Ala Met Val Arg Asp Tyr Val Arg Gln Thr Trp Lys 370 375 Leu Glu Gly Glu Ala Leu Glu Gln Ala Ile Ile Ser Gln Ala Pro Gln 385 390 Val Glu Lys Leu Ile Ala Thr Thr Ala His Glu Arg Met Pro Trp Tyr His Ser Ser Leu Thr Arg Glu Glu Ala Glu Arg Lys Leu Tyr Ser Gly Ala Gln Thr Asp Gly Lys Phe Leu Leu Arg Pro Arg Lys Glu Gln Gly Thr Tyr Ala Leu Ser Leu Ile Tyr Gly Lys Thr Val Tyr His Tyr Leu Ile Ser Gln Asp Lys Ala Gly Lys Tyr Cys Ile Pro Glu Gly Thr Lys Phe Asp Thr Leu Trp Gln Leu Val Glu Tyr Leu Lys Leu Lys Ala Asp Gly Leu Ile Tyr Cys Leu Lys Glu Ala Cys Pro Asn Ser Ser Ala Ser

505 500 Asn Ala Ser Gly Ala Ala Ala Pro Thr Leu Pro Ala His Pro Ser Thr 520 525 Leu Thr His Pro Gln Arg Arg Ile Asp Thr Leu Asn Ser Asp Gly Tyr 540 535 Thr Pro Glu Pro Ala Arg Ile Thr Ser Pro Asp Lys Pro Arg Pro Met 555 545 550 Pro Met Asp Thr Ser Val Tyr Glu Ser Pro Tyr Ser Asp Pro Glu Glu 570 Leu Lys Asp Lys Lys Leu Phe Leu Lys Arg Asp Asn Leu Leu Ile Ala 585 Asp Ile Glu Leu Gly Cys Gly Asn Phe Gly Ser Val Arg Gln Gly Val 605 600 Tyr Arg Met Arg Lys Lys Gln Ile Asp Val Ala Ile Lys Val Leu Lys 615 620 Gln Gly Thr Glu Lys Ala Asp Thr Glu Glu Met Met Arg Glu Ala Gln 635 630 Ile Met His Gln Leu Asp Asn Pro Tyr Ile Val Arg Leu Ile Gly Val 645 650 Cys Gln Ala Glu Ala Leu Met Leu Val Met Glu Met Ala Gly Gly 670 660 665 Pro Leu His Lys Phe Leu Val Gly Lys Arg Glu Glu Ile Pro Val Ser 685 680 Asn Val Ala Glu Leu Leu His Gln Val Ser Met Gly Met Lys Tyr Leu 690 695 700 Glu Glu Lys Asn Phe Val His Arg Asp Leu Ala Ala Arg Asn Val Leu 705 710 715 Leu Val Asn Arg His Tyr Ala Lys Ile Ser Asp Phe Gly Leu Ser Lys 725 730 Ala Leu Gly Ala Asp Asp Ser Tyr Tyr Thr Ala Arg Ser Ala Gly Lys 745 740 Trp Pro Leu Lys Trp Tyr Ala Pro Glu Cys Ile Asn Phe Arg Lys Phe 760 765 Ser Ser Arg Ser Asp Val Trp Ser Tyr Gly Val Thr Met Trp Glu Ala 775 780 Leu Ser Tyr Gly Gln Lys Pro Tyr Lys Lys Met Lys Gly Pro Glu Val 795 790 Met Ala Phe Ile Glu Gln Gly Lys Arg Met Glu Cys Pro Pro Glu Cys 810 805 Pro Pro Glu Leu Tyr Ala Leu Met Ser Asp Cys Trp Ile Tyr Lys Trp 825 820 Glu Asp Arg Pro Asp Phe Leu Thr Val Glu Gln Arg Met Arg Ala Cys 835 840 845 Tyr Tyr Ser Leu Ala Ser Lys Val Glu Gly Pro Pro Gly Ser Thr Gln Lys Ala Glu Ala Ala Cys Ala

(2) INFORMATION FOR SEQ ID NO:110:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2598 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:

(A) NAME/KEY: Coding Sequence
(B) LOCATION: 1...2595

(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:110:

	-	-	_													
ATG Met 1	CCA Pro	GAC Asp	CCC Pro	GCG Ala 5	GCG Ala	CAC His	CTG Leu	CCC Pro	TTC Phe 10	TTC Phe	TAC Tyr	GGC Gly	AGC Ser	ATC Ile 15	TCG Ser	48
CGT Arg	GCC Ala	GAG Glu	GCC Ala 20	GAG Glu	GAG Glu	CAC His	CTG Leu	AAG Lys 25	CTG Leu	GCG Ala	GGC Gly	ATG Met	GCG Ala 30	GAC Asp	GGG Gly	96
CTC Leu	TTC Phe	CTG Leu 35	CTG Leu	CGC Arg	CAG Gln	TGC Cys	CTG Leu 40	CGC Arg	TCG Ser	CTG Leu	GGC Gly	GGC Gly 45	TAT Tyr	GTG Val	CTG Leu	144
TCG Ser	CTC Leu 50	GTG Val	CAC His	GAT Asp	GTG Val	CGC Arg 55	TTC Phe	CAC His	CAC His	TTT Phe	CCC Pro 60	ATC Ile	GAG Glu	CGC Arg	CAG Gln	192
CTC Leu 65	AAC Asn	GGC Gly	ACC Thr	TAC Tyr	GCC Ala 70	ATT	GCC Ala	GGC	GGC Gly	AAA Lys 75	GCG Ala	CAC His	TGT Cys	GGA Gly	CCG Pro 80	240
GCA Ala	GAG Glu	CTC Leu	TGC Cys	GAG Glu 85	TTC Phe	TAC Tyr	TCG Ser	CGC Arg	GAC Asp 90	CCC Pro	GAC Asp	GGG Gly	CTG Leu	CCC Pro 95	TGC Cys	288
AAC Asn	CTG Leu	CGC Arg	AAG Lys 100	Pro	TGC Cys	AA.C Asn	CGG Arg	CCG Pro 105	Ser	GGC Gly	CTC Leu	GAG Glu	CCG Pro	Gln	CCG Pro	336
GGG Gly	GTC Val	TTC Phe 115	Asp	TGC Cys	CTG Leu	CGA Arg	GAC Asp 120	Ala	ATG Met	GTG Val	CGT Arg	GAC Asp 125	туг	GTG Val	CGC Arg	384
CAG Gln	ACG Thr 130	Tro	AAG Lys	CTG	GAG Glu	GGC Gly 135	Glu	GCC Ala	CTG Leu	GAC Glu	G CAG Glr 140	Ala	ATC	ATC	AGC Ser	432
CAG Gln 145	Ala	CCC	G CAC	GTC n Val	GAC Glu 150	ı Lys	CTC Lev	ATI	GCT Ala	ACC Thr 155	Thi	GCC Ala	CAC a His	GAC Glu	G CGG Arg 160	480
ATC Met	CCC Pro	TGG Tr	TAC Tyz	CAC His	s Sei	C AGO	CTC	ACC Thr	G CGT Arc	g Glu	G GAC	GCC 1 Ala	GA(G CGC 1 Arg 175	AAA A Lys	528
CTT	TAC 1 Ty1	TC: Se:	r GG(c Gly 180	/ Ala	G CAC	G ACC	GA0	GGC Gl ₂ 189	/ Lys	TTO Phe	C CTO	G CT(G AGG L Arg	g Pro	G CGG D Arg	576
AA(Lys	G GAC	G CAG I Gli 19	n Gl	C AC	A TAC	C GCC r Ala	CTY a Let 20	u Sei	CTC r Lei	TAT:	C TA' e Ty:	r GG r Gl; 20	y Ly	G ACC	G GTG r Val	624

		Tyr	CTC													672
	Gly		AAG Lys													720
			GAC Asp													768
			AGC Ser 260													816
			ACG Thr													864
			TAC Tyr													912
	Arg		ATG Met													960
			GAG Glu													1008
			GCT Ala 340													1056
			GTG Val													1104
			AAG Lys													1152
			CAG Gln													1200
			GTC Val													1248
			GGG Gly 420													1296
ATC	CCT	GTG	AGC	TAA	GTG	GCC	GAG	CTG	CTG	CAC	CAG	GTG	TCC	ATG	GGG	1344

Ile	Pro	Val 435	Ser	Asn	Val		Glu 440	Leu	Leu	His	Gln	Val 445	Ser	Met	Gly	
ATG Met	AAG Lys 450	TAC Tyr	CTG Leu	GAG Glu	GAG Glu	AAG Lys 455	AAC Asn	TTT Phe	GTG Val	CAC His	CGT Arg 460	GAC Asp	CTG Leu	GCG Ala	GCC Ala	1392
											AAG Lys					1440
GGC Gly	CTC Leu	TCC Ser	AAA Lys	GCA Ala 485	CTG Leu	GGT Gly	GCC Ala	GAC Asp	GAC Asp 490	AGC Ser	TAC Tyr	TAC Tyr	ACT Thr	GCC Ala 495	CGC Arg	1488
											CCC					1536
TTC Phe	CGC Arg	AAG Lys 515	TTC Phe	TCC Ser	AGC Ser	CGC Arg	AGC Ser 520	GAT Asp	GTC Val	TGG Trp	AGC Ser	ТАТ Туг 525	GGG Gly	GTC Val	ACC Thr	1584
ATG Met	TGG Trp 530	GAG Glu	GCC Ala	TTG Leu	TCC Ser	TAC Tyr 535	GGC Gly	CAG Gln	AAG Lys	CCC	TAC Tyr 540	Lys	AAG Lys	ATG Met	AAA Lys	1632
GGG G1y 545	Pro	GAG Glu	GTC Val	ATG Met	GCC Ala 550	TTC Phe	ATC Ile	GAG Glu	CAG Gln	GGC Gly 555	/ Lys	CGG Arg	ATG Met	GAG Glu	TGC Cys 560	1680
CC? Pro	CCA	GAC Glu	TGT Cys	CCA Pro	Pro	GAA Glu	CTG Leu	TAC Tyr	GCA Ala 570	Lev	ATC Met	AGT Ser	GAC Asp	TGC Cys 575	TGG Trp	1728
ATC Ile	TAC Tyr	: AAC : Lys	7GC Trp 580	Glu	GAT Asp	CGC Arg	CCC	GAC Asp 585	Phe	CTX	G ACC	GTC Val	G GAC L Glu 590	ı Glr	G CGC 1 Arg	1776
AT(Met	G CGA	GC0 J Ala 599	a Cys	TAC Tyr	TAC Tyr	: AGC Ser	CTC Lev	ı Ala	AGC A Ser	AAC Ly:	G GT(s Val	G GAZ 1 Glu 609	ı Gl	G CCC	C CCA Pro	1824
GG(Gl)	2 AGC / Ser 610	Th:	A CAC	G AAC	GCT Ala	GAC Glu 615	ı Ala	r GCC a Ala	TG:	r GCO s Alo	C TGG a Trj 621	p As	r cc	A CC	G GTC o Val	1872
GC6 Al-	a Thi	C ATY	G GTY	G AGG	AAC Lys 630	3 Gl	GAC Glv	G GA0	G CTO	G TT u Ph 63	e Th	C GG r Gl	G GT y Va	G GTV 1 Va	G CCC 1 Pro 640	1920
AT Il	C CTO	G GT u Va	C GAG	G CTY u Let 64"	ı Ası	C GG(C Gly	GA(As)	C GT.	A AAG 1 Ası 65	n Gl	с са у Ні	C AA s Ly	G TT s Ph	C AG e Se 65	C GTG r Val 5	1968
TC Se	c GG r Gl	C GA y Gl	G GG u Gl: 66	y Gl	G GG(u Gl	C GA' y As _l	r GCC p Al	C AC a Th 66	r Ty	c GG r Gl	Ю AA У Lу	G CT s Le	G AC u Th 67	r Le	G AAG u Lys	2016

												CCC Pro 685				2064
												TAC Tyr				2112
												GAA Glu				2160
												TAC Tyr				2208
												CGC Arg				2256
												GGG Gly 765				2304
GAG Glu	TAC Tyr 770	AAC Asn	TAC Tyr	AAC Asn	AGC Ser	CAC His 775	AAC Asn	GTC Val	TAT Tyr	ATC	ATG Met 780	GCC Ala	GAC Asp	AAG Lys	CAG Gln	2352
AAG Lys 785	AAC Asn	GGC Gly	ATC Ile	AAG Lys	GTG Val 790	AAC Asn	TTC Phe	AAG Lys	ATC	CGC Arg 795	CAC His	AAC Asn	ATC	GAG Glu	GAC Asp 800	2400
					Ala					Gln					GGC Gly	2448
GAC Asp	GGC Gly	CCC	GTG Val 820	Leu	CTG Leu	CCC Pro	GAC Asp	AAC Asn 825	His	TAC	: CTG	AGC Ser	ACC Thr 830	Glr	TCC Ser	2496
GCC Ala	CTG Leu	AGC Ser 835	Lys	GAC Asp	CCC Pro	AAC Asn	GAG Glu 840	Lys	CGC Arg	GAT Asp	CAC His	ATG Met 845	. Val	CTC Lev	CTG Leu	2544
GAG Glu	TTC Phe 850	. Val	ACC Thr	GCC Ala	GCC Ala	GGG Gly 855	Ile	ACT Thr	CTC	GGC Gly	ATC Met 860	Asp	GAC Glu	CTC	G TAC 1 Tyr	2592
AAG Lys 865																2598

(2) INFORMATION FOR SEQ ID NO:111:

⁽i) SEQUENCE CHARACTERISTICS:

⁽A) LENGTH: 865 amino acids

- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:111:

Met Pro Asp Pro Ala Ala His Leu Pro Phe Phe Tyr Gly Ser Ile Ser 10 Arg Ala Glu Ala Glu Glu His Leu Lys Leu Ala Gly Met Ala Asp Gly 25 20 Leu Phe Leu Leu Arg Gln Cys Leu Arg Ser Leu Gly Gly Tyr Val Leu 40 Ser Leu Val His Asp Val Arg Phe His His Phe Pro Ile Glu Arg Gln 55 Leu Asn Gly Thr Tyr Ala Ile Ala Gly Gly Lys Ala His Cys Gly Pro 75 70 Ala Glu Leu Cys Glu Phe Tyr Ser Arg Asp Pro Asp Gly Leu Pro Cys 85 90 Asn Leu Arg Lys Pro Cys Asn Arg Pro Ser Gly Leu Glu Pro Gln Pro 105 110 100 Gly Val Phe Asp Cys Leu Arg Asp Ala Met Val Arg Asp Tyr Val Arg 125 120 Gln Thr Trp Lys Leu Glu Gly Glu Ala Leu Glu Gln Ala Ile Ile Ser 140 135 Gln Ala Pro Gln Val Glu Lys Leu Ile Ala Thr Thr Ala His Glu Arg 155 150 Met Pro Trp Tyr His Ser Ser Leu Thr Arg Glu Glu Ala Glu Arg Lys 165 170 Leu Tyr Ser Gly Ala Gln Thr Asp Gly Lys Phe Leu Leu Arg Pro Arg 185 180 Lys Glu Gln Gly Thr Tyr Ala Leu Ser Leu Ile Tyr Gly Lys Thr Val 195 200 205 Tyr His Tyr Leu Ile Ser Gln Asp Lys Ala Gly Lys Tyr Cys Ile Pro 220 215 Glu Gly Thr Lys Phe Asp Thr Leu Trp Gln Leu Val Glu Tyr Leu Lys 230 235 Leu Lys Ala Asp Gly Leu Ile Tyr Cys Leu Lys Glu Ala Cys Pro Asn 250 245 Ser Ser Ala Ser Asn Ala Ser Gly Ala Ala Ala Pro Thr Leu Pro Ala 265 His Pro Ser Thr Leu Thr His Pro Gln Arg Arg Ile Asp Thr Leu Asn 280 Ser Asp Gly Tyr Thr Pro Glu Pro Ala Arg Ile Thr Ser Pro Asp Lys 295 Pro Arg Pro Met Pro Met Asp Thr Ser Val Tyr Glu Ser Pro Tyr Ser 310 315 Asp Pro Glu Glu Leu Lys Asp Lys Lys Leu Phe Leu Lys Arg Asp Asn 330 325 Leu Leu Ile Ala Asp Ile Glu Leu Gly Cys Gly Asn Phe Gly Ser Val 345 340 Arg Gln Gly Val Tyr Arg Met Arg Lys Lys Gln Ile Asp Val Ala Ile 360 365 Lys Val Leu Lys Gln Gly Thr Glu Lys Ala Asp Thr Glu Glu Met Met 375 380

Arg Glu Ala Gln Ile Met His Gln Leu Asp Asn Pro Tyr Ile Val Arg

205					390					395					400
385 Leu :	Ile	Glv	Val			Ala	Glu	Ala			Leu	Val	Met	Glu	Met
				405					410					415	
Ala(Gly	Gly	Gly 420	Pro	Leu	His	Lys	Phe 425	Leu	Val	Gly	Lys	Arg 430	Glu	Glu
Ile :	Pro	Val 435	Ser	Asn	Val	Ala	Glu 44 0	Leu	Leu	His	Gln	Val 445	Ser	Met	Gly
Met :	Lys 450	Tyr	Leu	Glu	Glu	Lys 455	Asn	Phe	Val	His	Arg 460	Asp	Leu	Ala	Ala
Arg . 465		Val	Leu	Leu	Val 470	Asn	Arg	His	Tyr	Ala 475	Lys	Ile	Ser	Asp	Phe 480
Gly	Leu	Ser	Lys	Ala 485		Gly	Ala	Asp	Asp	Ser	Tyr	Tyr	Thr	Ala 495	Arg
Ser	Ala	Gly	Lys 500		Pro	Leu	Lys	Trp 505		Ala	Pro	Glu	Cys 510	Ile	Asn
Phe	Arg	Lys 515	Phe	Ser	Ser	Arg	Ser 520	Asp	Val	Trp	Ser	Tyr 525	Gly	Val	Thr
	530	Glu				535	Gly				540				
545					550		Ile			555					560
				565			Leu		570					575	
			580				Pro	585					590		
		595					600					605			Pro
	610					615					620				Val
625					630					635					Pro 640
				645					650	l				655	
			660					665					670)	Lys
		675					680					685			ı Val
	690					695	5				700				His
705					710	1				715	,				720
				725	,				730)				735	
			740)				745	5				750)	ı Leu
		755	5				760)				765	5		s Leu
	770)				775	5				780)			s Gln
785					790)				799	5				u Asp
				809	5				810	0				81	
			820)				82	5				83	0	n Ser
		83	5				840)				84	5		u Leu
Glu	Phe	e Va	l Thu	r Ala	a Ala	a Gl	y Ile	e Th	r Le	u G1	y Me	t As	p Gl	u Le	u Tyr

850 855 860

Lys 865

(2) INFORMATION FOR SEQ ID NO:112:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1635 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence

(B) LOCATION: 1...1632

(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:112:

ATG Met 1	GAG Glu	AAC Asn	TTC Phe	CAA Gln 5	AAG Lys	GTG Val	GAA Glu	AAG Lys	ATC Ile 10	GGA Gly	GAG Glu	GGC Gly	ACG Thr	TAC Tyr 15	GGA Gly	48
									ACG Thr							96
AAG Lys	AAA Lys	ATC Ile 35	CGC Arg	CTG Leu	GAC Asp	ACT Thr	GAG Glu 40	ACT Thr	GAG Glu	GGT Gly	GTG Val	CCC Pro 45	AGT Ser	ACT Thr	GCC Ala	144
									CTT Leu							192
AAG Lys 65	CTG Leu	CTG Leu	GAT Asp	GTC Val	ATT Ile 70	CAC His	ACA Thr	GAA Glu	AAT Asn	AAA Lys 75	CTC Leu	TAC Tyr	CTG Leu	GTT Val	TTT Phe 80	240
GAA Glu	TTT Phe	CTG Leu	CAC His	CAA Gln 85	GAT Asp	CTC Leu	AAG Lys	AAA Lys	TTC Phe 90	ATG Met	GAT Asp	GCC Ala	TCT Ser	GCT Ala 95	CTC Leu	285
									AGC Ser					Leu	CTC Leu	336
CAG Gln	GGC Gly	CTA Leu 115	Ala	TTC Phe	TGC Cys	CAT His	TCT Ser 120	His	CGG	GTC Val	CTC Leu	CAC His	Arg	. GAC Asp	CTT Leu	384
AAA Lys	CCT Pro 130	CAG Gln	AAT Asn	CTG Leu	CTT Leu	ATT Ile 135	Asn	ACA Thr	GAG Glu	GGG Gly	GCC Ala 140	Ile	: AAG : Lys	CTA Leu	GCA Ala	432
GAC	TTT	GGA	CTA	GCC	AGA	GCT	TTI	GGA	GTC	CCI	GTI	CGT	ACT	TAC	2 ACC	480

Asp I 145	Phe	Gly	Leu	Ala	Arg 150	Ala	Phe	Gly		Pro 155	Va1	Arg	Thr	Tyr	Thr 160	
CAT (GAG Glu	GTG Val	GTG Val	ACC Thr 165	CTG Leu	TGG Trp	TAC Tyr	CGA Arg	GCT Ala 170	CCT Pro	GAA Glu	ATC Ile	CTC Leu	CTG Leu 175	GGC Gly	528
TCG A	AAA Lys	тат туг	ТАТ Туг 180	TCC Ser	ACA Thr	GCT Ala	GTG Val	GAC Asp 185	ATC Ile	TGG Trp	AGC Ser	CTG Leu	GGC Gly 190	TGC Cys	ATC Ile	576
TTT (GCT Ala	GAG Glu 195	ATG Met	GTG Val	ACT Thr	CGC Arg	CGG Arg 200	GCC Ala	CTG Leu	TTC Phe	CCT Pro	GGA Gly 205	GAT Asp	TCT Ser	GAG Glu	624
Ile	GAC Asp 210	CAG Gln	CTC Leu	TTC Phe	CGG Arg	ATC Ile 215	TTT Phe	CGG Arg	ACT Thr	CTG Leu	GGG Gly 220	ACC Thr	CCA Pro	GAT Asp	GAG Glu	672
GTG Val 225	GTG Val	TGG Trp	CCA Pro	GGA Gly	GTT Val 230	ACT Thr	TCT Ser	ATG Met	CCT Pro	GAT Asp 235	TAC Tyr	AAG Lys	CCA Pro	AGT Ser	TTC Phe 240	720
CCC Pro	AAG Lys	TGG Trp	GCC Ala	CGG Arg 245	CAA Gln	GAT Asp	TTT Phe	AGT Ser	AAA Lys 250	GTT Val	GTA Val	CCT Pro	CCC Pro	CTG Leu 255	GAT Asp	768
GAA Glu	GAT Asp	GGA Gly	CGG Arg 260	AGC Ser	TTG Leu	TTA Leu	TCG Ser	CAA Gln 265	ATG Met	CTG Leu	CAC His	TAC Tyr	GAC Asp 270	CCT Pro	AAC Asn	816
AAG Lys	CGG Arg	ATT Ile 275	Ser	GCC Ala	AAG Lys	GCA Ala	GCC Ala 280	Leu	GCT Ala	CAC	CCT Pro	TTC Phe 285	Phe	CAG Gln	GAT Asp	86 4
GTG Val	ACC Thr 290	Lys	CCA Pro	GTA Val	CCC Pro	CAT His 295	Leu	CGA Arg	CTC Leu	TGC Trp	GAT Asp 300	Pro	CCG Pro	GTC Val	GCC Ala	912
ACC Thr 305	ATG Met	GTC Val	G AGC Ser	: AAC	GGC Gly 310	Glu	GAC Glu	CTC Leu	TTC Phe	ACC Thr	G13	GTC Val	GTC Val	CCC Pro	ATC Ile 320	960
CTG Leu	GTC Val	GAC Glu	CTC	GAC Asp 325	o Gly	GAC Asp	GTA Val	A AAC L Asr	330	, His	AAC Lys	G TTC S Phe	C AGO	GTC Val 335	G TCC Ser	1008
GGC Gly	GAC Glu	GGG Gly	GAC Glu 340	ı Gly	C GAT y Asp	GCC Ala	ACC Thi	TAC Ty:	Gly	AA(G CTY	G ACC	C CTV c Let 350	ı Lys	TTC Phe	1056
ATC Ile	TG(2 ACC 35!	c Thi	GGC Gl	C AAC y Lys	G CTC	9 CC0 1 Pro 36	o Vai	G CCC	TGG Trj	G CCG	C ACC O Thi	r Le	C GTY	G ACC l Thr	1104
ACC Thr	CTY Lev	ı Th	C TAC	c GGG	C GTG y Val	G CAG 1 Glr 379	1 Су	C TTV s Ph	C AGG e Se:	C CG	С ТА g Ту 38	r Pr	C GA	C CA	C ATG s Met	1152

AAG Lys 385	CAG Gln	CAC His	GAC Asp	TTC Phe	TTC Phe 390	AAG Lys	TCC Ser	GCC Ala	ATG Met	CCC Pro 395	GAA Glu	GGC	TAC Tyr	GTC Val	CAG Gln 400		1200
						AAG Lys											1248
						GAC Asp											1296
						GAC Asp											1344
						AAC Asn 455											1392
						TTC Phe											1440
															GAC Asp		1488
GGC Gly	CCC	GTG Val	CTG Leu 500	CTG Leu	CCC	GAC Asp	AAC Asn	CAC His	Tyr	CTG Leu	AGC Ser	ACC Thr	CAG Gln 510	Ser	GCC Ala		1536
			Asp					Arg					Lev		GAG Glu		1584
TTC Phe	GTG Val	Thr	GCC Ala	GCC Ala	GGG Gly	ATC 11e 535	Thr	CTC	GGC Gly	ATC	GAC Asr 540	Glu	CTC Lev	TAC 1 TY1	AAG Lys	т	1633
AA																	1635

(2) INFORMATION FOR SEQ ID NO:113:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 544 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:113:

Met Glu Asn Phe Gln Lys Val Glu Lys Ile Gly Glu Gly Thr Tyr Gly
1 5 10 15

Val V	/al	Tyr	Lys 20	Ala	Arg	Asn	Lys	Leu 25	Thr	Gly	Glu	Val	Val 30	Ala	Leu
Lys I	_	Ile 35	Arg	Leu	Asp	Thr	Glu 40	Thr	Glu	Gly	Val	Pro 45	Ser	Thr	Ala
Ile i			Ile	Ser	Leu	Leu 55	Lys	Glu	Leu	Asn	His 60	Pro	Asn	Ile	Val
Lys 1		Leu	Asp	Val	Ile 70	His	Thr	Glu	Asn	Lys 75	Leu	Tyr	Leu	Val	Phe 80
Glu				85					90					95	
Thr			100					105					110		
Gln		115					120					125			
Lys	130					135					140				
Asp 145					150					155					160
				165			Tyr		170					175	
			180				Val	185					190		
		195					Arg 200					205			
	210					215	Phe				220				
225					230		Ser			235					240
				245					250					255	
			260					265					270		Asn
		275					280					285			Asp
	290					295					300				Ala
305					310					315					320
				325	•				330)				335	
			340)				345					350)	Phe Thr
		355	5				360	1				365	5		l Thr
	370					375	5				380)			l Gln
385					390)				395	5				400
				405	5				410)				41	g Ala 5 u Lys
			420)				425	5				430)	u Glu
		435	5				440)				445	5		
	450)				45	5				460)			n Lys
Asn 465		7 110	e ry:	s va.	476		= rÀs	o 11€	= wr	4 7.			. 01	ر در د	p Gly 480

Ser	Val	G	ln 1		Ala 485	Asp	His	Tyr		Gln 490	Asn	Thr	Pro :	Ile	GLY 495	Asp	
Gly	Pro	v	al 1			Pro	Asp			Tyr	Leu	Ser	Thr		Ser	Ala	
	6			500	Dro	λεη	Clu		505 Ara	Asn	His	Met	Val	510 Leu	Leu	Glu	
		5	15					520					525				
Phe	Val 530		hr.	Ala	Ala	Gly	Ile 535	Thr	Leu	Gly	Met	Asp 540	Glu	Leu	Tyr	Lys	
			(2)	INF	ORMA	TION	, FOF	SEQ	ID	NO:1	14:						
	,						ACTER bas										
		((B)	TYPE	: n	ıcle:	ic ac	id									
							S: si inear		2								
		1	(ע)	TOPC	LCG.		rnear	•									
		•		OLEC		TYP	E: cI	ONA									
			(A)	NAI	Œ/K	EY: (Codi	ng Se	eque	nce							
			(B)	LO	ITAC	ON:	1	1632									
			(D)	OT	HER	INFO	RMAT:	ION:									
		(×:	i) 5	EQU!	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	114:					
		_	300	N N C	ccc	Cac	GAG	CTG	بالمائث	ACC	GGG	GTG	GTG	CCC	ATC	CTO	3 48
Met	. Va	1	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Le	ג
1					5					10					15		
GTC	G GA	G	CTG	GAC	GGC	GAC	GTA	AAC	GGC	CAC	AAG	TTC	AGC	GTG	TCC	GG(c 96
Va.	l Gl	u	Leu	Asp	Gly	Asp	val	Asn	Gly	His	Lys	Phe	e Ser	Val	Ser	G1	Y
				20					25					30			
GAG	G GC	æ	GAG	GGC	GAT	GCC	ACC	TAC	GGC	AAC	CTO	ACC	CTG	AAG	TTC	TA	C 144
Gl	u G3	У		Gly	Asp	Ala	Thr	Tyr 40	Gly	. Lys	Let	ı Thi	Leu 45	Lys	Phe	• II	е
			35														
TG	CAC	CC	ACC	GGC	AAC	CTC	CCC	GTG	CCC	TGC	CCC	ACC	CTC	GTC	ACC	C AC	C 192
Cy:	s Th 50		Thr	Gly	Lys	s Lev	1 Pro) vai	Pro	111	PIC	60	r Leu	val			-
													- 010		3 NOV	~ ~ ~	.G 240
CT	G AG	CC	TAC	GGC	GTC	G CAG	TGC	TTC Phe	: AGC	CGC Arc	CVT t	r Pr	C GAC	His	. And	s AA t Ly	.G 240 'S
65		11	ıyı	O ₁ y	vu.	70	. 0,1				75		_			80)
				, mmc	n mmv	~ >>/	י יייי		יייי אייי	- 000	- GA	A GG	C TAC	· GTY	CA(G GA	AG 288
CA G1	G C. n H.	AC is	ASE	: TTC > Phe	Pne	e Ly:	s Sei	Ala	Met	e Pro	o Gli	u Gl	у Тул	Va.	Gl	n Gl	.u
			•		85					90					95		
CG	- L		ΥΤ .4	ጉ ጥጥር	ידידי	C AA	G GA	GAC	GG(C AA	C TA	C AA	G ACC	CGG	G GC	C GA	AG 336
Ar	g T	hr	Ile	Phe	e Ph	e Ly	s As	o Asi	o Gl	y As:	n Ty	r Ly	s Thi	r Ar	g Al	a Gl	lu
				100)				10	5				11	U		
GI	YG A	AG	TTC	GAG	G GG	C GA	C AC	CT	G GTV	g aa	c cg	C AT	C GA	G CT	g AA	.G G(GC 384
Va	l L	ys	Phe	e Glu	ı Gl	y As	p Th	r Le	u Va	l As	n Ar	g Il	e Gl	u Le	u Ly	's G	ly
			115	5				12	O				12	כ			

ATC Ile	GAC Asp 130	TTC Phe	AAG Lys	GAG Glu	GAC Asp	GGC . Gly . 135	AAC Asn	ATC Ile	C'I'G Leu	GGG Gly	CAC His	AAG Lys	CTG Leu	GAG Glu	TAC Tyr	432
AAC Asn 145	TAC Tyr	AAC Asn	AGC Ser	CAC His	AAC Asn 150	GTC Val	TAT Tyr	ATC Ile	ATG Met	GCC Ala 155	GAC Asp	AAG Lys	CAG Gln	AAG Lys	AAC Asn 160	480
GGC Gly	ATC Ile	AAG Lys	GTG Val	AAC Asn 165	TTC Phe	AAG Lys	ATC Ile	CGC Arg	CAC His 170	AAC Asn	ATC Ile	GAG Glu	GAC Asp	GGC Gly 175	AGC Ser	528
GTG Val	CAG Gln	CTC Leu	GCC Ala 180	GAC Asp	CAC His	TAC Tyr	CAG Gln	CAG Gln 185	AAC Asn	ACC Thr	CCC Pro	ATC Ile	GGC Gly 190	GAC Asp	GGC Gly	576
CCC Pro	GTG Val	CTG Leu 195	CTG Leu	CCC Pro	GAC Asp	AAC Asn	CAC His 200	TAC Tyr	CTG Leu	AGC Ser	ACC Thr	CAG Gln 205	Ser	GCC Ala	CTG Leu	624
AGC Ser	AAA Lys 210	GAC Asp	CCC	AAC Asn	GAG Glu	AAG Lys 215	CGC Arg	GAT Asp	CAC His	ATG Met	GTC Val 220	Leu	CTG Leu	GAG Glu	TTC Phe	672
GTG Val 225	Thr	GCC	GCC Ala	GGG Gly	ATC Ile 230	ACT Thr	CTC Leu	GGC Gly	ATG Met	GAC Asp 235	Glu	CTC Lev	ТАС 1 Тут	AAC Lys	S TCC Ser 240	720
GGA Gly	CTC Leu	AGA Arg	TCT Ser	CGA Arg 245	, Ala	ATG Met	GAG Glu	AAC Asn	TTC Phe 250	Glr	AAC Lys	GT(G GAZ L Glu	A AAC 1 Lys 255	ATC Ile	768
GGA Gly	GAG Glu	G17 GGC	260	туг	GGA Gly	GTT Val	GTC Val	TAC Tyr 265	Lys	GCC Alá	AGA A Arg	AAA iaA g	270	s Lei	ACG Thr	816
GGA Gly	A GAC	GTC Val 279	l Va	GCC L Alá	G CTI	AAG Lys	AAA Lys 280	: Ile	C CGC	CTC J Lev	G GAG	C AC o Th 28	r Gl	G AC' u Thi	r GAG r Glu	864
GG1 G13	r GTC 7 Val 290	Pro	C AG'	r ACT	r GCC r Ala	295	Arg	A GAG	TA E	TC' Se:	r CTG r Le	u Le	T AA u Ly	G GA	G CTT u Leu	912
AAC Asi 30!	n His	r cc	'AA T	r AT	r GT0 e Va: 310	l Lys	CTO	G CTY	G GA' u Asj	r GT p Va 31	1 11	т СА е Ні	C AC s Th	A GA r Gl	A AAT u Asn 320	960
AA: Ly:	A CTY	TA L Ty	C CT r Le	G GT u Va 32	1 Ph	r GA/ e Glu	A TT	T CT e Le	G CA u Hi 33	s Gl	A GA n As	T CI p Le	C AA	G AA 's Ly 33	A TTC s Phe	1008
ATY Me	G GA' t As	T GC p Al	C TC a Se 34	r Al	T CT a Le	C ACT	r GG c Gl	C AT y Il 34	e Pr	T CT o Le	T CC	C CI	C AT u Il 35	e Ly	AG AGC vs Ser	1056
ТА Ту	T CT r Le	G TI u Ph	C CA	G CT n Le	G CT	C CAG	G GG n Gl	с ст у Le	'A GC	T TI a Ph	ie C?	C C/ /s H:	AT TO	er Hi	AT CGG is Arg	1104

360 355 GTC CTC CAC CGA GAC CTT AAA CCT CAG AAT CTG CTT ATT AAC ACA GAG 1152 Val Leu His Arg Asp Leu Lys Pro Gln Asn Leu Leu Ile Asn Thr Glu 380 375 370 GGG GCC ATC AAG CTA GCA GAC TTT GGA CTA GCC AGA GCT TTT GGA GTC 1200 Gly Ala Ile Lys Leu Ala Asp Phe Gly Leu Ala Arg Ala Phe Gly Val 395 390 385 CCT GTT CGT ACT TAC ACC CAT GAG GTG GTG ACC CTG TGG TAC CGA GCT 1248 Pro Val Arg Thr Tyr Thr His Glu Val Val Thr Leu Trp Tyr Arg Ala 410 415 405 CCT GAA ATC CTC CTG GGC TCG AAA TAT TAT TCC ACA GCT GTG GAC ATC 1296 Pro Glu Ile Leu Leu Gly Ser Lys Tyr Tyr Ser Thr Ala Val Asp Ile 430 425 420 TGG AGC CTG GGC TGC ATC TTT GCT GAG ATG GTG ACT CGC CGG GCC CTG Trp Ser Leu Gly Cys Ile Phe Ala Glu Met Val Thr Arg Arg Ala Leu 445 435 440 TTC CCT GGA GAT TCT GAG ATT GAC CAG CTC TTC CGG ATC TTT CGG ACT 1392 Phe Pro Gly Asp Ser Glu Ile Asp Gln Leu Phe Arg Ile Phe Arg Thr 460 455 450 CTG GGG ACC CCA GAT GAG GTG GTG TGG CCA GGA GTT ACT TCT ATG CCT 1440 Leu Gly Thr Pro Asp Glu Val Val Trp Pro Gly Val Thr Ser Met Pro 475 470 GAT TAC AAG CCA AGT TTC CCC AAG TGG GCC CGG CAA GAT TTT AGT AAA 1488 Asp Tyr Lys Pro Ser Phe Pro Lys Trp Ala Arg Gln Asp Phe Ser Lys 490 485 GTT GTA CCT CCC CTG GAT GAA GAT GGA CGG AGC TTG TTA TCG CAA ATG Val Val Pro Pro Leu Asp Glu Asp Gly Arg Ser Leu Leu Ser Gln Met

CAC CCT TTC TTC CAG GAT GTG ACC AAG CCA GTA CCC CAT CTT CGA CTC T 1633 His Pro Phe Phe Gln Asp Val Thr Lys Pro Val Pro His Leu Arg Leu 540 535

1584

1635

505

CTG CAC TAC GAC CCT AAC AAG CGG ATT TCG GCC AAG GCA GCC CTG GCT

Leu His Tyr Asp Pro Asn Lys Arg Ile Ser Ala Lys Ala Ala Leu Ala 520

(2) INFORMATION FOR SEQ ID NO:115:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 544 amino acids
 - (B) TYPE: amino acid

500

515

GA

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:115:

1				5	Glu				10					15	
			20		Asp			25					30		
		35			Ala		40					45			
	50				Leu	55					60				
65	Thr				Gln 70					75					80
				85	Lys				90					95	
			100		Lys			105					110		
		115			Asp		120					125			
	130				Asp	135					140				
Asn 145	Tyr	Asn	Ser	His	Asn 150	Val	Tyr	Ile	Met	Ala 155	Asp	Lys	GIn	Lys	160
Gly	Ile	Lys	Val	Asn 165	Phe	Lys	Ile	Arg	His 170		Ile	Glu	Asp	Gly 175	Ser
			180		His			185					190		
		195			Asp		200					205			
	210				Glu	215					220				
225					11e 230					235					240
				245	Ala				250)				255	
			260)	Gly			265					270		
		275	5				280					285			Glu
	290)				295					300)			Leu
Asr 305		Pro) Asi	ı Ile	val 310		Leu	Leu	Ası	o Val 315		e His	Thr	GIU	320
Lys	Leu			325	Phe	Glu			330	3				335	
			34	0				345	,				350	}	Ser
		35	5				360)				365	5		Arg
	370	0				375	5				38	0			c Glu
Gl ₃ 38		a Il	e Ly	s Le	Ala د 390) Phe	e Gl3	/ Le	u Al. 39		g Ala	a Phe	e GI	y Val 400
Pro	o Va	l Ar	g Th	r Ty:	c Thr		s Glu	ı Val	l Va 41		r Le	u Tr	р Ту:	r Arg	g Ala 5
Pro	o Gl	u Il	e Le 42	u Le		/ Sei	r Lys	5 Ty:	с Ту		r Th	r Al	a Va 43	l As _l O	p Ile
Tr	p Se	r Le			s Ile	e Phe	e Ala	a Glu	ı Me	t Va	l Th	r Ar	g Ar	g Al	a Leu

Phe Pro	435					440					445				
450					455					460					
Leu Gly	Thr I	Pro A			Val	Val	Trp			Val	Thr	Ser	Met		
465 Asp Tyr	Lys 1		Ser :	470 Phe	Pro	Lys		Ala	475 Arg	Gln	Asp	Phe		480 Lys	
Val Val		Pro 1	485 Leu .	Asp (Glu	Asp	Gly	490 Arg	Ser	Leu	Leu		495 Gln	Met	
Leu His	Tyr .	500 Asp :	Pro .	Asn	Lys		505 Ile	Ser	Ala	Lys		510 Ala	Leu	Ala	
His Pro 530	515 Phe	Phe (Gln		Val 535	520 Thr	Lys	Pro	Val	Pro 540	525 His	Leu	Arg	Leu	
	(2)	INF	ORMA	TION	FOF	SEC) ID	NO:1	16:						
(:	, ,	LENG	TH:	HARA 2532 clei	bas	se pa									
	(C)	STRA	NDED	NESS : li	: si	ngle	e								
7	ii) M ix) F			TYPE	: cI	ONA									
	(B)	LOC	CITA	EY: C ON: 1 INFOR		2529	equer	nce							
t:	vil S														
١.	<u> </u>	SEQUE	NCE	DESC	RIP	LION	; SE	2 10	NO:	116:					
ATG GTG Met Val	AGC	AAG	GGC	GAG	GAG	CTG	TTC	ACC	GGG	GTG	GTG Val	CCC Pro	ATC Ile 15	CTG Leu	48
ATG GTG Met Val	AGC Ser	AAG Lys GAC	GGC Gly 5 GGC	GAG Glu GAC	GAG Glu GTA	CTG Leu AAC	TTC Phe GGC	ACC Thr 10	GGG Gly AAG	GTG Val	Val AGC	Pro	Ile 15 TCC	Leu	48 96
ATG GTG Met Val 1 GTC GAG	AGC Ser CTG Leu	AAG Lys GAC Asp 20 GGC	GGC Gly 5 GGC Gly	GAG Glu GAC Asp	GAG Glu GTA Val	CTG Leu AAC Asn	TTC Phe GGC Gly 25 GGC	ACC Thr 10 CAC His	GGG Gly AAG Lys	GTG Val TTC Phe	Val AGC Ser	GTG Val 30	Ile 15 TCC Ser	Leu GGC Gly	
ATG GTG Met Val 1 GTC GAG Val Glu GAG GGG	AGC Ser CTG Leu GAG Glu 35	AAG Lys GAC Asp 20 GGC Gly	GGC Gly 5 GGC Gly GAT Asp	GAG Glu GAC Asp GCC Ala	GAG Glu GTA Val ACC Thr	CTG Leu AAC Asn TAC Tyr 40	TTC Phe GGC Gly 25 GGC Gly	ACC Thr 10 CAC His AAG Lys	GGG Gly AAG Lys CTG Leu	GTG Val TTC Phe ACC Thr	AGC Ser CTG Leu 45	GTG Val 30 AAG Lys	TCC Ser TTC Phe	GGC GGY ATC FILE	96
ATG GTG Met Val 1 GTC GAG Val Glu GAG GGC Glu Gly TGC ACC Cys Thr	AGC Ser CTG Leu GAG Glu 35 ACC Thr	AAG Lys GAC Asp 20 GGC Gly GGC Gly	GGC Gly 5 GGC Gly GAT Asp AAG Lys	GAG Glu GAC Asp GCC Ala CTG Leu	GAG Glu GTA Val ACC Thr	CTG Leu AAC Asn TAC Tyr 40 GTG Val	TTC Phe GGC Gly 25 GGC Gly CCC Pro	ACC Thr 10 CAC His AAG Lys TGG Trp	GGG Gly AAG Lys CTG Leu CCC Pro	GTG Val TTC Phe ACC Thr 60	AGC Ser CTG Leu 45 CTG Leu GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	GTG Val 30 AAG Lys Val	TCC Ser TTC Phe ACC	GGC GGY ATC Ile	96 144
ATG GTG Met Val 1 GTC GAG Val Glu GAG GGC Glu Gly TGC ACC Cys Thr 50 CTG ACC Leu Thr	AGC Ser CTG Leu GAG Glu 35 ACC Thr	AAG Lys GAC Asp 20 GGC Gly GGC Gly	GGC Gly 5 GGC Gly GAT Asp AAG Lys	GAG Glu GAC Asp GCC Ala CTG Leu CAG Gln 70	GAG Glu GTA Val ACC Thr CCC Pro 55 TGC Cys	AAC Asn TAC Tyr 40 GTG Val	TTC Phe GGC Gly 25 GGC Gly Pro	ACC Thr 10 CAC His AAG Lys TCG Trp CGC Arg	GGG Gly AAG Lys CTG Leu CCC Pro	GTG Val TTC Phe ACC Thr ACC Thr 60	AGC Ser CTG Leu 45 CTG Leu 65 CTG	GTG Val 30 AAG Lys Val Val CAC His	Ilee 15 TCC Ser TTC Phe TTC: ACC Thu	GGC GGY ATC Ile CACC Thr	96 144 192

GTG Val	AAG Lys	TTC Phe 115	GAG Glu	GGC Gly	GAC . Asp	ACC Thr	CTG Leu 120	GTG Val	AAC Asn	CGC Arg	ATC Ile	GAG Glu 125	CTG Leu	AAG Lys	GGC Gly	384
ATC Ile	GAC Asp 130	TTC Phe	AAG Lys	GAG Glu	Asp	GGC Gly 135	AAC Asn	ATC Ile	CTG Leu	GGG Gly	CAC His 140	AAG Lys	CTG Leu	GAG Glu	TAC Tyr	432
AAC Asn 145	TAC Tyr	AAC Asn	AGC Ser	CAC His	AAC Asn 150	GTC Val	ТАТ Тух	ATC Ile	ATG Met	GCC Ala 155	GAC Asp	AAG Lys	CAG Gln	AAG Lys	AAC Asn 160	480
GGC Gly	ATC Ile	AAG Lys	GTG Val	AAC Asn 165	TTC Phe	AAG Lys	ATC Ile	CGC Arg	CAC His 170	AAC Asn	ATC Ile	GAG Glu	GAC Asp	GGC Gly 175	AGC Ser	528
GTG Val	CAG Gln	CTC Leu	GCC Ala 180	Asp	CAC His	TAC Tyr	CAG Gln	CAG Gln 185	AAC Asn	ACC Thr	CCC	ATC	GGC Gly 190	Asp	GGC Gly	576
CCC Pro	GTG Val	CTG Leu 195	Leu	CCC Pro	GAC Asp	AAC Asn	CAC His 200	Tyr	CTG Leu	AGC Ser	ACC	CAG Gln 205	Ser	GCC Ala	CTG Leu	624
AGC Ser	AAA Lys 210	Asp	CCC Pro	AAC Asn	GAG Glu	AAG Lys 215	Arg	GAT Asp	CAC His	ATG Met	GTC Val 220	Leu	CTC Lev	GAC 1 Glu	TTC Phe	672
GT0 Val 225	Thr	GCC Ala	GCC Ala	GGG Gly	ATC Ile 230	Thr	CTC	GGC Gly	ATG Met	GAC Asp 235	Gli	G CTC	TAC	AAC Lys	G TCC S Ser 240	720
GG# Gly	CTC	AGA	A TCT g Ser	CGA Arg 245	Glu	ATC Met	CTC	TCC Ser	CGT Arg 250	; Gl	TC	G TT	r CAG ∋ Hi:	C CG S Arg 25	A GAC g Asp 5	768
CT(Lev	C AGT	r GGK	G CTC y Let 260	ı Asp	GCA Ala	GAC Glu	ACO 1 Thi	C CTC r Leu 265	ı Let	Ly:	G GG S Gl	c cg y Ar	A GG g Gl 27	y Va	C CAC 1 His	816
GG' Gl	T AGO y Se:	2 TT r Ph 27	e Le	G GCT u Ala	CGG Arg	CCC Pro	28	r Arg	D AAG	G AAG	c CA n Gl	G GG n Gl; 28	y As	C TT p Ph	C TCG e Ser	864
CT(C TCC u Se: 29	r Va	C AG	G GTK	GG(GA' / Asj 29	o G1	G GT(n Va)	G ACC	C CA r Hi	T AT s Il 30	e Ar	G AI g Il	C CA e Gl	G AAC n Asn	912
TC Se 30	r Gl	G GA y As	T TT p Ph	C TA' e Ty:	r GAG r Ası 310	Le	д ТА u Ту	T GG r Gl	A GG y Gl	G GA y Gl 31	u Ly	AG TT /s Ph	T GC Le Al	G AC	T CTG ir Leu 320	
AC Th	A GA r Gl	G CT u L∈	G GT eu Va	G GA 1 G1 32	и Ту	TA r Ty	C AC r Th	T CA	G CA n Gl 33	n Gl	.G GC	GT GT ly Va	C C	eu G.	AG GAC ln Asp 35	1008
CG	C GA	.c ga	C AC	C AT	C AT	C CA	C CI	C AA	G TA	c cc	G C	rg A	AC TY	GC TY	CC GAT	1056

Arg	Asp	Gly	Thr 340	Ile	Ile	His	Leu	Lys 345	Tyr	Pro	Leu	Asn	Cys 350	Ser	Asp	
			GAG Glu													1104
			CTG Leu													1152
			AGC Ser													1200
			GCT Ala													1248
GTC Val	ATG Met	TGC Cys	GAG Glu 420	GGT Gly	GGA Gly	CGC Arg	TAC Tyr	ACA Thr 425	GTG Val	GGT Gly	GGT Gly	TTG Leu	GAG Glu 430	ACC Thr	TTC Phe	1296
			ACG Thr													1344
			GGC Gly									Tyr				1392
	Val		GCG Ala			Ile					Leu				AAG Lys 480	1440
					Asp					Gly					TTT Phe	1488
				Lys					Asn					, Lei	G GAA 1 Glu	1536
			Pro					/ Lys					a Ası		r CTC e Leu	1584
		Asp					Ile					g Asi			C ATC	1632
	Gly					e Asr					e Ly:				G CTA Leu 560	1680
GG(C CCT	GA?	r GAC o Glu	AAC Asr 565	n Ala	r aac a Lys	ACC Thi	TAC	C ATC	Ala	C AGG a Se	C CAG	g GG n Gl	C TG y Cy 57	T CTG s Leu 5	1728

GAG Glu	GCC Ala	ACG Thr	GTC Val 580	AAT Asn	GAC Asp	TTC Phe	TGG Trp	CAG Gln 585	ATG Met	GCG Ala	TGG Trp	CAG Gln	GAG Glu 590	AAC Asn	AGC Ser	1776
CGT Arg	GTC Val	ATC Ile 595	GTC Val	ATG Met	ACC Thr	ACC Thr	CGA Arg 600	GAG Glu	GTG Val	GAG Glu	AAA Lys	GGC Gly 605	CGG Arg	AAC Asn	AAA Lys	1824
TGC Cys	GTC Val 610	CCA Pro	TAC Tyr	TGG Trp	CCC Pro	GAG Glu 615	GTG Val	GGC Gly	ATG Met	CAG Gln	CGT Arg 620	GCT Ala	TAT Tyr	GGG Gly	CCC Pro	1872
TAC Tyr 625	TCT Ser	GTG Val	ACC Thr	AAC Asn	TGC Cys 630	GGG Gly	GAG Glu	CAT His	GAC Asp	ACA Thr 635	ACC Thr	GAA Glu	TAC Tyr	AAA Lys	CTC Leu 640	1920
CGT Arg	ACC Thr	TTA Leu	CAG Gln	GTC Val 645	TCC Ser	CCG Pro	CTG Leu	GAC Asp	AAT Asn 650	GGA Gly	GAC Asp	CTG Leu	ATT	CGG Arg 655	Glu	1968
ATC Ile	TGG Trp	CAT	TAC Tyr 660	CAG Gln	TAC Tyr	CTG Leu	AGC Ser	TGG Trp 665	Pro	GAC Asp	CAT His	GGG Gly	GTC Val 670	Pro	AGT Ser	2016
GAG Glu	CCT Pro	GGG Gly 675	Gly	GTC Val	CTC Leu	AGC Ser	TTC Phe 680	Leu	GAC Asp	CAG Gln	ATC Ile	AAC Asn 685	Gln	CGC Arg	CAG Gln	2064
GAA Glu	AGT Ser 690	Leu	CCT Pro	CAC His	GCA Ala	GGG Gly 695	CCC	: ATC	: ATC	GTG Val	CAC His	Cys	AGC Ser	GCC Ala	G GGC	2112
ATC 11e 705	e Gly	CGC Arg	ACA Thr	Gly	ACC Thr	Ile	ATT Ile	GTC Val	ATC Ile	GAC Asp 715	Met	CTC	ATC Met	GA(AAC Asn 720	2160
ATC Ile	TCC Ser	ACC Thi	AAC Lys	GGG G1 ₃ 725	/ Leu	GAC Asp	TGT Cys	GAC Asp	730	e Asp	TATY O Ile	C CAC	AAC 1 Lys	ACC Th	C ATC r Ile 5	2208
CA(Glr	TA E	GTC Val	l Arg	G GCC g Ala	a Glr	CGC Arg	Sei	c Gly	y Met	G GTY	G CAG	G ACC	G GA0 r G1: 75:	u Al	G CAG a Gln	2256
TAC Ty:	C AAG	5 TT0 5 Pho 75	e Ile	TAC	c GTC	GCC Ala	76	e Ala	C CAG	G TTO	e Il	T GA e Gl	u Th	C AC	T AAG r Lys	2304
AA(Ly:	G AA(s Ly: 77	s Le	G GA0	G GTV	CTX l Lev	G CAC J Glr 775	n Se	G CAG	G AA(n Ly:	G GG s Gl	C CA y G1 78	n Gl	G TC u Se	G GA r Gl	G TAC u Tyr	2352
GG G1 78	y As	C AT n Il	C AC	C TA	T CCC r Pro 79	o Pro	A GC o Al	C AT a Me	G AA t Ly	G AA s As 79	n Al	C CA a Hi	T GC s Al	C AF a Ly	G GCC s Ala 800	2400
TC	c cg	C AC	C TC	G TC	C AA	A CA	C AA	G GA	G GA	T GT	G TA	T GA	G Aª	C C	rg CAC	2448

Ser Arg Thr Ser Ser Lys His Lys Glu Asp Val Tyr Glu Asn Leu His 805 810 815

ACT AAG AAC AAG AGG GAG GAG AAA GTG AAG AAG CAG CGG TCA GCA GAC

Thr Lys Asn Lys Arg Glu Glu Lys Val Lys Lys Gln Arg Ser Ala Asp

820

825

830

AAG GAG AAG AGC AAG GGT TCC CTC AAG AGG AAG TGA Lys Glu Lys Ser Lys Gly Ser Leu Lys Arg Lys 835 840 2532

(2) INFORMATION FOR SEQ ID NO:117:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 843 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:117:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 20 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 45 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 60 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 75 70 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 90 85 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 105 100 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 120 115 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 135 140 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 155 150 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 170 165 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 185 190 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 205 200 Ser Lys Asp Prc Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 220 215 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 225 230 235 Gly Leu Arg Ser Arg Glu Met Leu Ser Arg Gly Trp Phe His Arg Asp 250 245 Leu Ser Gly Leu Asp Ala Glu Thr Leu Leu Lys Gly Arg Gly Val His

			260					265					270		
Gly	Ser	Phe 275	Leu	Ala	Arg	Pro	Ser 280	Arg	Lys	Asn	Gln	Gly 285	Asp	Phe	Ser
Leu	Ser 290	Val	Arg	Val	Gly	Asp 295	Gln	Val	Thr	His	Ile 300	Arg	Ile	Gln	Asn
305					310					315				Thr	320
				325					330					Gln 335	
			340					345					350	Ser	_
		355					360					365		Gln	
	370					375					380			Val	
385					390					395				Ser	400
				405					410					Ile 415	
			420					425					430	Thr	
		435					440					445		Ile	
	450					455					460			Ala	
465					470					475				Asn	480
				485					490					Glu 4 95	
			500					505					510	Leu	
		515					520					525		Ile	
	530					535					540			Asn	
545					550					555				Leu	560
				565					570					Cys 575	
			580					585					590	Asn	
		595					600					605		Asn	
	610					615					620			Gly	
625					630					635				Lys	640
				645					650					Arg 655	
			660					665					670	Pro	
		675					680					685		Arg	
	690					695					700			Ala	
Ile 705	Gly	Arg	Thr	Gly	Thr 710	Ile	Ile	Val	Ile	Asp 715	Met	Leu	Met	Glu	Asn 720
Ile	Ser	Thr	Lys	Gly	Leu	Asp	Cys	Asp	Ile	Asp	Ile	Gln	Lys	Thr	Ile

	7:	25			7	30					735		
Gln Met Val	Arg A:	la Gln		-	745					750			
Tyr Lys Phe 755			•	760				•	765				
Lys Lys Leu 770			775				7	780					
Gly Asn Ile 785		790				-	795					800	
Ser Arg Thr	8	05				B10					815		
Thr Lys Asn	820				825			Gln .	Arg	830	Ala	Asp	
Lys Glu Lys 835		ys Gly		Leu 840	Lys .	Arg 1	Lys						
(2) INFC	RMATIO	N FOR	SEQ	ID	NO:1	18:						
		E CHAR											
		H: 256 nucle			irs								
(C)	STRAN	IDEDNES	S: si	ngle	<u> </u>								
(D)	TOPOL	.OGY: 1	inear										
, ,	MOLECU FEATUR	LE TYP RE:	E: cI	ALIO									
		E/KEY:			equer	ice							
(1	B) LOCA	: NOITA	12	2559									
(!	O) OTH	ER INFO	TAMAC										
				ION:	: SEC) ID	NO:1	.18:					
(xi)	SEQUE	NCE DES	SCRIP:	ION:					ccc	· CTYC	GAT.	GC A	48
(xi) ATG CTG TCG Met Leu Se	SEQUEI	NCE DES	SCRIPT	ION: TION: CAC	CGA	GAC	CTC	AGT	GGG Gly	CTG Leu	GAT Asp 15	GCA Ala	48
(xi) ATG CTG TCG Met Leu Se	SEQUEI C CGT (r Arg (NCE DES GGG TGG Gly Trj 5	SCRIPT G TTT p Phe	ION: FION: CAC His	CGA Arg	GAC Asp 10	CTC Leu	AGT Ser	Gly	Leu	Asp 15	Ala	4 8 96
(xi) ATG CTG TCG Met Leu Se	SEQUEI C CGT (r Arg (NCE DES GGG TGG Gly Trj 5	SCRIPT G TTT p Phe	ION: ION: CAC His	CGA Arg GTC	GAC Asp 10 CAC	CTC Leu GGT	AGT Ser AGC	Gly	Leu CTC	Asp 15 GCT	Ala	
(xi) ATG CTG TCG Met Leu Se 1 GAG ACC CTG Glu Thr Le	SEQUEI C CGT (r Arg (G CTC :	NCE DES GGG TGG Gly Tr 5 AAG GGG Lys Gl:	SCRIPT G TTT D Phe C CGA Y Arg	CAC His GGT Gly	CGA Arg GTC Val 25	GAC Asp 10 CAC His	CTC Leu GGT Gly	AGT Ser AGC Ser	Gly TTC Phe	CTC Leu 30	Asp 15 GCT Ala	Ala CGG Arg	
(xi) ATG CTG TCG Met Leu Se. 1 GAG ACC CTG Glu Thr Le CCC AGT CG Pro Ser Ar	SEQUEI C CGT (Arg (C CTC) Leu : 20	NCE DES	SCRIPT G TTT D Phe C CGA y Arg	CAC His GGT Gly GAC Asp	CGA Arg GTC Val 25	GAC Asp 10 CAC His	CTC Leu GGT Gly	AGT Ser AGC Ser	Gly TTC Phe	CTG Leu 30	Asp 15 GCT Ala	Ala CGG Arg	96
(xi) ATG CTG TCC Met Leu Se 1 GAG ACC CTC Glu Thr Le CCC AGT CG Pro Ser Ar 35	SEQUEI C CGT (Arg (Leu : 20 C AAG : g Lys :	NCE DES GGG TGG G1y Try 5 AAG GGG Lys G1; AAC CA Asn G1	SCRIPT TO Phe C CGA y Arg G GGT n Gly	CAC His GGT Gly GAC Asp	CGA Arg GTC Val 25 TTC Phe	GAC Asp 10 CAC His	CTC Leu GGT Gly CTC Leu	AGT Ser AGC Ser TCC Ser	TTC Phe GTC Val 45	CTC Leu 30 AGC	Asp 15 GCT Ala GCT Val	Ala CGG Arg GGG Gly	96 144
(xi) ATG CTG TCG Met Leu Se. 1 GAG ACC CTG Glu Thr Le CCC AGT CG Pro Ser Ar	SEQUEI C CGT (Arg (Leu : 20 C AAG : g Lys :	NCE DES GGG TGG G1y Try 5 AAG GGG Lys G1; AAC CA Asn G1	SCRIPT TO Phe C CGA Y Arg G GGT n Gly T CGG	CAC His GGT Gly GAC Asp 40 ATC	CGA Arg GTC Val 25 TTC Phe	GAC Asp 10 CAC His TCG Ser	CTC Leu GGT Gly CTC Leu	AGT Ser AGC Ser TCC Ser	TTC Phe GTC Val 45	CTC Leu 30 AGC Arg	Asp 15 GCT Ala GGTC Val	CGG Arg GGG Gly	96
(xi) ATG CTG TCC Met Leu Se 1 GAG ACC CTC Glu Thr Le CCC AGT CG Pro Ser Ar 35 GAT CAG GT	SEQUEI C CGT (Arg (Leu : 20 C AAG : g Lys :	NCE DES GGG TGG G1y Try 5 AAG GGG Lys G1; AAC CA Asn G1	SCRIPT TO Phe C CGA Y Arg G GGT n Gly T CGG	CAC His GGT Gly GAC Asp 40 ATC	CGA Arg GTC Val 25 TTC Phe	GAC Asp 10 CAC His TCG Ser	CTC Leu GGT Gly CTC Leu	AGT Ser AGC Ser TCC Ser	TTC Phe GTC Val 45	CTC Leu 30 AGC Arg	Asp 15 GCT Ala GGTC Val	CGG Arg GGG Gly	96 144
(xi) ATG CTG TCC Met Leu Se 1 GAG ACC CTC Glu Thr Le CCC AGT CG Pro Ser Ar 35 GAT CAG GT Asp Gln Va 50 CTG TAT GG	SEQUEI C CGT (Arg (Leu : 20 C AAG : G Lys : G ACC 1 Thr	NCE DES GGG TGG G1y Try 5 AAG GGG Lys G1; AAC CA Asn G1 CAT AT His I1	G GGT CGG Arg 55	CAC His GGT Gly ASP 40 ATC Ile	CGA Arg GTC Val 25 TTC Phe CAG	GAC Asp 10 CAC His TCG Ser AAC Asn	CTC Leu GGT CTC Leu TCA Ser	AGT Ser AGC Ser TCC Ser GGG Gly 60	GTC Phe GTC Val 45 GAT Asi	CTC Leu 30 CAGC Arc	Asp 15 GCT Ala GCT Val	CGG Arg GGG GJy GAC Asp	96 144
(xi) ATG CTG TCC Met Leu Se 1 GAG ACC CTC Glu Thr Le CCC AGT CG Pro Ser Ar 35 GAT CAG GT Asp Gln Va 50	SEQUEI C CGT (Arg (Leu : 20 C AAG : G Lys : G ACC 1 Thr	NCE DES GGG TGG G1y Try 5 AAG GGG Lys G1; AAC CA Asn G1 CAT AT His I1	GCRIPT G TTT C Phe C CGA Y Arg G GGT n Gly T CGG E Arg 55 G TTT S Phe	CAC His GGT Gly ASP 40 ATC Ile	CGA Arg GTC Val 25 TTC Phe CAG	GAC Asp 10 CAC His TCG Ser AAC Asn	CTC Leu GGT CTC Leu TCA Ser	AGT Ser AGC Ser TCC Ser GGG Gly 60	GTC Phe GTC Val 45 GAT Asi	CTC Leu 30 CAGC Arc	Asp 15 GCT Ala GCT Val	CGG Arg GGG GJy GAC Asp	96 144 192
(xi) ATG CTG TCC Met Leu Se 1 GAG ACC CTC Glu Thr Le CCC AGT CG Pro Ser Ar 35 GAT CAG GT Asp Gln Va 50 CTG TAT GG Leu Tyr Gl 65	SEQUEI C CGT (Arg (Leu : 20 C AAG : G Lys : G ACC Thr A GGG y Gly	NCE DES	G GGT CGG Arg S5 Phe	CAC His GGT Gly GAC Asp 40 ATC Ile Ala	GGA Arg GTC Val 25 TTC Phe CAG Gln ACT Thr	GAC Asp 10 CAC His TCG Ser AAC Asn CTG Leu	CTC Leu GGT Gly CTC Leu TCA Ser ACA Thr 75	AGT Ser AGC Ser TCC Ser GGG Gly 60 GAG Glu	GTC Val. 45 Asp. CTC Let	C ACC	Asp 15 GCT Ala GCTA' Val Tyr	CGG Arg GGG GGY Asp GTAC TYr 80	96 144 192
(xi) ATG CTG TCC Met Leu Se 1 GAG ACC CTC Glu Thr Le CCC AGT CG Pro Ser Ar 35 GAT CAG GT Asp Gln Va 50 CTG TAT GG Leu Tyr Gl 65	SEQUEI C CGT (Arg (Leu : 20 C AAG : G Lys : G ACC Thr A GGG y Gly	NCE DES	G GGT CGG Arg S5 Phe	CAC His GGT Gly GAC Asp 40 ATC Ile Ala	GGA Arg GTC Val 25 TTC Phe CAG Gln ACT Thr	GAC Asp 10 CAC His TCG Ser AAC Asn CTG Leu	CTC Leu GGT Gly CTC Leu TCA Ser ACA Thr 75	AGT Ser AGC Ser TCC Ser GGG Gly 60 GAG Glu	GTC Val. 45 Asp. CTC Let	C ACC	Asp 15 GCT Ala GCTA' Val Tyr	CGG Arg GGG GGY Asp GTAC TYr 80	96 144 192 240

His	Leu	Lys	Tyr 100	Pro	Leu	Asn	Cys	Ser 105	Asp	Pro	Thr	Ser	Glu 110	Arg	Trp	
TAC Tyr	CAT His	GGC Gly 115	CAC His	ATG Met	TCT Ser	GGC	GGG Gly 120	CAG Gln	GCA Ala	GAG Glu	ACG Thr	CTG Leu 125	CTG Leu	CAG Gln	GCC Ala	384
AAG Lys	GGC Gly 130	GAG Glu	CCC Pro	TGG Trp	ACG Thr	TTT Phe 135	CTT Leu	GTG Val	CGT Arg	GAG Glu	AGC Ser 140	CTC Leu	AGC Ser	CAG Gln	CCT Pro	432
GGA Gly 145	GAC Asp	TTC Phe	GTG Val	CTT Leu	TCT Ser 150	GTG Val	CTC Leu	AGT Ser	GAC Asp	CAG Gln 155	CCC Pro	AAG Lys	GCT Ala	GGC Gly	CCA Pro 160	480
GGC Gly	TCC Ser	CCG Pro	CTC Leu	AGG Arg 165	GTC Val	ACC Thr	CAC His	ATC Ile	AAG Lys 170	GTC Val	ATG Met	TGC Cys	GAG Glu	GGT Gly 175	GGA Gly	528
CGC Arg	TAC Tyr	ACA Thr	GTG Val 180	GGT Gly	GGT Gly	TTG Leu	GAG Glu	ACC Thr 185	TTC Phe	GAC Asp	AGC Ser	CTC Leu	ACG Thr 190	GAC Asp	CTG Leu	576
GTA Val	GAG Glu	CAT His	Phe	AAG Lys	AAG Lys	ACG Thr	GGG Gly 200	ATT	GAG Glu	GAG Glu	GCC Ala	TCA Ser 205	Gly	GCC Ala	TTT Phe	624
GTC Val	TAC Tyr 210	Leu	CGG Arg	CAG Gln	CCG Pro	TAC Tyr 215	ТАТ Туг	GCC Ala	ACG Thr	AGG Arg	GTG Val 220	Asn	GCG Ala	GCT Ala	GAC Asp	672
ATT 11e 225	e Glu	AAC Asn	CGA Arg	GTC Val	TTG Leu 230	Glu	CTC Leu	AAC Asn	AAG Lys	AAC Lys 235	Glr	GAC Glu	TCC Ser	GAC Glu	GAT Asp 240	720
AC <i>A</i> Thr	A GCC	: AAC	GCT Ala	GGC Gly 245	/ Phe	TGG Trp	GAC Glu	GAG ıGlu	TTT Phe 250	Glu	G AGT	TTC Lev	G CAC	AAC Lys 255	G CAG Gln	768
GA(G GTC	AAC Lys	AAC Asn 260	Leu	G CAC	CAG Gln	CG?	r CTG g Lev 265	Glu	GG(G CAC	G CG(G CCA g Pro 270	o Glu	AAC Asn	816
AA(Lys	G GG(B Gl)	275 275	s Asr	CGC Arg	ТАС Туг	: AAG Lys	AA(ASI 28(n Ile	CTC	CCC Pro	TT'	T GAG e Ası 28!	p Hi	C AGG S Sen	C CGA r Arg	864
GT(Va)	G ATS	e Lei	G CAC	G GGZ	A CGC	GAC Asp 295	se:	r AAC r Asr	TATO	C CC	G GGG G G1; 30	y Se	C GA	С ТАО р Ту	C ATC	912
AA As:	n Al	C AAG a Asi	TAC	C ATY	C AAC e Lys 310	s Ası	C CAC	G CT(n Lev	CT/	A GG u Gl: 31	y Pr	T GA o As	T GA p Gl	G AA u As	C GCT n Ala 320	960
AA Ly	G AC	C TAI	C ATO	C GC e Al 32	a Se	CAC	G GG n Gl	C TG' y Cy:	r CTG s Leg	u Gl	G GC u Al	C AC a Th	G GT r Va	C AA 1 As 33	T GAC n Asp 5	1008

TTC Phe	TGG Trp	CAG Gln	ATG Met 340	GCG Ala	TGG Trp	CAG Gln	GAG Glu	AAC Asn 345	AGC Ser	CGT Arg	GTC Val	ATC Ile	GTC Val 350	ATG Met	ACC Thr	1056
ACC Thr	CGA Arg	GAG Glu 355	GTG Val	GAG Glu	AAA Lys	GGC Gly	CGG Arg 360	AAC Asn	AAA Lys	TGC Cys	GTC Val	CCA Pro 365	TAC Tyr	TGG Trp	CCC Pro	1104
GAG Glu	GTG Val 370	GGC Gly	ATG Met	CAG Gln	CGT Arg	GCT Ala 375	TAT Tyr	GGG Gly	CCC Pro	TAC Tyr	TCT Ser 380	GTG Val	ACC Thr	AAC Asn	TGC Cys	1152
GGG Gly 385	GAG Glu	CAT His	GAC Asp	ACA Thr	ACC Thr 390	GAA Glu	TAC Tyr	AAA Lys	CTC Leu	CGT Arg 395	ACC Thr	TTA Leu	CAG Gln	GTC Val	TCC Ser 400	1200
CCG Pro	CTG Leu	GAC Asp	AAT Asn	GGA Gly 405	GAC Asp	CTG Leu	ATT Ile	CGG Arg	GAG Glu 410	ATC Ile	TGG Trp	CAT His	TAC Tyr	CAG Gln 415	TAC Tyr	1248
CTG Leu	AGC Ser	TGG Trp	CCC Pro 420	GAC Asp	CAT His	GGG Gly	GTC Val	CCC Pro 425	AGT Ser	GAG Glu	CCT Pro	GGG Gly	GGT Gly 430	Val	Leu	1296
AGC Ser	TTC Phe	CTG Leu 435	Asp	CAG Gln	ATC Ile	AAC Asn	CAG Gln 440	Arg	CAG Gln	GAA Glu	AGT Ser	CTG Leu 445	CCT	CAC His	GCA Ala	1344
GGG Gly	CCC Pro 450	Ile	: ATC	GTG Val	CAC His	TGC Cys 455	AGC Ser	GCC Ala	GGC Gly	ATC Ile	GGC Gly 460	Arg	ACA Thr	GG(ACC Thr	1392
ATC Ile 465	Ile	GTC Val	: ATC	GAC Asp	Met 470	Leu	ATC Met	GAG Glu	AAC Asn	11e	Ser	ACC Thr	Lys	G GGG G Gly	CTG Leu 480	1440
GAC Asp	TGT Cys	GAC	ATT Ile	GAC Asp 485	Ile	CAG Gln	AAC Lys	ACC Thr	11e	Glr	ATC Met	GTC Val	CGC L Arg	g GCc g Al 49	G CAG a Gln 5	1488
CGC Arg	TCG Ser	GG(ATC Met 500	. Val	G CAC	ACG Thr	GAC Glu	ı Ala	G CAC Glr	тул	Lys	TTC Phe	C ATG	е Ту	C GTG r Val	1536
GCC Ala	ATC	GCC Ala 515	a Glr	TTC	C ATT	GAÆ Glu	A ACC 1 Th: 520	r Thr	AAC Lys	AA(ELY:	G AAG s Ly:	G CTO s Lev 52!	ı Gl	G GT u Va	C CTG l Leu	1584
CAC Glr	TCC Ser 530	Gli	g AAC n Lys	GGGG Gly	CAC Gli	G GAC n Glu 535	ı Se	G GAC r Glu	ТА (C GGG r Gl	G AA y As 54	n Il	C AC e Th	С ТА r Ту	T CCC T Pro	1632
CCA Pro 545	o Ala	TATY Me	G AAG t Ly:	AA S ASI	r GCC n Ala 55	a Hi	r GC s Al	C AA(a Ly:	G GCG S Ala	C TC a Se 55	r Ar	C AC g Th	C TC r Se	G TC	C AAA r Lys 560	1680
CAG	C AAC	G GA	G GA	r GT	G TA	T GA	g aa	CT	G CA	C AC	T AA	G AA	C AA	G AC	G GAG	1728

His	Lys	Glu	Asp	Val 565	Tyr	Glu	Asn	Leu	His 570	Thr	Lys	Asn	Lys	Arg 575	Glu	
GAG Glu	AAA Lys	GTG Val	AAG Lys 580	AAG Lys	CAG Gln	CGG Arg	TCA Ser	GCA Ala 585	GAC Asp	AAG Lys	GAG Glu	AAG Lys	AGC Ser 590	AAG Lys	GGT Gly	1776
TCC Ser	CTC Leu	AAG Lys 595	AGG Arg	AAG Lys	CGA Arg	ATT Ile	CTG Leu 600	CAG Gln	TCG Ser	ACG Thr	GTA Val	CCG Pro 605	CGG Arg	GCC Ala	CGG Arg	1824
GAT Asp	CCA Pro 610	CCG Pro	GTC Val	GCC Ala	ACC Thr	ATG Met 615	GTG Val	AGC Ser	AAG Lys	GGC Gly	GAG Glu 620	GAG Glu	CTG Leu	TTC Phe	ACC Thr	1872
GGG Gly 625	GTG Val	GTG Val	CCC Pro	ATC Ile	CTG Leu 630	GTC Val	GAG Glu	CTG Leu	GAC Asp	GGC Gly 635	GAC Asp	GTA Val	AAC Asn	GGC Gly	CAC His 640	1920
AAG Lys	TTC Phe	AGC Ser	GTG Val	TCC Ser 645	GGC Gly	GAG Glu	GGC Gly	GAG Glu	GGC Gly 650	GAT Asp	GCC Ala	ACC Thr	TAC Tyr	GGC Gly 655	AAG Lys	1968
CTG Leu	ACC Thr	CTG Leu	AAG Lys 660	TTC Phe	ATC Ile	TGC Cys	ACC Thr	ACC Thr 665	GGC Gly	AAG Lys	CTG Leu	CCC Pro	GTG Val 670	Pro	TGG Trp	2016
CCC	ACC Thr	CTC Leu 675	Val	ACC Thr	ACC Thr	CTG Leu	ACC Thr 680		GGC Gly	GTG Val	CAG Gln	TGC Cys 685	Phe	AGC Ser	CGC Arg	2064
TAC Tyr	CCC Pro 690	Asp	CAC His	ATG Met	AAG Lys	CAG Gln 695	His	GAC Asp	TTC Phe	TTC	AAG Lys 700	Ser	GCC Ala	: ATC	CCC Pro	2112
GA2 G1u 705	Gly	ТАС	GTC Val	CAG Gln	GAG Glu 710	Arg	ACC	T ATC	TTC Phe	TTC Phe 715	Lys	GAC Asp	GAC Asp	GG(AAC Asn 720	2160
ТАС Туг	AAC Lys	ACC Thr	CGC Arg	GCC Ala 725	Glu	GTG Val	AAC Lys	TTC Phe	GAG Glu 730	Gly	GAC Asp	ACC Thr	CTC Lev	GTX 1 Va: 73!	AAC Asn	2208
CG(Arg	C ATO	GAC Glu	CTC Lev 740	ı Lys	GGC Gly	ATC	GAC Asi	745	E Lys	GAC	GAC 1 Asp	GGG Gly	C AAC 3' Asi 750	n Ile	CTG e Leu	2256
GG(Gly	G CAC / His	2 AA0 3 Lys 75!	s Lev	G GAC	TAC 1 Tyr	AAC Asr	ТАО 1 Ту: 760	r Ası	C AGO	C CAG	C AAG	C GTO n Val	1 Ty:	r AT	C ATG e Met	2304
GC(C GAG a Ası 770) Ly:	G CAC	3 AAC 1 Lys	G AAC s Asr	775	/ Il	C AAG e Ly:	G GTY	G AA l As	n Pho	e Ly	G ATV s Il	C CG e Ar	C CAC g His	2352
AA As:	n Il	C GAG	G GA(u Asj	C GGG C Gl	C AGG y Sei 790	r Val	G CA	G CTO	C GCC	C GA a As 79	p Hi	C TA s Ty	C CA r Gl	G CA n Gl	G AAC n Asn 800	2400

							TAC Tyr 815	2448
							GAT Asp	2496
							GGC Gly	2544
 GAG Glu 850		TAA						2562

(2) INFORMATION FOR SEQ ID NO:119:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 853 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
 (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:119:

Met 1	Leu	Ser	Arg	Gly 5	Trp	Phe	His	Arg	Asp 10	Leu	Ser	Gly	Leu	Asp 15	Ala
Glu	Thr	Leu	Leu 20	Lys	Gly	Arg	Gly	Val 25	His	Gly	Ser	Phe	Leu 30	Ala	Arg
Pro	Ser	Arg 35	Lys	Asn	Gln	Gly	Asp 40	Phe	Ser	Leu	Ser	Val 45	Arg	Val	Gly
_	50					55					60			Tyr	
Leu 65	Tyr	Gly	Gly	Glu	Lys 70	Phe	Ala	Thr	Leu	Thr 75	Glu	Leu	Val	Glu	Tyr 80
Tyr	Thr	Gln	Gln	Gln 85	Gly	Val	Leu	Gln	Asp 90	Arg	Asp	Gly	Thr	Ile 95	Ile
His	Leu	Lys	Tyr 100	Pro	Leu	Asn	Суѕ	Ser 105	Asp	Pro	Thr	Ser	Glu 110	Arg	Trp
Tyr	His	Gly 115	His	Met	Ser	Gly	Gly 120	Gln	Ala	Glu	Thr	Leu 125	Leu	Gln	Ala
Lys	Gly 130	Glu	Pro	Trp	Thr	Phe 135	Leu	Val	Arg	Glu	Ser 140	Leu	Ser	Gln	Pro
Gly 145	Asp	Phe	Val	Leu	Ser 150	Val	Leu	Ser	Asp	Gln 155	Pro	Lys	Ala	Gly	Pro 160
	Ser	Pro	Leu	Arg 165	Val	Thr	His	Ile	Lys 170	Val	Met	Суѕ	Glu	Gly 175	Gly
Arg	Tyr	Thr	Val 180	Gly	Gly	Leu	Glu	Thr 185	Phe	Asp	Ser	Leu	Thr 190		Leu
Val	Glu	His 195	Phe	Lys	Lys	Thr	Gly 200		Glu	Glu	Ala	Ser 205	Gly	Ala	Phe
Val	Tyr		Arg	Gln	Pro	Tyr	Tyr	Ala	Thr	Arg	Val	Asn	Ala	Ala	Asp

2	10					215					220				
Ile G	ilu A	Asn	Arg		Leu (230	Glu	Leu	Asn	Lys	Lys 235	Gln	Glu	Ser	Glu	Asp 240
Thr A	Ala I	Lys	Ala			Trp	Glu	Glu	Phe 250	Glu	Ser	Leu	Gln	Lys 255	Gln
Glu V	/al 1	Lys	Asn 260		His	Gln	Arg	Leu 265	Glu	Gly	Gln	Arg	Pro 270	Glu	Asn
Lys (Lys 275	Asn	Arg	Tyr	Lys	Asn 280	Ile	Leu	Pro	Phe	Asp 285	His	Ser	Arg
Val 1	Ile :	Leu	Gln	Gly	Arg	Asp 295	Ser	Asn	Ile	Pro	Gly 300	Ser	Asp	Tyr	Ile
Asn 2 305	Ala	Asn	Tyr	Ile	Lys 310	Asn	Gln	Leu	Leu	Gly 315	Pro	Asp	Glu	Asn	Ala 320
Lys :	Thr	Tyr	Ile	Ala 325		Gln	Gly	Cys	Leu 330	Glu	Ala	Thr	Val	Asn 335	Asp
Phe '	Trp	Gln	Met 340	Ala	Trp	Gln	Glu	Asn 345	Ser	Arg	Val	Ile	Val 350	Met	Thr
Thr	Arg	Glu 355	Val	Glu	Lys	Gly	Arg 360	Asn	Lys	Cys	Val	Pro 365	Tyr	Trp	Pro
Glu	370	Gly				375					380				
Gly :	Glu				390					395					400
Pro	Leu	Asp	Asn	Gly 405	Asp	Leu	Ile	Arg	Glu 410	Ile	Trp	His	Tyr	Gln 415	Tyr
			420	Asp				425					430		Leu
		435	Asp	Gln			440					445			Ala
	450	Ile	Ile			455					460				Thr
465	Ile				470					475	5				Leu 480
Asp				485					490)				495	
			500)				505	5				510)	Val
		515	ò				520)				525)		l Leu
	530					535	5				540)			r Pro
545					550	٢				55	5				r Lys 560
His				565	5				57	0				5/	
			580)				58	5				59	U	s Gly
		599	5				600	0				60)		a Arg
	610)				61	5				62	0			e Thr
625	Val	l Va			630)				63	5				y His 640
Lys	Phe			64	5				65	0				65	
			66	0				66	5				67	0	o Trp
Pro	Thi	r Le	u Va	1 Th	r Th	r Le	u Th	r Ty	r Gl	y Vā	il Gl	n Cy	s Ph	ne Se	er Arg

		675					680					685				
Tyr	Pro 690	Asp	His	Met	Lys	Gln 695	His	qzA	Phe	Phe	Lys 700	Ser	Ala	Met	Pro	
Glu 705		Tyr	Val	Gln	Glu 710	Arg	Thr	Ile	Phe	Phe 715	Lys	Asp	Asp	Gly	Asn 720	
Tyr	Lys	Thr	Arg	Ala 725	Glu	Val	Lys	Phe	Glu 730	Gly	Asp	Thr	Leu	Val 735	Asn	
Arg	Ile	Glu	Leu 740	Lys	Gly	Ile	Asp	Phe 745	Lys	Glu	Asp	Gly	Asn 750	Ile	Leu	
Gly	His	Lys 755	Leu	Glu	Tyr	Asn	Tyr 760	Asn	Ser	His	Asn	Val 765	Tyr	Ile	Met	
Ala	Asp 770	Lys	Gln	Lys	Asn	Gly 775	Ile	Lys	Val	Asn	Phe 780	Lys	Ile	Arg	His	
785					Ser 790					795					800	
				805	Gly				810					815		
			820		Leu			825					830			
Met	Val	Leu 835		Glu	Phe	Val	Thr 840		Ala	Gly	Ile	Thr 845		Gly	Met	
Asp	Glu 850		Tyr	Lys												
	((D) (ii) (ix) (F	TOF MOLE FEAT A) NF B) LO D) OT	CULE URE: ME/K CATI	EY: ON: INFO	inea E: c Codi 1	DNA ng S 2991	eđne			100					
					E DES											40
ATO Med 1	G GTY	G AGO	C AAC C Lys	GGG Gly 5	GAC Glu	G GAC	CTC	J TTC 1 Phe	Thi	GGC Gl	g GTC y Vai	G GTV l Va:	CCC Pr	2 ATC 5 Ile 15	c CTG e Leu	48
GTY Va	C GAG	G CTG	G GAG L Asi 20	c GG(c Gl ₂	GAC Asi	C GTA	A AAG L Asi	GGC n Gly 25	CAC His	C AAG s Ly:	G TTO	C AGG e Se:	C GTV r Va 30	G TC	C GGC r Gly	96
GA G1	G GGG	C GAG y Gli 35	G GG(u Gly	C GA' Y Ası	r GCC o Ala	a Thi	TAC TY:	c GG(r Gly	Z AAG y Ly:	G CT	G AC	C CTV r Le ² 45	G AA u Ly	G TT s Ph	C ATC e Ile	144
TG Cy	C AC s Th	r Th	C GGG r Gl	C AAG y Ly:	G CTY s Le	G CCG u Pro 55	C GT O Va	G CC	C TG	G CC p Pr	C AC o Th 60	r Le	C GT u Va	G AC 1 Th	C ACC	192

CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG 240

Leu 65	Thr	Tyr	Gly	Val	Gln 70	Cys	Phe	Ser	Arg	Tyr 75	Pro	Asp	His	Met	Lys 80	
							GCC Ala				_		_			288
							GAC Asp	_				_				336
							CTG Leu 120									384
				_		_	AAC Asn	_		_				_		432
				· .			TAT Tyr						_			480
							ATC Ile									528
							CAG Gln									576
							CAC His 200					_		_		624
							CGC Arg									672
							CTC Leu									720
							GCT Ala									768
							GGC Gly				_			Glu		816
							AAC Asn 280								GAA Glu	864
							AAG Lys								ACC Thr	912

AAA Lys 305	AAC Asn	AGA Arg	GAA Glu	Arg	TGG Trp 310	TGC Cys	CAT His	GAA Glu	Ile	CAG Gln 315	ATT Ile	ATG Met	AAG Lys	AAG Lys	TTG Leu 320	960
AAC Asn	CAT His	GCC Ala	AAT Asn	GTT Val 325	GTA Val	AAG Lys	GCC Ala	TGT Cys	GAT Asp 330	GTT Val	CCT Pro	GAA Glu	GAA Glu	TTG Leu 335	AAT Asn	1008
ATT Ile	TTG Leu	ATT Ile	CAT His 340	GAT Asp	GTG Val	CCT Pro	CTT Leu	CTA Leu 345	GCA Ala	ATG Met	GAA Glu	TAC Tyr	TGT Cys 350	TCT Ser	GGA Gly	1056
GGA Gly	GAT Asp	CTC Leu 355	CGA Arg	AAG Lys	CTG Leu	CTC Leu	AAC Asn 360	AAA Lys	CCA Pro	GAA Glu	AAT Asn	TGT Cys 365	TGT Cys	GGA Gly	CTT Leu	1104
AAA Lys	GAA Glu 370	AGC Ser	CAG Gln	ATA Ile	CTT Leu	TCT Ser 375	TTA Leu	CTA Leu	AGT Ser	GAT Asp	ATA Ile 380	GGG Gly	TCT Ser	GGG	ATT Ile	1152
CGA Arg 385	Tyr	TTG Leu	CAT His	GAA Glu	AAC Asn 390	AAA Lys	ATT Ile	ATA Ile	CAT His	CGA Arg 395	GAT Asp	CTA Leu	AAA Lys	CCT Pro	GAA Glu 400	1200
AAC Asn	ATA Ile	GTT Val	CTT Leu	CAG Gln 405	GAT Asp	GTT Val	GGT Gly	GGA Gly	AAG Lys 410	ATA Ile	ATA Ile	CAT His	AAA Lys	ATA Ile 415	Ile	1248
GAT Asp	CTG Leu	GGA Gly	TAT Tyr 420	Ala	AAA Lys	GAT Asp	GTT Val	GAT Asp 425	Gln	GGA Gly	AGT Ser	CTG	TGT Cys 430	Thr	TCT Ser	1296
TTI Phe	GTG Val	GGA Gly 435	Thr	CTG	CAG Gln	ТАТ Туг	CTC Lev 440	ı Ala	CCA Pro	GAG Glu	CTC Lev	TTT Phe 445	e Glu	AA7 Asr	AAG Lys	1344
CCT Pro	TAC Tyr 450	Thi	A GCC	ACT Thr	GTT Val	GAT Asp 455	ТУз	r TGG r Trp	AGC Ser	TTT Phe	GGC G1 ₂ 460	Thi	ATC Me	G GTA	A TTT l Phe	1392
GAF Glu 469	2 Cys	TAT:	r GCT e Ala	r GGA a Gly	TAT Ty:	Arg	CC!	r TTI o Phe	TTC	CAT His 475	Hi:	r CT(s Le:	G CAG	g CC n Pro	A TTT o Phe 480	1440
ACC Thi	TGC Trp	G CA'	r GAC s Glv	3 AAC 1 Lys 485	3 Ile	r AAC e Lys	AAG Ly:	G AAC s Lys	G GAT S Asi 490	Pro	A AAG	G TG' s Cy:	T AT s Il	A TT e Ph 49	T GCA e Ala 5	1488
CĀ:	T GA S Glu	A GA	G ATO u Met 500	t Ser	A GG r G1	A GAA	A GT ı Va	T CGC 1 Arg 505	g Phe	T AG' e Se:	r AG r Se	C CA r Hi	T TT s Le 51	u Pr	T CAA o Gln	1536
CC. Pr	A AA' o Asi	r AG n Se 51	r Le	T TGʻ u Cy:	T AG' s Se	T TT/	A AT 1 11 52	e Va	A GAI	A CC	C AT o Me	G GA t Gl 52	u As	C TG	G CTA	1584
CA	G TT	G AT	G TT	g AA	T TG	G GA	c cc	T CA	G CA	g ag	A GG	A GO	A CC	T GI	T GAC	1632

Gln	Leu 530	Met	Leu	Asn		Asp 535	Pro	Gln	Gln	Arg	Gly 540	Gly	Pro	Val	Asp	
											ATG Met					1680
											TCT Ser					1728
TCT Ser	TTT Phe	CTG Leu	TTA Leu 580	CCA Pro	CCT Pro	GAT Asp	GAA Glu	AGT Ser 585	CTT Leu	CAT His	TCA Ser	CTA Leu	CAG Gln 590	TCT Ser	CGT Arg	1776
											CAA Gln					1824
											GCC Ala 620					1872
	Asp										GTT Val					1920
											TCC Ser					1968
				Tyr					Ser		ATA Ile			Pro		2016
ATA Ile	CAG Gln	CTG Leu 675	Arg	AAA Lys	GTG Val	TGG Trp	GCT Ala 680	Glu	GCA Ala	GTC Val	CAC His	ТАТ Тут 685	Val	TCT Ser	GGA Gly	2064
		Glu					Leu					Arg			ATG Met	2112
TTA Let 705	ı Ser	CTI Leu	CTI Lev	'AGA Arg	ТАТ Туг 710	Asn	GCI Ala	AAC Asr	TTA Leu	ACA Thi 715	Lys	ATC Met	AAC Lys	AAC a Asr	ACT Thr 720	2160
TT(Let	OTA E	TCA Ser	A GCA c Ala	TCA Ser 725	Glr	CAA Gln	CTC	S AAA 1 Lys	A GCT s Ala 730	Ly	A TTC	GAC 1 Glu	TT.	TTT Phe 735	CAC His	2208
				ı Lev					y Tyr					t Thi	TAT Tyr	2256
			r Sei					ı Ly:					ı Me		A GAA u Glu	2304

AAG (Lys .	GCC Ala 770	ATC Ile	CAC His	TAT (Ala (GAG (Glu '	GTT Val	GGT Gly	GTC . Val	Ile	GGA G1y 780	TAC Tyr	CTG Leu	GAG Glu	GAT Asp	2352
CAG Gln 785	ATT Ile	ATG Met	TCT Ser	Leu	CAT (His .	GCT (GAA Glu	ATC Ile	Met	GGG Gly 795	CTA Leu	CAG Gln	AAG Lys	AGC Ser	CCC Pro 800	2400
TAT Tyr	GGA Gly	AGA Arg	CGT Arg	CAG Gln 805	GGA Gly	GAC Asp	TTG Leu	ATG Met	GAA Glu 810	TCT Ser	CTG Leu	GAA Glu	CAG Gln	CGT Arg 815	GCC Ala	2448
ATT Ile	GAT Asp	CTA Leu	тат туг 820	AAG Lys	CAG Gln	TTA Leu	AAA Lys	CAC His 825	AGA Arg	CCT Pro	TCA Ser	GAT Asp	CAC His 830	TCC Ser	TAC Tyr	2496
AGT Ser	GAC Asp	AGC Ser 835	ACA Thr	GAG Glu	ATG Met	GTG Val	AAA Lys 840	ATC Ile	ATT Ile	GTG Val	CAC His	ACT Thr 845	GTG Val	CAG Gln	AGT Ser	2544
CAG Gln	GAC Asp 850	CGT Arg	GTG Val	CTC Leu	AAG Lys	GAG Glu 855	CTG Leu	TTT Phe	GGT Gly	CAT His	TTG Leu 860	Ser	AAG Lys	TTG Leu	TTG Leu	2592
GGC Gly 865	Cys	AAG Lys	CAG Gln	AAG Lys	ATT Ile 870	ATT Ile	GAT Asp	CTA Leu	CTC Leu	CCT Pro 875	Lys	GTG Val	GAA Glu	GTC Val	GCC Ala 880	2640
CTC Leu	AGT Ser	AAT Asr	T ATC	AAA Lys 885	Glu	GCT Ala	GAC Asp	AAT Asn	ACT Thr 890	Val	ATG Met	TTC Phe	ATC Met	Glr 895	G GGA n Gly	2688
AAA Lys	AGG Arg	Glr	AAA Lys	Glu	ATA Ile	TGG Trp	CAT His	CTC Lev	Lev	`AAA	ATT	GCC Ala	TG1 Cys 910	Th	A CAG r Gln	2736
AGT Ser	TCT Ser	GC0 Ala	a Arc	TCT Ser	CTT	GTA Val	GG/ G1 ₃ 920	/ Ser	AGI Ser	CTA Lev	A GAA	A GG' 1 Gl; 92	y Ala	A GT. a Va	A ACC l Thr	2784
CCT Pro	CAC Glr 930	ı Th	A TCA	A GCA	A TGG	935	ı Pro	C CCC	G ACI	r TCA	A GCA c Ala 941	a Gl	A CA' u Hi:	r GA s As	T CAT p His	2832
TC: Sei	r Le	G TC. u Se	A TG:	r GTC s Val	G GT# 1 Va] 950	Thi	CC'	T CAI	A GAT	r GG(p Gl; 95	y Gl	G AC u Th	T TC. r Se	A GC r Al	A CAA a Gln 960	2880
ATC Met	G AT	A GA e Gl	A GAI u Gli	A AA' u Asi 96!	n Lei	AAC 1 Asi	TG n Cy	C CT	T GG u Gl; 97	y Hi	T TT s Le	A AG u Se	C AC	T AT r Il 97	T ATT e Ile	2928
CA' Hi	T GA s Gl	G GC u Al	A AA a As 98	n Gl	G GAA	A CAG J Gli	G GG n Gl	С АА у Аs 98	n Se	T AT r Me	G AT t Me	G AA et As	T CT in Le	u As	AT TGG Sp Trp	2976
AG	T TG	G TI	'A AC	A GA	A TG	A										2994

Ser Trp Leu Thr Glu 995

(2) INFORMATION FOR SEQ ID NO:121:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 997 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:121:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 20 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 60 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 70 75 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 90 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 120 125 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 135 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 150 155 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 185 190 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 200 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 215 220 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 235 Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Thr Met Glu Arg Pro 245 250 Pro Gly Leu Arg Pro Gly Ala Gly Gly Pro Trp Glu Met Arg Glu Arg 260 265 270 Leu Gly Thr Gly Gly Phe Gly Asn Val Cys Leu Tyr Gln His Arg Glu 280 Leu Asp Leu Lys Ile Ala Ile Lys Ser Cys Arg Leu Glu Leu Ser Thr 295 300 Lys Asn Arg Glu Arg Trp Cys His Glu Ile Gln Ile Met Lys Lys Leu 310 315 Asn His Ala Asn Val Val Lys Ala Cys Asp Val Pro Glu Glu Leu Asn

				325					330					335	
			His 340					345					350		
		355	Arg				360					365			
	370		Gln			375					380				
385			His		390					395					400
Asn	Ile	Val	Leu	Gln 405	Asp	Val	Gly	Gly	Lys 410	Ile	Ile	His	Lys	Ile 415	Ile
Asp	Leu	Gly	Туг 420	Ala	Lys	Asp	Val	Asp 425	Gln	Gly	Ser	Leu	Cys 430	Thr	Ser
Phe	Val	Gly 435	Thr	Leu	Gln	Tyr	Leu 440	Ala	Pro	Glu	Leu	Phe 445	Glu	Asn	Lys
Pro	Tyr 450	Thr	Ala	Thr	Val	Asp 455	Tyr	Trp	Ser	Phe	Gly 460	Thr	Met	Val	Phe
465			Ala		470					475					480
			Glu	485					490					495	
			Met 500					505					510		
		515					520					525			
	530		Leu			535					540				
545			Lys		550					555					560
				565	,				570					575	
			580					585					590)	Arg
		595	,				600					605			Ser
	610					615					620	•			: Val
625					630)				635					Asp 640
_				645	5				650)				655	
			660)				665					670)	lle
		675	5				680)				685	5		Gly
	690)				695	5				700)			a Met
705	5				710)				715	5				720
				725	5				730)				73	
			740)				745	5				75	0	r Tyr
		75	5				760)				76	5		u Glu
	770)				775	5				78	0			u Asp
Gli	n Ile	е Ме	t Se	r Le	u His	s Ala	a Glu	ı Ile	e Me	t Gl	y Le	u Gl:	n Ly	s Se	r Pro

785					790					795					800
Tyr	Gly	Arg	Arg	Gln 805	Gly	Asp	Leu	Met	Glu 810	Ser	Leu	Glu	Gln	Arg 815	Ala
Ile	Asp	Leu	Tyr 820	Lys	Gln	Leu	Lys	His 825	Arg	Pro	Ser	Asp	His 830	Ser	Tyr
		835				Val	840					845			
Gln	Asp 850	Arg	Val	Leu	Lys	Glu 855	Leu	Phe	Gly	His	Leu 860	Ser	Lys	Leu	Leu
Gly 865	Суѕ	Lys	Gln	Lys	Ile 870	Ile	Asp	Leu	Leu	Pro 875	Lys	Val	Glu	Val	Ala 880
Leu	Ser	Asn	Ile	Lys 885	Glu	Ala	Asp	Asn	Thr 890	Val	Met	Phe	Met	Gln 895	Gly
Lys	Arg	Gln	Lys 900	Glu	Ile	Trp	His	Leu 905	Leu	Lys	Ile	Ala	Cys 910	Thr	Gln
Ser	Ser	Ala 915	Arg	Ser	Leu	Val	Gly 920	Ser	Ser	Leu	Glu	Gly 925	Ala	Val	Thr
Pro	Gln 930	Thr	Ser	Ala	Trp	Leu 935	Pro	Pro	Thr	Ser	Ala 940	Glu	His	Asp	His
Ser 945	Leu	Ser	Cys	Val	Val 950	Thr	Pro	Gln	Asp	Gly 955	Glu	Thr	Ser	Ala	Gln 960
Met	Ile	Glu	Glu	Asn 965	Leu	Asn	Cys	Leu	Gly 970	His	Leu	Ser	Thr	Ile 975	Ile
His	Glu	Ala	Asn 980	Glu	Glu	Gln	Gly	Asn 985	Ser	Met	Met	Asn	Leu 990	Asp	Trp
Ser	Trp	Leu 995	Thr	Glu											

(2) INFORMATION FOR SEQ ID NO:122:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2991 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...2988
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:122:

	GAG Glu															48
	CGG Arg															96
	CAT His															144
GAG	CTA	AGT	ACC	AAA	AAC	AGA	GAA	CGA	TGG	TGC	САТ	GAA	ATC	CAG	ATT	192

Glu	Leu 50	Ser	Thr	Lys		Arg (55	Glu	Arg	Trp (Cys	His 60	Glu	Ile	Gln	Ile	
ATG Met 65	AAG Lys	AAG Lys	TTG Leu	AAC Asn	CAT His 70	GCC Ala	AAT Asn	GTT Val	Val	AAG Lys 75	GCC Ala	TGT Cys	GAT Asp	GTT Val	CCT Pro 80	240
GAA Glu	GAA Glu	TTG Leu	AAT Asn	ATT Ile 85	TTG Leu	ATT Ile	CAT His	GAT Asp	GTG Val 90	CCT Pro	CTT Leu	CTA Leu	GCA Ala	ATG Met 95	GAA Glu	288
TAC Tyr	TGT Cys	TCT Ser	GGA Gly 100	GGA Gly	GAT Asp	CTC Leu	CGA Arg	AAG Lys 105	CTG Leu	CTC Leu	AAC Asn	AAA Lys	CCA Pro 110	GAA Glu	AAT Asn	336
						AGC Ser										384
GGG Gly	TCT Ser 130	GGG Gly	ATT Ile	CGA Arg	TAT Tyr	TTG Leu 135	CAT His	GAA Glu	AAC Asn	AAA Lys	ATT Ile 140	ATA Ile	CAT	CGA Arg	GAT Asp	432
CTA Leu 145	Lys	CCT Pro	GAA Glu	AAC Asn	ATA Ile 150	Val	CTT Leu	CAG Gln	GAT Asp	GTT Val 155	Gly	GGA Gly	AAG Lys	ATA Ile	ATA Ile 160	480
CAT His	AAA Lys	ATA Ile	ATT	GAT Asp 165	Leu	GGA Gly	тат туг	GCC Ala	AAA Lys 170	GAT Asp	GTT Val	GAT Asp	CAA Gln	GGA Gly 175	AGT Ser	528
CTC	TGI 1 Cys	ACA Thi	TCT Ser 180	Ph∈	GTG Val	GGA Gly	ACA Thr	CTG Leu 185	Gln	ТАТ Тут	CTC	GCC Ala	CCA Pro	Glu	CTC Leu	576
TT.	r GAC e Glu	AAN Asr 199	ı Lys	CCT Pro	TAC Tyr	ACA Thr	GCC Ala 200	Thr	GTT Val	GA? Ası	r TAI	TGC Trp 205	Ser	TTT Phe	r GGG e Gly	624
ACC Th:	C ATO	. Va	A TTT	GAF Glu	A TGT 1 Cys	ATT Ile 215	Ala	GGA Gly	тат Туг	ACC	g CC: g Pro) Phe	r TTC e Lev	G CAS	r CAT s His	672
CT Le	u Gli	G CC	A TTI	r ACC	TGC Trp 230	His	GAC	AAC Lys	ATT	AA(Ly: 23!	s Ly:	G AAG s Ly:	G GA' s Asi	r CC	A AAG o Lys 240	720
TG Cy	T AT	A TT	T GCZ e Ala	A TG' a Cy: 24!	s Glu	A GAC u Glu	ATC	TC/	A GGA c Gly 250	/ G1	A GT u Va	T CG	G TT g Ph	T AG e Se 25	T AGC r Ser 5	768
CA Hi	T TT. s Le	A CC u Pr	T CA o Gl: 26	n Pr	A AA' o Asi	T AGC n Sei	CT:	r TG: 1 Cy: 26:	s Ser	TT Le	A AT u Il	A GT e Va	A GA 1 G1 27	u Pr	C ATG o Met	816
GA G1	A AA u As	C TG n Tr 27	p Le	A CA u Gl	G TTO	G ATO	TTX Lev 28	u Ası	r TGC n Trī	G GA p As	C CC p Pr	T CA o Gl 28	n Gl	G AG n Ar	a GGA g Gly	864

	CCT Pro 290															912
	CAC His						_									960
	AAG Lys															1008
	CAG Gln															1056
	CTT Leu															1104
	CAA Gln 370															1152
	TTG Leu															1200
	AGT Ser									_	_					1248
	CTT Leu		_	_					_		_	_	_	_		1296
	GTG Val		_									_	_			1344
	GCA Ala 450															1392
	AAG Lys															1440
	TTT Phe										_					1488
_	ATG Met	_		_	_			_					_			1536
GAA	ATG	GAA	GAA	AAG	GCC	ATC	CAC	TAT	GCT	GAG	GTT	GGT	GTC	TTA	GGA	1584

Glu	Met	Glu 515	Glu	Lys	Ala		His 520	Tyr	Ala	Glu		Gly 525	Val	Ile	Gly	
TAC Tyr	CTG Leu 530	GAG Glu	GAT Asp	CAG Gln	ATT Ile	ATG Met 535	TCT Ser	TTG Leu	CAT His	GCT Ala	GAA Glu 540	ATC Ile	ATG Met	GGG Gly	CTA Leu	1632
CAG Gln 545	AAG Lys	AGC Ser	CCC Pro	TAT Tyr	GGA Gly 550	AGA Arg	CGT Arg	CAG Gln	GGA Gly	GAC Asp 555	TTG Leu	ATG Met	GAA Glu	TCT Ser	CTG Leu 560	1680
GAA Glu	CAG Gln	CGT Arg	GCC Ala	ATT Ile 565	GAT Asp	CTA Leu	TAT Tyr	AAG Lys	CAG Gln 570	TTA Leu	AAA Lys	CAC His	AGA Arg	CCT Pro 575	TCA Ser	1728
GAT Asp	CAC His	TCC Ser	TAC Tyr 580	AGT Ser	GAC Asp	AGC Ser	ACA Thr	GAG Glu 585	ATG Met	GTG Val	AAA Lys	ATC Ile	ATT Ile 590	GTG Val	CAC His	1776
ACT Thr	GTG Val	CAG Gln 595	Ser	CAG Gln	GAC Asp	CGT Arg	GTG Val 600	CTC Leu	AAG Lys	GAG Glu	CTG Leu	TTT Phe 605	GGT Gly	CAT His	TTG Leu	1824
AGC Ser	AAG Lys 610	Leu	TTG Leu	GGC Gly	TGT Cys	AAG Lys 615	CAG Gln	AAG Lys	ATT Ile	ATT	GAT Asp 620	CTA Leu	CTC	CCT Pro	AAG Lys	1872
GTC Val	GAA Glu	GTG Val	GCC Ala	CTC Leu	AGT Ser 630	AAT Asn	ATC	: AAA : Lys	GAA Glu	GCT Ala 635	Asp	AAT Asn	ACT Thr	GTC Val	ATG Met 640	1920
TTC Phe	ATG Met	CAC Glr	GGA Gly	AAA Lys 645	Arg	CAG Gln	AAA Lys	GAA Glu	ATA Ile 650	Tr	CAT His	CTC Leu	CTI Lev	AAA Lys 655	ATT	1968
GC(TGT Cys	ACA	CAC Glr 660	ser	TCT Ser	GCC	CGC	TCI Ser 665	Leu	GTA Val	A GGA L Gly	TCC Ser	Ser 670	Leu	A GAA 1 Glu	2016
GG' G1	r GCA y Ala	GTA Val 675	l Thi	CCI Pro	CAG	ACA Thr	TCA Ser 680	c Ala	TGC	CTC	ı Pro	CCC Pro 685	Th:	r TCA r Sei	A GCA Ala	2064
GA. G1	A CAT u His 690	a Ası	CAT His	r TC1 s Ser	CTC	TCA Ser 695	Cys	r GTC s Val	GTA L Val	A AC'	r CCS r Pro 700	o Gli	A GA' n Asj	r GG(p Gl)	G GAG y Glu	2112
AC Th 70	r Sei	A GC	A CAJ a Gli	A ATC	710	e Glu	A GAZ	A AA' u Asi	r TTC n Lei	3 AA 1 As: 71	n Cy	CT" s Le	r GG	C CA' y Hi	T TTA s Leu 720	2160
AG Se	C AC' r Thi	r AT	T AT'	r CAS e His 725	s Glu	G GCA 1 Ala	A AA' a As	T GAG	G GAA u Glu 730	ı Gl	G GG n Gl	y As:	T AG n Se	T AT r Me 73	G ATG t Met 5	2208
AA As	T CT n Le	T GA u A s	T TG p Tr 74	p Se	T TGO T Tr	G TT	A AC.	A GA r Gl 74	u Trj	G GT p Va	A CC 1 Pr	G CG o Ar	G GC g Al 75	a Ar	G GAT g Asp	2256

CCA Pro	CCG Pro	GTC Val 755	GCC Ala	ACC Thr	ATG Met	GTG Val	AGC Ser 760	AAG Lys	GGC Gly	GAG Glu	GAG Glu	CTG Leu 765	TTC Phe	ACC Thr	GGG Gly	2304
GTG Val	GTG Val 770	CCC Pro	ATC Ile	CTG Leu	GTC Val	GAG Glu 775	CTG Leu	GAC Asp	GGC Gly	GAC Asp	GTA Val 780	AAC Asn	GGC Gly	CAC His	AAG Lys	2352
TTC Phe 785	AGC Ser	GTG Val	TCC Ser	GGC Gly	GAG Glu 790	GGC Gly	GAG Glu	GGC Gly	GAT Asp	GCC Ala 795	ACC Thr	TAC Tyr	GGC Gly	AAG Lys	CTG Leu 800	2400
ACC Thr	CTG Leu	AAG Lys	TTC Phe	ATC Ile 805	TGC Cys	ACC Thr	ACC Thr	GGC Gly	AAG Lys 810	CTG Leu	CCC Pro	GTG Val	CCC Pro	TGG Trp 815	CCC Pro	2448
ACC Thr	CTC Leu	GTG Val	ACC Thr 820	ACC Thr	CTG Leu	ACC Thr	TAC Tyr	GGC Gly 825	GTG Val	CAG Gln	TGC Cys	TTC Phe	AGC Ser 830	CGC Arg	TAC Tyr	2496
													ATG Met			2544
GGC Gly	TAC Tyr 850	Val	CAG Gln	GAG Glu	CGC	ACC Thr 855	ATC	TTC Phe	TTC Phe	AA G Lys	GAC Asp 860	GAC Asp	GGC Gly	AAC Asn	TAC Tyr	2592
AAG Lys 865	ACC Thr	CGC Arg	GCC Ala	GAG Glu	GTG Val 870	AAG Lys	TTC Phe	GAG Glu	GGC Gly	GAC Asp 875	Thr	CTG	GTG Val	AAC Asn	CGC Arg 880	2640
ATC Ile	GAG Glu	CTG Leu	AAG Lys	GGC Gly 885	Ile	GAC Asp	TTC	AAG Lys	GAG Glu 890	Asp	GGC Gly	AAC Asr	ATC	CTG Leu 895	Gly	2688
CAC His	AAG Lys	CTG Leu	GAG Glu 900	Tyr	AAC Asn	TAC Tyr	AAC Asn	AGC Ser 905	His	AAC Asr	GTC Val	TAT Tyr	11e	Met	GCC Ala	2736
GAC Asp	Lys	Glr	Lys	Asn	GGC Gly	Ile	Lys	GTG Val	AAC Asn	TTC	AAC Lys	925	e Arc	CAC His	AAC Asn	2784
ATC Ile	GAG Glu 930	Asp	GGC Gly	AGC Ser	GTG Val	Gln 935	Lei	GCC Ala	GAC Asp	CAC His	TAC 5 Ty: 940	Glr	G CAC	AAC Asn	ACC Thr	2832
CCC Pro 945	Ile	GGC Gly	GAC Asp	GGC G1y	CCC Pro 950	Val	CTO Let	CTC Leu	CCC Pro	GA6 Asj 95	p Ası	C CAG	ТА(5 Ту)	CTC	AGC Ser 960	2880
ACC Thr	CAC Glr	TCC Sei	GCC Ala	CTC Lev 969	ı Ser	AAA Lys	GA(CCC Pro	AAC Asr 970	ı Gl	G AAG u Ly:	G CG s Ar	C GA' g Ası	r CAC P His 975	ATG Met	2928
GTC	CTC	G CT	G GAC	TT	GT	ACC	GC	C GC(GGC	TA E	C AC'	т ст	c GG	YEA C	G GAC	2976

Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp 980 985 990

GAG CTG TAC AAG TAA Glu Leu Tyr Lys 995 2991

(2) INFORMATION FOR SEQ ID NO:123:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 996 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:123:

Met Glu Arg Pro Pro Gly Leu Arg Pro Gly Ala Gly Gly Pro Trp Glu 10 5 Met Arg Glu Arg Leu Gly Thr Gly Gly Phe Gly Asn Val Cys Leu Tyr 20 25 Gln His Arg Glu Leu Asp Leu Lys Ile Ala Ile Lys Ser Cys Arg Leu 40 Glu Leu Ser Thr Lys Asn Arg Glu Arg Trp Cys His Glu Ile Gln Ile 60 55 Met Lys Lys Leu Asn His Ala Asn Val Val Lys Ala Cys Asp Val Pro 75 70 Glu Glu Leu Asn Ile Leu Ile His Asp Val Pro Leu Leu Ala Met Glu 90 85 Tyr Cys Ser Gly Gly Asp Leu Arg Lys Leu Leu Asn Lys Pro Glu Asn 105 110 100 Cys Cys Gly Leu Lys Glu Ser Gln Ile Leu Ser Leu Leu Ser Asp Ile 120 125 115 Gly Ser Gly Ile Arg Tyr Leu His Glu Asn Lys Ile Ile His Arg Asp 135 130 Leu Lys Pro Glu Asn Ile Val Leu Gln Asp Val Gly Gly Lys Ile Ile 155 150 145 His Lys Ile Ile Asp Leu Gly Tyr Ala Lys Asp Val Asp Gln Gly Ser 165 170 175 Leu Cys Thr Ser Phe Val Gly Thr Leu Gln Tyr Leu Ala Pro Glu Leu 185 190 180 Phe Glu Asn Lys Pro Tyr Thr Ala Thr Val Asp Tyr Trp Ser Phe Gly 200 205 Thr Met Val Phe Glu Cys Ile Ala Gly Tyr Arg Pro Phe Leu His His 220 215 Leu Gln Pro Phe Thr Trp His Glu Lys Ile Lys Lys Lys Asp Pro Lys 230 235 Cys Ile Phe Ala Cys Glu Glu Met Ser Gly Glu Val Arg Phe Ser Ser 255 250 245 His Leu Pro Gln Pro Asn Ser Leu Cys Ser Leu Ile Val Glu Pro Met 260 265 Glu Asn Trp Leu Gln Leu Met Leu Asn Trp Asp Pro Gln Gln Arg Gly 280 285 Gly Pro Val Asp Leu Thr Leu Lys Gln Pro Arg Cys Phe Val Leu Met

	290					295					300				
Asp 305	His	Ile	Leu	Asn	Leu 310	Lys	Ile	Val	His	Ile 315	Leu	Asn	Met	Thr	Ser 320
Ala	Lys	Ile	Ile	Ser 325	Phe	Leu	Leu	Pro	Pro 330	Asp	Glu	Ser	Leu	His 335	Ser
			340					345			Asn		350		
Glu	Leu	Leu 355	Ser	Glu	Thr	Gly	Ile 360	Ser	Leu	Asp	Pro	Arg 365	Lys	Pro	Ala
	370					375					Asp 380				
385					390					395	Gly				400
				405					410		Gln			415	
			420					425			Ala		430		
		435					440				Leu	445			
	450					455					Ala 460				
Met 465		Asn	Thr	Leu	Ile 470	Ser	Ala	Ser	Gln	Gln 475	Leu	Lys	Ala	Lys	Leu 480
Glu	Phe	Phe	His	Lys 485	Ser	Ile	Gln	Leu	Asp 490	Leu	Glu	Arg	Tyr	Ser 495	Glu
			500					505			Leu		510		
		515					520				Val	525			
-	530					535					Glu 540				
545	,				550					555					560
				565	<u>,</u>				570)				575	
			580					585	,				590)	His
		595	5				600)				605	.		Leu
	610	}				615	,				620				Lys
Va] 629		va]			Ser 630				GIU	635		ASI	1 1111	. vai	Met 640
Phe	e Met			645	5				650)				655	
			660)				665	5				670)	ı Glu
		675	5				680)				689	5		Ala
	690)				695	5				700)			/ Glu
70	5				710)				715	5				720
				72	5				730	0				73	
			740	C				74	5				75	0	g Asp
Pr	o Pro	o Vai	l Ala	a Th	r Me	t Va	l Se	r Ly	s Gl	y Gl	u Gl	ı Le	u Ph	e Th	r Gly

760 755 Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys 775 780 Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu 790 795 Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro 815 810 805 Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr 825 Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu 840 Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr 855 860 Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg 870 875 Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly 895 890 885 His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala 910 900 905 Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn 920 925 Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr 935 940 Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser 950 955 Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met 965 970 Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp 990 980 985 Glu Leu Tyr Lys 995

- (2) INFORMATION FOR SEQ ID NO:124:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1908 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...1905
 - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:124:

								TTC Phe					Pro			48
								GGC Gly 25				Ser				96
GAG	GGC	GAG	GGC	GAT	GCC	ACC	TAC	GGC	AAG	CTG	ACC	CTG	AAG	TTC	ATC	144

Glu	Gly	Glu 35	Gly	Asp	Ala	Thr	Tyr 40	Gly	Lys	Leu	Thr	Leu 45	Lys	Phe	Ile	
TGC Cys																192
												GAC Asp				240
												TAC Tyr				288
												ACC Thr				336
												GAG Glu 125				384
												AAG Lys				432
											Asp	AAG Lys				480
										Asn		GAG Glu			Ser	528
				Asp					Asn					Asp	GGC	576
CCC Pro	GTG Val	CTG Leu 195	Leu	CCC Pro	GAC Asp	AAC Asn	CAC His	Tyr	CTG Leu	AGC Ser	ACC Thr	CAG Gln 205	Ser	GCC Ala	CTG Leu	624
AGC Ser	AAA Lys 210	Asp	CCC	: AAC Asn	GAG Glu	AAG Lys 215	Arg	GAT Asp	CAC His	ATC Met	GTC Val 220	Leu	CTC	G GAC	TTC Phe	672
GTG Val 225	Thr	GCC Ala	GCC Ala	GGG Gly	ATC 11e 230	Thr	CTC Lev	GGC Gly	ATC	GAC Asp 235	o Glu	CTC	ТАС ТУ1	AAC Lys	TCC Ser 240	720
GGA Gly	CTC Lev	AGA Arg	TC1	CGA Arg 245	Ala	CAA Glr	GCT Ala	TCC Ser	ATC Met	: Sei	GAC	G ACC	GTY Vai	25!	ATG Met	768
				l Ile					g Ala					и Ту	r GAT r Asp	816

TAD qzA	GGC Gly	AAC Asn 275	AAG Lys	CGA Arg	TGG Trp	Leu	CCT Pro 280	GCT Ala	GGC Gly	ACG Thr	GGT Gly	CCC Pro 285	CAG Gln	GCC Ala	TTC Phe	864
AGC Ser	CGC Arg 290	GTC Val	CAG Gln	ATC Ile	TAC Tyr	CAC His 295	AAC Asn	CCC Pro	ACG Thr	GCC Ala	AAT Asn 300	TCC Ser	TTT Phe	CGC Arg	GTC Val	912
GTG Val 305	GGC Gly	CGG Arg	AAG Lys	ATG Met	CAG Gln 310	CCC Pro	GAC Asp	CAG Gln	CAG Gln	GTG Val 315	GTC Val	ATC Ile	AAC Asn	TGT Cys	GCC Ala 320	960
ATC Ile	GTC Val	CGG Arg	GGT Gly	GTC Val 325	AAG Lys	TAT Tyr	AAC Asn	CAG Gln	GCC Ala 330	ACC Thr	CCC Pro	AAC Asn	TTC Phe	CAT His 335	CAG Gln	1008
TGG Trp	CGC Arg	GAC Asp	GCT Ala 340	CGC Arg	CAG Gln	GTC Val	TGG Trp	GGC Gly 345	CTC Leu	AAC Asn	TTC Phe	GGC Gly	AGC Ser 350	Lys	GAG Glu	1056
GAT Asp	GCG Ala	GCC Ala 355	CAG Gln	TTT Phe	GCC Ala	GCC Ala	GGC Gly 360	Met	GCC Ala	AGT Ser	GCC Ala	CTA Leu 365	Glu	GCG Ala	TTG Leu	1104
GAA Glu	GGA Gly 370	Gly	GGG Gly	CCC Pro	CCT Pro	CCA Pro 375	CCC	CCA Pro	GCA Ala	CTT	CCC Pro 380	Thr	TGG Trp	TCG Ser	GTC Val	1152
CCG Pro 385	Asn	GGC Gly	CCC Pro	TCC Ser	CCG Pro 390	Glu	GAG Glu	GTG Val	GAG Glu	Gln 395	Gln	AAA Lys	AGG Arg	G CAC	CAG Gln 400	1200
CCC	GGC Gly	CCG Pro	TCG Ser	GAG Glu 405	His	ATA Ile	GAC Glu	G CGC	CGG Arg 410	Val	TCC Ser	AAT Asr	GCA n Alá	A GGA A Gly 415	A GGC / Gly	1248
CCA Pro	CCI Pro	GCI Ala	CCC Pro 420	Pro	GCT Ala	GGG Gly	GGT Gly	r CCA / Pro 425	Pro	CCA Pro	A CCA	A CCA	430	y Pro	r CCC p Pro	1296
CCI Pro	CCI Pro	r CCA Pro 435	Gly	CCC Pro	Pro	Pro	Pro	C CCA D Pro	Gl3	, Te	ı Pro	C CC' D Pro 445	Se:	G GGG r Gl	G GTC y Val	1344
CC; Pro	A GCT Ala 450	a Ala	A GCC a Ala	G CAC	GG? Gly	A GCA Ala 455	Gly	g GG/ y Gl	A GG/ / Gl ₂	A CC	A CCC O Pro 460	o Pro	T GC.	A CC	C CCT o Pro	1392
CT0 Lev 465	ı Pro	G GCZ o Ala	A GCA a Ala	A CAC	G GGG 1 Gly 470	y Pro	GG' Gly	r GG y Gly	r GG(y Gly	G GG. y G1; 47	y A1	T GG a Gl	g GC y Al	C CC a Pr	A GGC o Gly 480	1440
CT(Let	G GCC	C GC. a Al	A GC' a Ala	r AT a Ile 48	e Ala	r GG/ a Gly	A GC	C AA a Ly:	A CTO s Lev 49	u Ar	G AA. g Ly	A GT s Va	C AG 1 Se	C AA r Ly 49	G CAG s Gln	1488
GA(G GA	G GC	C TC	A. GGK	G GG	G CC	CAC	A GC	c cc	C AA	A GC	T GA	G AG	T GG	T CGA	1536

Glu	Glu	Ala	Ser 500	Gly	Gly	Pro	Thr	Ala 505	Pro	Lys	Ala	Glu	Ser 510	Gly	Arg	
			_											GCC Ala		1584
														GAA Glu		1632
														GAA Glu		1680
														ATG Met 575		1728
														CCC Pro		1776
														CTG Leu		1824
														GAA Glu		1872
						AAG Lys			-		TGA					1908

(2) INFORMATION FOR SEQ ID NO:125:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 635 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:125:

 Met
 Val
 Ser
 Lys
 Gly
 Glu
 Glu
 Leu
 Phe
 Thr
 Gly
 Val
 Pro
 Ile
 Leu

 Val
 Glu
 Leu
 Asp
 Gly
 Asp
 Gly
 His
 Lys
 Phe
 Ser
 Val
 Ser
 Gly

 Glu
 Gly
 Gly
 Asp
 Ala
 Thr
 Tyr
 Gly
 Leu
 Thr
 Leu
 Leu
 Thr
 Ile
 Lys
 Phe
 Ile
 Ile
 Leu
 Ile
 Leu
 Ile
 Ile

65			7	70					75					80
Gln His			85					90					95	
Arg Thr		100					105					110		
Val Lys	115					120					125			
Ile Ası)			-	135					140				
Asn Ty	r Asn	Ser		Asn \ 150	Val	Tyr	Ile	Met	A1a 155	Asp	Lys	GIn	гуѕ	160
145 Gly Il	e Lys	Val			Lys	Ile	Arg	His 170		Ile	Glu	Asp	Gly 175	Ser
Val Gl		180					185					190		
Pro Va	195	,				200					205			
Ser Ly 21	0				215					220				
Val Th	r Ala	Ala	Gly	11e 230	Thr	Leu	Gly	Met	. Asp 235	GIU	Leu	ТУГ	БĀЗ	240
Gly Le	u Arg	g Ser		Ala	Gln	Ala	Ser	Met 250	Ser	Glu	Thr	Val	11e 255	Met
Ser Gl	u Thi	val 260		Cys	Ser	Ser	Arg 265	Ala		· Val	Met	Leu 270	Тут	
Asp Gl	Ly Asi	n Lys	Arg	Trp	Leu	Pro 280		Gl7	/ Thr	Gly	Prc 285	Gln	Ala	Phe
Ser A	rg Va	l Gln			295					300)			
Val G	ly Ar	g Lys	Met		Pro	Asp	Glr	Glr	n Va. 31:	l Val	. Il€	e Asr	СУ	320
305 Ile Va	al Ar	g Gly	. Val	310 Lys	Tyr	Asn	Glr	Ala) Asr	n Phe	His	s Gln
Trp A			325					33	0				33)
		340)				345	5				350)	
Asp A	35	5				360)				36	5		
Glu G 3	70				375	5				38	0			
	sn Gl	y Pro	Ser	Pro 390		ı Glı	ı Va	l Gl	u G1 39	n Gl: 5	n Ly	s Ar	g Gl	n Gln 400
385 Pro G	ly Pr	o Se	c Glu	His	Ile	≘ Glı	u Ar	g Ar 41	g Va		r As	n Al	a Gl 41	y Gly 5
Pro P	ro Al	a Pro	o Pro	Ala	Gl			o Pr				o Gl 43	y Pr O	o Pro
Pro F		co Gl;	y Pro	Pro	Pro	o Pr	o Pr		y Le	eu Pr	o Pr 44	o Se 5	r Gl	y Val
4	la A	la Al			45	5				46	0			o Pro
Leu E	Pro A			470)				47	75				60 Gly
Leu A	Ala A	la Al	a Ile 489		G1;	y Al	a Ly	s Le	eu Ar 90	g Ly	's Va	al S∈	r Ly 49	/s Gln 95
		50	r Gly	y Gly			50	a Pi	ro L			51	er G:	ly Arg
	5	ly Gl 15	y Gl			52	20				52	25		la Arg
Arg A	Arg L	ys Al	a Th	r Gli	n Va	l Gl	y Gl	u L	ys T	hr Pi	co L	ys As	p G	lu Ser

530		535			540				
Ala Asn Gln G		o Glu Ala	Arg V	al Pro 555	Ala G	ln Ser	Glu	Ser 560	
545 Val Arg Arg P	55) ro Trp Gl	u Lys Asn	Ser T	hr Thr	Leu P	ro Arg	g Met		
Ser Ser Ser S	565		5	70			575		
5	80		585			590	J		
Ser Ser Asp T		600)		6	05			
Glu Val Lys L		615			620	le Il	e Glu	Ala	
Phe Val Gln G 625	lu Leu Ar 63		g Gly S	Ser Pro 635					
(2)	INFORMATI	ON FOR S	EQ ID 1	NO:126:					
	UENCE CHA								
	LENGTH: 13 TYPE: nucl		pairs						
(C) :	STRANDEDNE	ESS: sing	le						
	TOPOLOGY:								
	DLECULE TY	YPE: cDNA							
	NAME/KEY			ce					
	LOCATION OTHER IN								
/ C	בטו זוביאור בי די	ECCRIPTIO	N SEC	ON CIT	:126:				
	EQUENCE D					CTY - C	ርር <i>እ</i> ሞየ	r CTG	48
MENC CITY &CCC	ANG GGC G	AG GAG CI	G TTC	ACC GG	G GTG	GTG C	ro II	C CTG e Leu	48
	ANG GGC G	AG GAG CI	G TTC	ACC GG	G GTG	GTG C	CC ATO ro Ilo 15	C CTG e Leu	48
ATG GTG AGC Met Val Ser 1	AAG GGC G Lys Gly G 5 GAC GGC G	AG GAG CT lu Glu Le	TTC Phe AC GGC	ACC GG Thr Gl 10	G GTG y Val	AGC G	TG TC	c GGC	48 96
ATG GTG AGC Met Val Ser 1	AAG GGC G Lys Gly G 5 GAC GGC G	AG GAG CT lu Glu Le	TTC Phe AC GGC	ACC GG Thr Gl 10	G GTG y Val	AGC G Ser V	TG TC	c GGC	
ATG GTG AGC Met Val Ser 1 GTC GAG CTG Val Glu Leu	AAG GGC G Lys Gly G 5 GAC GGC G Asp Gly A 20	AG GAG CT lu Glu Le AC GTA AA ASP Val AS	ng TTC eu Phe AC GGC sn Gly 25 AC GGC	ACC GG Thr Gl 10 CAC AA His Ly	G GTG y Val G TTC rs Phe	AGC G Ser V 3	TG TCG TG TCG TAG TCG TAG TTG TAG TTG	c GGC r Gly	
ATG GTG AGC Met Val Ser 1 GTC GAG CTG Val Glu Leu GAG GGC GAG Glu Gly Glu	AAG GGC G Lys Gly G 5 GAC GGC G Asp Gly A 20	AG GAG CT lu Glu Le AC GTA AA ASP VAI AS	AC GGC Shall GGC GGC GGC GGC GGC GGC GGC GGC GGC G	ACC GG Thr Gl 10 CAC AA His Ly	G GTG y Val G TTC rs Phe	AGC G Ser V 3 CTG A Leu L	TG TCG TG TCG TAG TCG TAG TTG TAG TTG	c GGC r Gly	96
ATG GTG AGC Met Val Ser 1 GTC GAG CTG Val Glu Leu GAG GGC GAG Glu Gly Glu 35	AAG GGC G Lys Gly G 5 GAC GGC G Asp Gly A 20 GGC GAT G	AG GAG CT lu Glu Le AC GTA AA sp Val As GCC ACC TA Ala Thr T	rG TTC eu Phe AC GGC 5n Gly 25 AC GGC yr Gly 0	ACC GG Thr G1 10 CAC AA His Ly AAG C1 Lys Le	G GTG y Val G TTC rs Phe TG ACC eu Thr	AGC G Ser V 3 CTG A Leu I	TG TC (al Se 0	C GGC r Gly C ATC	96 144
ATG GTG AGC Met Val Ser 1 GTC GAG CTG Val Glu Leu GAG GGC GAG Glu Gly Glu 35	AAG GGC G Lys Gly G 5 GAC GGC G Asp Gly A 20 GGC GAT G Gly Asp A	AG GAG CT lu Glu Le AC GTA AA SP Val AS SCC ACC TA Ala Thr T	rG TTC eu Phe AC GGC sn Gly 25 AC GGC yr Gly 0	ACC GG Thr G1 10 CAC AA His Ly AAG C1 Lys Le	G GTG y Val G TTC rs Phe rg ACC eu Thr	AGC G Ser V 3 CTG A Leu L 45	TG TC	C GGC r Gly C ATC e Ile	96
ATG GTG AGC Met Val Ser 1 GTC GAG CTG Val Glu Leu GAG GGC GAG Glu Gly Glu 35	AAG GGC G Lys Gly G 5 GAC GGC G Asp Gly A 20 GGC GAT G Gly Asp A	AG GAG CT lu Glu Le AC GTA AA SP Val AS SCC ACC TA Ala Thr T	rG TTC eu Phe AC GGC sn Gly 25 AC GGC yr Gly 0	ACC GG Thr G1 10 CAC AA His Ly AAG C1 Lys Le	G GTG y Val G TTC rs Phe rg ACC eu Thr	AGC G Ser V 3 CTG A Leu L 45	TG TC	C GGC r Gly C ATC e Ile	96 144
ATG GTG AGC Met Val Ser 1 GTC GAG CTG Val Glu Leu GAG GGC GAG Glu Gly Glu 35 TGC ACC ACC Cys Thr Thr 50	AAG GGC G Lys Gly G 5 GAC GGC G Asp Gly A 20 GGC GAT G GGC AAG G Gly Lys I	AG GAG CT lu Glu Le AC GTA AA ASP Val AS ACC ACC TA Ala Thr TA ACC CCC G Leu Pro V 55	ng TTC eu Phe AC GGC sn Gly 25 AC GGC yr Gly 0 TG CCC al Pro	ACC GG Thr G1 10 CAC AA His Ly AAG CT Lys Le TGG CC Trp Pr	G GTG y Val G TTC rs Phe TG ACC Thr GC ACC TO Thr GO ACC TO Thr	AGC G Ser V 3 CTG A Leu I 45 CTC C Leu V	TG TC: TG TC: AG TT	C GGC r Gly C ATC e Ile C ACC nr Thr	96 144
ATG GTG AGC Met Val Ser 1 GTC GAG CTG Val Glu Leu GAG GGC GAG Glu Gly Glu 35 TGC ACC ACC Cys Thr Thr 50 CTG ACC TAC Leu Thr Tyr	AAG GGC G Lys Gly G S GAC GGC G Asp Gly A 20 GGC GAT G Gly Asp A GGC AAG C Gly Lys I GGC GTG C Gly Val C	AG GAG CT lu Glu Le AC GTA AA ASP Val AS ACC ACC TA Ala Thr TA ACC CCC G Leu Pro V 55	ng TTC eu Phe AC GGC sn Gly 25 AC GGC yr Gly 0 TG CCC al Pro	ACC GG Thr G1 10 CAC AA His Ly AAG CT Lys Le TGG CC Trp Pr	G GTG y Val G TTC rs Phe G ACC eu Thr GC ACC ro Thr 60 AC CCC yr Pro	AGC G Ser V 3 CTG A Leu I 45 CTC C Leu V	TG TC: TG TC: AG TT	C GGC r Gly C ATC e Ile C ACC nr Thr	96 144 192
ATG GTG AGC Net Val Ser 1 GTC GAG CTG Val Glu Leu GAG GGC GAG Glu Gly Glu 35 TGC ACC ACC Cys Thr Thr 50 CTG ACC TAC Leu Thr Tyr 65	AAG GGC G Lys Gly G 5 GAC GGC G Asp Gly A 20 GGC GAT G Gly Asp A GGC AAG G Gly Lys I GGC GTG G Gly Val G	AG GAG CTIU GIU Le AC GTA AM ASP VAI AS ACC ACC TI AIA THE TI ACC CCC G Leu Pro V 55 CAG TGC T	AC GGC Sn Gly 25 AC GGC yr Gly 0 TG CCC al Pro	ACC GG Thr G1 10 CAC AA His Ly AAG CT Lys Le TGG CC Trp Pr CGC Tr Arg T	G GTG y Val G TTC rs Phe G ACC ro Thr 60 AC CCC yr Pro 5	AGC G Ser V 3 CTG A Leu I 45 CTC C Leu V	TG TC: TG TC: AG TT LAG TT LYS Ph CAC AT His Me	C GGC r Gly C ATC e Ile C ACC nr Thr CG AAG et Lys 80	96 144 192
ATG GTG AGC Met Val Ser 1 GTC GAG CTG Val Glu Leu GAG GGC GAG Glu Gly Glu 35 TGC ACC ACC Cys Thr Thr 50 CTG ACC TAC Leu Thr Tyr	AAG GGC G Lys Gly G 5 GAC GGC G Asp Gly A 20 GGC GAT G Gly Asp A GGC AAG C Gly Lys I GGC GTG C Gly Val C TTC TTC Phe Phe :	AG GAG CTIU GIU Le AC GTA AM ASP VAI AS ACC ACC TI AIA THE TI AIC CCC G Leu Pro V 55 CAG TGC T GIN Cys P	AC GGC En Gly 25 AC GGC yr Gly 0 TG CCC al Pro	ACC GG Thr G1 10 CAC AA His Ly AAG CT Lys Le TGG CC Trp Pr CGC Tr Arg Tr 7 CCC G Pro G	G GTG y Val G TTC rs Phe G ACC ro Thr 60 AC CCC yr Pro 5	AGC G Ser V 3 CTG A Leu I 45 CTC C Leu V	TG TC: AG TT A	C GGC r Gly C ATC e Ile C ACC IT Thr C AAG	96 144 192 240
ATG GTG AGC Net Val Ser 1 GTC GAG CTG Val Glu Leu GAG GGC GAG Glu Gly Glu 35 TGC ACC ACC Cys Thr Thr 50 CTG ACC TAC Leu Thr Tyr 65 CAG CAC GAC Gln His Asp	AAG GGC G Lys Gly G 5 GAC GGC G Asp Gly A 20 GGC GAT G Gly Asp A GGC AAG G Gly Lys I GGC GTG G Gly Val G TTC TTC TTC TTC TTC TTC TTC TTC TTC 85	AG GAG CTILL GAG GAG GAG AG AG TCC GAG TCC GAG TCC TG GAG TCC GAG TCC GAG TCC TCG CCC TCG TCC TCC	AC GGC Sn Gly 25 AC GGC yr Gly 0 TG CCC al Pro TC AGC he Ser	ACC GG Thr G1 10 CAC AA His Ly AAG CT Lys Le TGG CC Trp Pr CGC Tr Arg Tr 7 CCC G Pro G	G GTG y Val G TTC rs Phe TG ACC Thr 60 AC CCC yr Pro 5 AA GGC lu Gly	AGC G Ser V 3 CTG A Leu I 45 CTC C Leu V	TG TC: AG TT VS Ph CAC AT His Me GTC CA Val G: 9!	C GGC r Gly C ATC le Ile C ACC lr Thr CG AAG et Lys 80 AG GAG ln Glu 5	96 144 192 240
ATG GTG AGC Net Val Ser 1 GTC GAG CTG Val Glu Leu GAG GGC GAG Glu Gly Glu 35 TGC ACC ACC Cys Thr Thr 50 CTG ACC TAC Leu Thr Tyr 65	AAG GGC G Lys Gly G 5 GAC GGC G Asp Gly A 20 GGC GAT G Gly Asp A GGC AAG G Gly Lys I GGC GTG G Gly Val G TTC	AG GAG CT Lu Glu Le La	AC GGC Sn Gly 25 AC GGC Yr Gly 0 TG CCC al Pro TC AGC Ser AGGC ATG AGC AGGC ATG AGC AGGC AGGC AGG	ACC GG Thr G1 10 CAC AA His Ly AAG CT Lys Le TGG CC Trp Pr CGC Tr Arg Tr 7 CCC G Pro G 90 AAC T	G GTG y Val G TTC rs Phe TG ACC PU Thr GC ACC TO Thr 60 AC CCC yr Pro 5 AA GGC lu Gly AC AAC	AGC G Ser V 3 CTG A Leu I 45 CTC C Leu V GAC C Tyr S	TG TC: AG TTC AG TTT AG TT A	C GGC r Gly C ATC le Ile C ACC lr Thr CG AAG et Lys 80 AG GAG ln Glu 5	96 144 192 240

GTG Val	AAG Lys	TTC Phe 115	GAG Glu	GGC Gly	GAC . Asp '	Thr	CTG Leu 120	GTG Val	AAC Asn	CGC Arg	ATC Ile	GAG Glu 125	CTG Leu	AAG Lys	GGC Gly	384
ATC Ile	GAC Asp 130	TTC Phe	AAG Lys	GAG Glu	Asp	GGC Gly 135	AAC Asn	ATC Ile	CTG Leu	GGG Gly	CAC His 140	AAG Lys	CTG Leu	GAG Glu	TAC Tyr	432
AAC Asn 145	TAC Tyr	AAC Asn	AGC Ser	CAC His	AAC Asn 150	GTC Val	тат Туг	ATC Ile	ATG Met	GCC Ala 155	GAC Asp	AAG Lys	CAG Gln	AAG Lys	AAC Asn 160	480
GGC Gly	ATC Ile	AAG Lys	GTG Val	AAC Asn 165	TTC Phe	AAG Lys	ATC Ile	CGC Arg	CAC His 170	AAC Asn	ATC Ile	GAG Glu	GAC Asp	GGC Gly 175	AGC Ser	528
GTG Val	CAG Gln	CTC Leu	GCC Ala 180	GAC Asp	CAC His	TAC Tyr	CAG Gln	CAG Gln 185	AAC Asn	ACC Thr	CCC Pro	ATC	GGC Gly 190	GAC Asp	GGC	576
CCC Pro	GTG Val	CTG Leu 195	Leu	CCC Pro	GAC Asp	AAC Asn	CAC His	Tyr	CTG Leu	AGC Ser	ACC Thr	CAG Gln 205	Ser	GCC Ala	CTG	624
AGC Ser	AAA Lys 210	Asp	CCC Pro	AAC Asn	GAG Glu	AAG Lys 215	CGC	GAT Asp	CAC His	ATG Met	GTC Val 220	Lev	CTG Leu	GAG Glu	TTC Phe	672
GTC Val 225	. Thr	GCC Ala	GCC Ala	GGG Gly	ATC Ile 230	Thr	CTC	GGC Gly	: ATG / Met	GAC Asp 235	Glu	CTC Leu	TAC Tyr	AAC Lys	G TCC Ser 240	720
GG? Gly	CTC	: AGA	A TCI J Ser	CGA Arg	, Ala	CAA Gln	GCT Ala	r TCA a Ser	A ATC Met 250	Ala	GCC Ala	OTA C	CGC Arg	E AAC J Lys 255	G AAA S Lys	768
CT(Lev	GTC ı Val	AT'	r GT1 e Val	. Gl ₃	GAT Asp	GGA Gly	GCC Ala	TG:	s Gly	A AAC	F ACA	A TGO	TTX s Let 270	ı Lei	C ATA u Ile	816
GTY Va	C TTO	2 AG	r Lys	G GAC	CAC Glr	TTC	CC Pro	o Gl	G GTG u Val	G TAT	r GTV	G CC 1 Pr 28	o Thi	A GT r Va	G TTT l Phe	864
GA:	G AAG u Asi 29	а Ту	T GT(r Vai	G GCZ l Ala	A GAT a Asp	7 ATC 5 Ile 299	e Gl	G GTv u Va	G GA' 1 Asi	r GG/ p Gly	A AA y Ly 30	s Gl	G GT n Va	A GA 1 G1	G TTG u Leu	912
GC Al 30	a Le	G TG u Tr	G GAG p As	C AC.	A GCT r Ala 310	a Gl	g CA y Gl	G GA n Gl	A GA' u As	т та р ту: 31	r As	T CG p Ar	C CT	G AG u Ar	G CCC g Pro 320	
CT Le	C TC u Se	C TA r Ty	C CC r Pr	A GA o As 32	p Th	C GA	T GT p Va	T AT	A CT e Le 33	u Me	G TG t Cy	T TI	T TO	C AT	C GAC Le Asp 35	1008
AG	e cc	T GA	T AG	т тт	A GA	A AA	C AI	rc cc	A GA	AA A.	G TO	G AC	cc cc	A G	AA GTC	1056

Ser	Pro	Asp	Ser 340	Leu	Glu	Asn	Ile	Pro 345	Glu	Lys	Trp	Thr	Pro 350	Glu	Val	
						GTG Val										1104
						CAC His 375										1152
						GAA Glu										1200
						GAG Glu										1248
						GCT Ala					-					1296
						TGC Cys				TGA						1329

(2) INFORMATION FOR SEQ ID NO:127:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 442 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:127:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 1 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 20 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 45 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 60 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 70 75 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 105 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 120 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr

135 130 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 150 155 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 180 185 190 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 200 205 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 220 210 215 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 235 230 Gly Leu Arg Ser Arg Ala Gln Ala Ser Met Ala Ala Ile Arg Lys Lys 250 255 245 Leu Val Ile Val Gly Asp Gly Ala Cys Gly Lys Thr Cys Leu Leu Ile 270 265 260 Val Phe Ser Lys Asp Gln Phe Pro Glu Val Tyr Val Pro Thr Val Phe 2.85 275 280 Glu Asn Tyr Val Ala Asp Ile Glu Val Asp Gly Lys Gln Val Glu Leu 300 295 Ala Leu Trp Asp Thr Ala Gly Gln Glu Asp Tyr Asp Arg Leu Arg Pro 310 315 Leu Ser Tyr Pro Asp Thr Asp Val Ile Leu Met Cys Phe Ser Ile Asp 330 335 325 Ser Pro Asp Ser Leu Glu Asn Ile Pro Glu Lys Trp Thr Pro Glu Val 350 340 345 Lys His Phe Cys Pro Asn Val Pro Ile Ile Leu Val Gly Asn Lys Lys 360 365 355 Asp Leu Arg Asn Asp Glu His Thr Arg Arg Glu Leu Ala Lys Met Lys 380 375 Gln Glu Pro Val Lys Pro Glu Glu Gly Arg Asp Met Ala Asn Arg Ile 395 390 Gly Ala Phe Gly Tyr Met Glu Cys Ser Ala Lys Thr Lys Asp Gly Val 405 410 415 Arg Glu Val Phe Glu Met Ala Thr Arg Ala Ala Leu Gln Ala Arg Arg 425 Gly Lys Lys Lys Ser Gly Cys Leu Val Leu 440

(2) INFORMATION FOR SEQ ID NO:128:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1140 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...1137
 - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:128:

Met 1	Asp	His	Tyr	Asp 5	Ser	Gln	Gln	Thr	Asn 10	Asp	Tyr	Met	Gln	Pro 15	Glu	
GAG Glu	GAC Asp	TGG Trp	GAC Asp 20	CGG Arg	GAC Asp	CTG Leu	CTC Leu	CTG Leu 25	GAC Asp	CCG Pro	GCC Ala	TGG Trp	GAG Glu 30	AAG Lys	CAG Gln	96
CAG Gln	AGA Arg	AAG Lys 35	ACA Thr	TTC Phe	ACG Thr	GCA Ala	TGG Trp 40	TGT Cys	AAC Asn	TCC Ser	CAC His	CTC Leu 45	CGG Arg	AAG Lys	GCG Ala	144
GGG Gly	ACA Thr 50	CAG Gln	ATC	GAG Glu	AAC Asn	ATC Ile 55	GAA Glu	GAG Glu	GAC Asp	TTC Phe	CGG Arg 60	GAT Asp	GGC Gly	CTG Leu	AAG Lys	192
CTC Leu 65	ATG Met	CTG Leu	CTG Leu	CTG Leu	GAG Glu 70	GTC Val	ATC Ile	TCA Ser	GGT Gly	GAA Glu 75	CGC Arg	TTG Leu	GCC Ala	AAG Lys	CCA Pro 80	240
GAG Glu	CGA Arg	GGC Gly	AAG Lys	ATG Met 85	AGA Arg	GTG Val	CAC His	AAG Lys	ATC Ile 90	TCC Ser	AAC Asn	GTC Val	: AAC . Asr	AAC Lys 95	GCC Ala	288
CTG Leu	GAT Asp	TTC Phe	ATA	Ala	AGC Ser	AAA Lys	GGC Gly	GTC Val	Lys	CTG	GTC Val	TCC Ser	110	e Gly	A GCC / Ala	336
GAA Glu	GAA Glu	ATC 11e	val	GAT Asp	Gly	AAT Asn	GTG Val 120	Lys	ATG Met	ACC Thr	CTC	GGG Gl _y 125	/ Me	G ATO	TGG Trp	384
ACC Thr	130	Ile	CTC	G CGC	AGG Arg	GAT Asp 135	Pro	CCC Pro	GTC Val	GCC Ala	ACC Thi	Me	G GTV	G AGG	C AAC r Lys	432
GGC Gly 145	/ Glu	GAC Glu	G CTC	TTC 1 Phe	ACC Thr	Gl3	GTC Val	GTC Val	CCC Pro	110 15	e Lei	G GT u Va	C GA 1 G1	G CT u Le	G GAG u Ası 160)
GG(Gl _y	C GAC	GTA Val	AAA laa l	C GG(n Gl _y 165	/ His	AAC Lys	TTC Phe	: AGC e Sei	GTC r Val	l Se:	c GGG r Gl	C GA y Gl	G GG u Gl	C GA y Gl 17	G GGG u Gly 5	528 Y
GAT Ası	r GCC p Ala	C ACC	C TAC r Ty: 18	r Gly	C AAC y Ly:	G CTO	G ACC	CTX r Lev 18	u Lys	G TT	C AT e Il	C TG e Cy	C AC s Tr	ir Th	c GG r Gl	C 576 Y
AA(Ly:	G CTY s Le	G CC u Pr 19	o Va	G CCO	C TGO	G CCO	20	r Le	C GTY u Va	3 AC 1 Th	C AC r Th	C CT r Le	u Tì	C TA	AC GG Mr Gl	C 624 Y
GTY Va	G CA 1 G1 21	n Cy	C TT s Ph	C AG e Se	c cg r Ar	C TA g Ty 21	r Pr	C GA o As	C CA p Hi	C AT s Me	G AA t Ly 22	's G]	AG CZ ln H:	AC GA	AC TI sp Ph	C 672 le
TT Ph 22	e Ly	G TC s Se	c GC r Al	C AT a Me	G CC t Pr 23	o Gl	A GG u Gl	с та у ту	C GT T Va	C CA 1 G1 23	n Gl	kG CC	GC AG	CC Ar hr I	TC TT le Ph 24	re

TTC Phe	AAG Lys	GAC Asp	GAC Asp	GGC Gly 245	AAC Asn	TAC Tyr	AAG Lys	ACC Thr	CGC Arg 250	GCC Ala	GAG Glu	GTG Val	AAG Lys	TTC Phe 255	GAG Glu	768
GGC Gly	GAC Asp	ACC Thr	CTG Leu 260	GTG Val	AAC Asn	CGC Arg	ATC Ile	GAG Glu 265	CTG Leu	AAG Lys	GGC Gly	ATC Ile	GAC Asp 270	TTC Phe	AAG Lys	816
GAG Glu	GAC Asp	GGC Gly 275	AAC Asn	ATC Ile	CTG Leu	GGG Gly	CAC His 280	AAG Lys	CTG Leu	GAG Glu	TAC Tyr	AAC Asn 285	TAC Tyr	AAC Asn	AGC Ser	864
CAC His	AAC Asn 290	GTC Val	TAT Tyr	ATC Ile	ATG Met	GCC Ala 295	GAC Asp	AAG Lys	CAG Gln	AAG Lys	AAC Asn 300	GGC Gly	ATC Ile	AAG Lys	GTG Val	912
AAC Asn 305	TTC Phe	AAG Lys	ATC Ile	CGC Arg	CAC His 310	AAC Asn	ATC	GAG Glu	GAC Asp	GGC Gly 315	Ser	GTG Val	CAG Gln	CTC Leu	GCC Ala 320	960
GAC Asp	CAC His	TAC Tyr	CAG Gln	CAG Gln 325	AAC Asn	ACC Thr	CCC	ATC Ile	GGC Gly 330	Asp	GGC Gly	CCC Pro	GTG Val	CTG Leu 335	Leu	1008
CCC Pro	GAC Asp	AAC Asn	CAC His	Туг	CTG Leu	AGC Ser	ACC Thr	CAG Gln 345	Ser	GCC Ala	CTO	AGC Ser	Lys 350	Asp	CCC Pro	1056
AAC Asn	GAG	AAG Lys	Arg	GAT Asp	CAC His	ATG Met	GTC Val 360	Leu	CTC	GAC 1 Glu	TTC Phe	GTC Val 365	Thr	GCC Ala	GCC Ala	1104
		Thi			ATC		Gl					Ą				1140

(2) INFORMATION FOR SEQ ID NO:129:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 379 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:129:

 Leu Met Leu Leu Glu Val Ile Ser Gly Glu Arg Leu Ala Lys Pro 75 Glu Arg Gly Lys Met Arg Val His Lys Ile Ser Asn Val Asn Lys Ala 90 Leu Asp Phe Ile Ala Ser Lys Gly Val Lys Leu Val Ser Ile Gly Ala 105 Glu Glu Ile Val Asp Gly Asn Val Lys Met Thr Leu Gly Met Ile Trp 120 125 Thr Ile Ile Leu Arg Arg Asp Pro Pro Val Ala Thr Met Val Ser Lys 135 140 Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp 150 155 Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly 165 170 Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly 185 Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly 200 Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp Phe 215 Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe 230 235 Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu 245 250 Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys 260 265 Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser 280 His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val 295 Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala 310 315 Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu 325 330 Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro 340 345 350 Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala 360 Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 375

(2) INFORMATION FOR SEQ ID NO:130:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3516 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...3513
 - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:130:

ATG Met 1	GTG Val	AGC Ser	AAG Lys	GGC Gly 5	GAG Glu	GAG Glu	CTG Leu	TTC Phe	ACC Thr 10	GGG Gly	GTG Val	GTG Val	CCC Pro	ATC Ile 15	CTG Leu	48
GTC Val	GAG Glu	CTG Leu	GAC Asp 20	Gly	GAC Asp	GTA Val	AAC Asn	GGC Gly 25	CAC His	AAG Lys	TTC Phe	AGC Ser	GTG Val 30	TCC Ser	GGC Gly	96
GAG Glu	GGC Gly	GAG Glu 35	GGC Gly	GAT Asp	GCC Ala	ACC Thr	TAC Tyr 40	GGC Gly	AAG Lys	CTG Leu	ACC Thr	CTG Leu 45	AAG Lys	TTC Phe	ATC Ile	144
TGC Cys	ACC Thr 50	ACC Thr	GGC Gly	AAG Lys	CTG Leu	CCC Pro 55	GTG Val	CCC Pro	TGG Trp	CCC Pro	ACC Thr 60	CTC Leu	GTG Val	ACC Thr	ACC Thr	192
CTG Leu 65	ACC Thr	TAC Tyr	GGC	GTG Val	CAG Gln 70	TGC Cys	TTC Phe	AGC Ser	CGC Arg	TAC Tyr 75	CCC Pro	GAC Asp	CAC His	ATG Met	AAG Lys 80	240
CAG Gln	CAC His	GAC Asp	TTC Phe	TTC Phe 85	AAG Lys	TCC Ser	GCC Ala	ATG Met	CCC Pro 90	GAA Glu	GGC	TAC Tyr	GTC Val	CAG Gln 95	GAG Glu	288
CGC Arg	ACC Thr	ATC	TTC Phe 100	Phe	AAG Lys	GAC Asp	GAC Asp	GGC Gly 105	Asn	TAC	AAG Lys	ACC Thr	CGC Arg 110	Ala	GAG Glu	336
GTG Val	AAG Lys	TTC Phe	e Glu	GGC Gly	GAC Asp	ACC Thr	CTG Leu 120	Val	AAC Asn	CGC Arg	: ATC	GAG Glu 125	Lev	AAC Lys	GGC Gly	384
ATC Ile	GAC Asp 130	Phe	AAG Lys	GAG Glu	GAC Asp	GGC Gly 135	Asr	ATC	CTG Lev	GGG GI	CAC His	Lys	CTC Lev	GAC 1 Glu	TAC Tyr	432
AAC Asr 145	туг	AAC Ası	AGC N Ser	CAC His	AAC Asr 150	Val	TAT	T ATC	ATC Met	GCC : Ala 155	a Asp	AAC Lys	G CAC	AAC Lys	AAC ASN 160	480
GG(Gl _y	C ATO	C AAG ≥ Lys	G GTY	AAC L Asr 165	n Phe	AAC Lys	ATC	C CGC	C CAC g His 170	. Asr	TATO	C GA0 ≥ Glu	G GAG	G GGG G G1:	AGC y Ser	528
GT(Va:	G CAC	G CTO	C GCC u Ala 180	a Asp	CAC His	TAC 5 Tyı	CAG	G CAC n Glr 185	n Ası	C ACC	c ccc	TA C	C GG e Gl; 19	y As	c GGC p Gly	576
CC(Pro	C GTY o Vai	G CT l Le	u Lei	G CCC	C GAG	C AAG p Asi	CA h Hi 20	s Ty:	C CTV	G AG u Se	C ACC	C CA r Gl: 20	n Se	c GC r Al	C CTG a Leu	624
AG Se	C AA r Ly: 21	s As	C CC	C AAG	C GA	G AAG u Ly: 21!	s Ar	C GA' g Asi	T CA p Hi	C AT s Me	G GT t Va 22	l Le	G CT u Le	G GA u Gl	G TTC u Phe	672
GT	G AC	c GC	c GC	c gg	G AT	C AC'	г ст	C GG	C AT	g ga	C GA	G CT	G TA	C AA	G TCC	720

Val Thr A	la Ala Gly	Ile Thr 230	Leu Gly	Met Asp 235	Glu Leu	Tyr Lys	Ser 240
	GA TCT CGA rg Ser Arg 245	Ala Met					
	GG GAC GCC ly Asp Ala 260						
Glu Leu A	AC TTC TCC sp Phe Ser 75						
	AG CCG AAT lu Pro Asn						
	CC GAT GAT ro Asp Asp						
	GT CTC TCT er Leu Ser 325	Gly Glu					Asp
	GG CCG CAG ly Pro Gln 340						
Ser Gly L	TG AGC CCT eu Ser Pro 55						
	TG GGG CCC al Gly Pro						
	CC CTG GCC ro Leu Ala						
	GC TTC GAG ly Phe Glu 405	Gly Tyr					Ala
	GC TCC TCT ly Ser Ser 420						
Thr Ser P	CC TGC GTC ro Cys Val 35						
	TT CAA AAC he Gln Asn						

ATA I Ile I 465	ATG Met	TCA Ser	CCT Pro	CGA Arg	ACC Thr 470	AGC Ser	CTC Leu	GCC Ala	Glu	GAC Asp 475	AGC Ser	TGC Cys	CTG Leu	GGC Gly	CGC Arg 480	1440
CAC His	TCG Ser	CCC Pro	GTG Val	CCC Pro 485	CGT Arg	CCG Pro	GCC Ala	TCC Ser	CGC Arg 490	TCC Ser	TCA Ser	TCG Ser	CCT Pro	GGT Gly 495	GCC Ala	1488
AAG Lys	CGG Arg	AGG Arg	CAT His 500	TCG Ser	TGC Cys	GCC Ala	GAG Glu	GCC Ala 505	TTG Leu	GTT Val	GCC Ala	CT G Leu	CCG Pro 510	CCC Pro	GGA Gly	1536
GCC Ala	TCA Ser	CCC Pro 515	CAG Gln	CGC Arg	TCC Ser	CGG Arg	AGC Ser 520	CCC Pro	TCG Ser	CCG Pro	CAG Gln	CCC Pro 525	TCA Ser	TCT Ser	CAC His	1584
GTG Val	GCA Ala 530	CCC Pro	CAG Gln	GAC Asp	CAC His	GGC Gly 535	TCC Ser	CCG Pro	GCT Ala	GGG Gly	ТАС Туг 540	CCC Pro	CCT Pro	GTG Val	GCT Ala	1632
GGC Gly 545	TCT Ser	GCC Ala	GTG Val	ATC Ile	ATG Met 550	GAT Asp	GCC Ala	CTG Leu	AAC Asn	AGC Ser 555	CTC Leu	GCC Ala	ACG Thr	GAC Asp	TCG Ser 560	1680
CCT Pro	TGT Cys	GGG Gly	ATC	CCC Pro 565	Pro	AAG Lys	ATG Met	TGG	AAG Lys 570	Thr	AGC Ser	CCT Pro	GAC Asp	CCC Pro 575	TCG Ser	1728
CCG Pro	GTG Val	TCT Ser	GCC Ala 580	Ala	CCA Pro	TCC Ser	AAC Lys	GCC Ala 585	Gly	Len CIG	CCT Pro	CGC Arg	CAC His	Ile	TAC Tyr	1776
CCG Pro	GCC	GTC Val 595	Glu	TTC Phe	CTG Leu	GGG Gly	Pro 600	CA:	GAG Glu	CAG Gln	GGC G13	GAC Glu 605	Arg	AG/ Arg	A AAC g Asn	1824
TCG Ser	GCT Ala 610	Pro	GAA Glu	TCC Sei	ATC	CTC Lev 615	ı Lei	G GT	r CCG	CCC Pro	C ACT	Tr	CCC Pro	AA(G CCG S Pro	1872
CTG Leu 625	(Val	CC'	r GCC	AT.	r ccc e Pro 630	lle	TG(C AGG S Se:	T Ile	C CCA Pro 635	o Vai	G ACT	r GC	A TC	C CTC r Leu 640	1920
CCI Pro	CCA Pro	A CT	T GAG	G TGG u Trj 64	p Pro	G CTO	TCC L Se:	C AG r Se	T CAC r Glr 650	n Se	A GG r Gl	C TC' y Se:	T TA	C GA r G1 65	G CTG u Leu 5	1968
CGC Arç	TATO	C GA	G GTY u Va:	1 G1	G CCO	C AAG o Ly:	G CC. s Pr	A CA O Hi 66	s Hi	c cg	g GC g Al	C CA a Hi	С ТА s Ту 67	r Gl	G ACA u Thr	2016
GA/ Glu	A GGG	C AG y Se 67	r Ar	A GG g Gl	g gc y Al	T GTV a Va	C AA 1 Ly 68	s Al	T CC. a Pr	A AC o Th	T GG r Gl	A GG y Gl 68	у Ні	C CC s Pr	T GTG	2064
GT.	r ca	G CT	C CA	T GG	с та	C AT	g ga	AA AL	C AA	.G CC	T CI	rg gg	A CI	T CA	G ATC	2112

Val	Gln 690	Leu	His	Gly		Met (695	Glu	Asn	Lys	Pro	Leu 700	Gly	Leu	Gln	Ile	
TTC Phe 705	ATT Ile	GGG Gly	ACA Thr	GCT Ala	GAT Asp 710	GAG Glu	CGG Arg	ATC Ile	CTT Leu	AAG Lys 715	CCG Pro	CAC His	GCC Ala	TTC Phe	TAC Tyr 720	2160
CAG Gln	GTG Val	CAC His	CGA Arg	ATC Ile 725	ACG Thr	GGG Gly	AAA Lys	ACT Thr	GTC Val 730	ACC Thr	ACC Thr	ACC Thr	AGC Ser	тат туг 735	GAG Glu	2208
AAG Lys	ATA Ile	GTG Val	GGC Gly 740	AAC Asn	ACC Thr	AAA Lys	GTC Val	CTG Leu 745	GAG Glu	ATC Ile	CCC Pro	TTG Leu	GAG Glu 750	CCC Pro	AAA Lys	2256
AAC Asn	AAC Asn	ATG Met 755	Arg	GCA Ala	ACC Thr	ATC Ile	GAC Asp 760	TGT Cys	GCG Ala	GGG Gly	ATC	TTG Leu 765	Lys	CTT Leu	AGA Arg	2304
AAC Asn	GCC Ala 770	GAC Asp	ATT Ile	GAG Glu	CTG Leu	CGG Arg 775	AAA Lys	GGC Gly	GAG Glu	ACG Thr	GAC Asp 780	Ile	GGA Gly	AGA Arg	AAG Lys	2352
AAC Asn 785	Thr	CGG Arg	GTG Val	AGA Arg	CTG Leu 790	GTT Val	TTC Phe	CGA Arg	GTT Val	CAC His	; 11e	CCA Pro	GAG Glu	TCC Ser	Ser 800	2400
GG(Gly	AGA Arg	ATC	GTC Val	TCT Ser 805	Leu	CAG Gln	ACT Thr	GCA Ala	TCT Ser 810	Ası	C CCC	TATO	C GAC ∈ Glv	TG0 1 Cys 815	TCC S Ser	2448
CAC Gl:	G CGA	TCT Sei	C GCT Ala 820	a His	GAG Glu	CTG Leu	CCC	ATO Met 825	. Val	GAI Glu	A AGA	A CAI g Gli	A GAG n Ası 830	o Thi	A GAC r Asp	2496
AG(Se:	TGC Cys	CTC Lev 83!	. Va	TAT l Tyr	GGC Gly	GGC Gly	CAC Glr 840	ı Glr	YFA A	ATO	C CTO	C ACC u Th	r Gl	G CA	G AAC n Asn	2544
TT Ph	r ACA e Thi 850	s Se	C GAG	G TCC	AAA Lys	GTI Val 855	. Va	G TT	r AC' e Thi	r GA	G AA u Ly 86	s Th	C AC r Th	A GA r As	T GGA p Gly	2592
CA G1 86	n Gl	A AT	T TG	G GAG p Glu	ATC Met 870	: Glu	A GC0	C ACC	G GTG r Va	G GA 1 As 87	p Ly	G GA s As	C AA p Ly	G AG s Se	c CAG r Gln 880	2640
CC Pr	C AAG o Asi	C AT	G CT t Le	T TT u Ph	e Vai	r GAC l Glu	G AT	C CC e Pr	T GA o G1 89	и Ту	T CG	G AA g As	C AA	G CA rs Hi 89	T ATC s Ile	2688
CG Ar	C AC.	A CC r Pr	T GT o Va 90	l Ly	A GTO	G AAG l Ası	TT n Ph	С ТА е Ту 90	r Va	C AT 1 II	C AA e As	T GC	G A. Ly Ly 91	's Ai	GA AAA ng Lys	2736
CG Ar	GA AG g Se	T CA r Gl 91	n Pr	T CA	G CA n Hi	C TT s Ph	T AC e Th 92	r Ty	C CA	C CC s Pi	CA G1	al Pi	CA GC co Al 25	CC A	TC AAG le Lys	2784

ACG (GAG Glu 930	CCC Pro	ACG Thr	GAT Asp	GAA Glu	тат Туг 935	GAC Asp	CCC Pro	ACT Thr	CTG Leu	ATC Ile 940	TGC Cys	AGC Ser	CCC Pro	ACC Thr	2832
CAT His 945	GGA Gly	GGC Gly	CTG Leu	GGG Gly	AGC Ser 950	CAG Gln	CCT Pro	TAC Tyr	TAC Tyr	CCC Pro 955	CAG Gln	CAC His	CCG Pro	ATG Met	GTG Val 960	2880
GCC Ala	GAG Glu	TCC Ser	CCC Pro	TCC Ser 965	TGC Cys	CTC Leu	GTG Val	GCC Ala	ACC Thr 970	ATG Met	GCT Ala	CCC Pro	TGC Cys	CAG Gln 975	CAG Gln	2928
TTC Phe	CGC Arg	ACG Thr	GGG Gly 980	CTC Leu	TCA Ser	TCC Ser	CCT Pro	GAC Asp 985	GCC Ala	CGC Arg	TAC Tyr	CAG Gln	CAA Gln 990	CAG Gln	AAC Asn	2976
CCA Pro	GCG Ala	GCC Ala 995	GTA Val	CTC Leu	TAC Tyr	Gln	CGG Arg 1000	Ser	AAG Lys	AGC Ser	Leu	AGC Ser 1005	CCC Pro	AGC Ser	CTG Leu	3024
Leu	GGC Gly 1010	тат туг	CAG Gln	CAG Gln	Pro	GCC Ala 1015	CTC Leu	ATG Met	GCC Ala	Ala	CCG Pro 1020	Leu	TCC Ser	CTT Leu	GCG Ala	3072
GAC Asp 1025	GCT Ala	CAC	CGC	TCT Ser	GTG Val	Leu	GTG Val	CAC His	GCC	GGC Gly 1035	Ser	CAG Gln	GGC Gly	CAC Glr	AGC Ser 1040	3120
TCA Ser	GCC Ala	CTG Leu	CTC	CAC His	Pro	TCT Ser	CCC	ACC Thr	AAC Asr 1050	Glr	CAG Glr	GCC Ala	TCC Ser	Pro 1055	r GTG Val	3168
ATC Ile	CAC	TAC	TCA Ser	Pro	ACC Thr	AAC Asn	CAC Glr	G CAG n Glr 1065	Lev	G CGC	TGC Cys	C GGA S Gly	AGC Sei 1070	His	C CAG s Gln	3216
GAG Glu	TTC	CAC Glr 1075	n His	TATO	C ATC	TAC	TG(Cys	s Glu	AAT ASI	r TTC	GCA Ala	A CCA a Pro 1089	Gl:	C ACC	C ACC r Thr	3264
Arg	CCT Pro 1090	Gly	CCC Pro	Pro	c ccc	val	. Se	T CAA	A GG' n Gly	r cac y Gl:	3 AGG 1 Arg 1100	g Lei	G AG	c cc r Pr	G GGT o Gly	3312
TCC Ser 1105	Typ	C CCC	C ACZ	A GTY	C AT 1 110	e Gl:	G CAG	G CA(G AA' n As:	T GCO n Al-	a Th	G AG r Se	C CA r Gl	A AG n Ar	A GCC g Ala 1120	3360
GCC Ala	: AAA	A AA(s Asi	c GGZ n Gl	A CC y Pr 112	o Pr	G GTY	C AG l Se	T GAG	C CA p Gl 113	n Ly	G GA s Gl	A GT u Va	A TT l Le	A CC u Pr 113	T GCG TO Ala	3408
GG(Gl _y	G GTV 7 Va:	G AC	C AT	e Ly	A CA s Gl	G GAG	G CA u Gl	G AA n As 114	n Le	G GA u As	C CA p Gl	G AC n Th	С ТА т Ту 115	r Le	G GAT	3456
GA7	r GT	AA T	T GA	A AT	T AT	C AG	G AA	.G GA	G TI	т тс	A GG	SA CC	T CC	T GO	CC AGA	3504

Asp Val Asn Glu Ile Ile Arg Lys Glu Phe Ser Gly Pro Pro Ala Arg 1155 1160 1165

CG TAA

AAT CAG ACG TAA Asn Gln Thr 1170

- (2) INFORMATION FOR SEQ ID NO:131:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1171 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:131:

	Val	Ser	Lys	Gly 5	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile 15	Leu
l Val	Glu	Leu	Asp 20		Asp	Val	Asn	Gly 25		Lys	Phe	Ser	Val 30	Ser	Gly
		35	Gly				40					45	Lys		
	50					55					60		Val		
65					70					75			His		80
				85					90				Val	95	
			100					105					Arg 110		
		115					120					125	Leu		
	130					135					140		Leu		
145					150					155					Asn 160
				165					170				Asp	175	
			180					185					190		Gly
		195					200					205			Leu
	210					215	1				220)			Phe
225					230)				235	5				Ser 240
				245	<u>,</u>				250)				255	
			260)				269	5				270)	Asp
		275	5				280)				289	5		Asn
Glu	Glu	Gli	ı Pro	Asr	n Ala	a His	Lys	s Val	l Ala	a Se	r Pro	o Pro	o Ser	: G1)	/ Pro

3516

	290					295					300				
305	Tyr				310		Asp '			315					320
Leu				325			Pro		330					335	
			340					345					350		
		355					Glu 360					365			
	370					375	Met				380				
385					390		Ala			395					400
				405			Arg		410					415	
			420				Phe	425					430		
		435					Asn 440					445			
	450					455	Ala				460				
465					470		Leu			475					480
				485			Ala		490					495	
			500				Glu	505					510		
		515	,				Ser 520					525			
	530					535					540				Ala
545	,				550		Ala			555	5				560
				565					570)				575	
			580)				585)				590)	Tyr
		595	5				600					605)		Asn
	610)				615	•				620)			Pro
625	5				630)				63	5				640
				645	5				65	0				655	
			660)				669	5				670)	1 Thr
		67	5				680)				689	5		Val
	69	0				695	5				70	C			n Ile
70	5				710)				71	5				720
				72	5				73	0				73	_
			74	0				74	5				75	0	o Lys
As	n As	n Me	t Ar	g Al	a Th	r Il	e Asp	o CA	s Al	a Gl	y Il	e Le	u Ly	s Le	u Arg

		755					760					765			
	770					775				Thr	780				
785					790					His 795					008
				805					810	Asn				815	
			820					825		Glu			830		
		835					840			Ile		845			
	850					855				Glu	860				
865					870					Asp 875					880
				885					890					895	
			900					905		Ile			910		
		915					920			Pro		925			
	930					935				Leu	940				
945					950					955					Val 960
				965					970					975	
			930					985					990	1	Asn
		995					1000					1005	l .		Leu
	1010					1015					1020				Ala
025					1030	1				1035	1				1040
				1045					1050)				105	
			1060)				1065	5				1070)	Gln
		1075	5				1080)				1089	5		r Thr
	1090)				1095	5				1100)			o Gly
105	,				1110)				1119	5				g Ala 1120
				1125	5				113	0				113	
			1140)				114	5				115	O	u Asp
		115	5	ı Ile	e Ile	e Ar	116		u Ph	e Se:	r Gly	y Pr 116	o Pr 5	o al	a Arg
Asr	Gl: 1170	n Thi	r												

(2) INFORMATION FOR SEQ ID NO:132:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3546 base pairs

(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence

(B) LOCATION: 1...3543

(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:132:

Met	AAC Asn	GCC Ala	CCC Pro	GAG Glu 5	CGG Arg	CAG Gln	CCC Pro	CAA Gln	CCC Pro	GAC Asp	GGC Gly	GGG Gly	GAC Asp	GCC Ala 15	CCA Pro	48
1 GGC Gly	CAC His	GAG Glu	CCT Pro 20	GGG	GGC Gly	AGC Ser	CCC Pro	CAA Gln 25	GAC Asp	GAG Glu	CTT Leu	GAC Asp	TTC Phe 30	TCC Ser	ATC Ile	96
CTC Leu	TTC Phe	GAC Asp 35	TAT Tyr	GAG Glu	TAT Tyr	TTG Leu	AAT Asn 40	CCG Pro	AAC Asn	GAA Glu	GAA Glu	GAG Glu 45	CCG Pro	AAT Asn	GCA Ala	144
CAT His	AAG Lys 50	GTC Val	GCC Ala	AGC Ser	CCA Pro	CCC Pro 55	TCC Ser	GGA Gly	CCC Pro	GCA Ala	TAC Tyr 60	CCC Pro	GAT Asp	GAT Asp	GTA Val	192
ATG Met 65	GAC Asp	TAT Tyr	GGC Gly	CTC Leu	AAG Lys 70	CCA Pro	TAC Tyr	AGC Ser	CCC Pro	CTT Leu 75	GCT Ala	AGT Ser	CTC	TCT Ser	GGC Gly 80	240
GAG Glu	CCC Pro	CCC	GGC Gly	CGA Arg 85	TTC Phe	GGA Gly	GAG Glu	CCG Pro	GAT Asp 90	AGG Arg	GTA Val	GGG Gly	CCG Pro	CAG Gln 95	AAG Lys	288
TTT Phe	CTG Leu	AGC Ser	GCG Ala 100	Ala	AAG Lys	CCA Pro	GCA Ala	GGG Gly 105	GCC Ala	TCG Ser	GGC Gly	CTG Leu	AGC Ser 110	Pro	CGG	336
ATC Ile	GAG Glu	ATC Ile 115	Thr	CCG Pro	TCC Ser	CAC His	GAA Glu 120	Lev	ATC	CAG Glr	GCA Ala	GTG Val 125	GJ?	CCC Pro	CTC Leu	384
CGC Arg	ATG Met	Arg	GAC Asp	GCG Ala	GGC Gly	CTC Leu 135	Leu	GTC Val	GAC Glu	CAC Glr	CCT Pro 140	Pro	CTC Lev	GCC 1 Ala	GGG Gly	432
GTG Val	Ala	GCC Ala	: AGC	CCG Pro	AGG Arg 150	Phe	ACC Thr	CTC	G CCC	GTC Val 155	l Pro	GIY	TTY Phe	C GAC	GGC Gly 160	480
TAC Tyr	CGC	GAC GGL	CCC Pro	CTI Lev 165	т СХ.	TTG Leu	AGC Sei	C CCC	C GC:	a Ser	C AGO	GGG Gly	TCC Se:	r Sei	r GCC c Ala	528
AGO	TT	C ATC	r TC	r GAC	ACC	TTC	TCC	C CC	C TAC	C AC	C TC	G CC	TG	CGT	C TCG	576

Ser	Phe	Ile	Ser 180	Asp	Thr	Phe	Ser	Pro 185	T/r	Thr	Ser	Pro	Cys 190	Val	Ser	
					CCC Pro											624
	_				CCC Pro											672
		_	_		AGC Ser 230											720
	_				TCA Ser											768
					GCC Ala											816
					CAG Gln											864
					TAC Tyr											912
					CTC Leu 310											960
					AGC Ser											1008
		_	_		CCT Pro											1056
					GGC Gly											1104
					ACT Thr											1152
					GTG Val 390											1200
					GGC Gly											1248

AAG Lys	CCA Pro	CAT His	CAC His 420	CGG Arg	GCC Ala	CAC His	TAT Tyr	GAG Glu 425	ACA Thr	GAA Glu	GGC Gly	AGC Ser	CGA Arg 430	GGG Gly	GCT Ala	1296
GTC Val	AAA Lys	GCT Ala 435	CCA Pro	ACT Thr	GGA Gly	Gly	CAC His 440	CCT Pro	GTG Val	GTT Val	CAG Gln	CTC Leu 445	CAT His	GGC Gly	TAC Tyr	1344
ATG Met	GAA Glu 450	AAC Asn	AAG Lys	CCT Pro	CTG Leu	GGA Gly 455	CTT Leu	CAG Gln	ATC Ile	TTC Phe	ATT Ile 460	GGG Gly	ACA Thr	GCT Ala	GAT Asp	1392
GAG Glu 465	CGG Arg	ATC Ile	CTT Leu	AAG Lys	CCG Pro 470	CAC His	GCC Ala	TTC Phe	TAC Tyr	CAG Gln 475	GTG Val	CAC His	CGA Arg	ATC Ile	ACG Thr 480	1440
GGG Gly	AAA Lys	ACT Thr	GTC Val	ACC Thr 485	ACC Thr	ACC Thr	AGC Ser	TAT Tyr	GAG Glu 490	AAG Lys	ATA Ile	GTG Val	GGC	AAC Asn 495	ACC Thr	1488
AAA Lys	GTC Val	CTG Leu	GAG Glu 500	ATC Ile	CCC Pro	TTG Leu	GAG Glu	CCC Pro 505	AAA Lys	AAC Asn	AAC Asn	ATG Met	AGG Arg 510	GCA Ala	ACC Thr	1536
ATC Ile	GAC Asp	TGT Cys 515	Ala	GGG Gly	ATC	TTG Leu	AAG Lys 520	Leu	AGA Arg	AAC Asn	GCC Ala	GAC Asp 525	ATT	GAG Glu	CTG Leu	1584
CGG	AAA Lys 530	Gly	GAG Glu	ACG Thr	GAC Asp	ATT Ile 535	GG.A Gly	AGA Arg	AAG Lys	AAC Asn	ACG Thr 540	Arg	GTG Val	AGA Arg	CTG	1632
GTT Val 545	Phe	CGA Arg	GTT Val	CAC His	ATC Ile 550	Pro	GAC Glu	TCC Ser	AGT Ser	GGC Gly 555	Arg	ATC	GTC Val	TCT Ser	Leu 560	1680
					Pro					Glr					GAG Glu	1728
CTC Lev	CCC Pro	ATC Met	GTT Val 580	Glu	AGA Arg	CAA Gln	GAC Asi	C ACA Thi 585	Asp	AGC Ser	TGC Cys	CTY Lev	GT(1 Va: 590	l Tyr	GGC Gly	1776
G17 GG(CAC Glr	G CAA n Glr 595	n Net	F ATC	CTC	ACG Thr	GG(G1 ₂ 60)	y Glr	AAC n Asr	TTC Phe	T ACA ∋ Thi	A TCC	r Gl	G TCC	AAA Lys	1824
GT' Va	r GTC l Val 610	l Phe	r ACT	GAC	AAC 1 Lys	ACC Thr	Th	A GA' r Asi	r GG/ o Gly	A CAG	G CAM n Gli 620	n Il	r TG	G GAG p Gli	G ATG u Met	1872
GA: G1: 62:	u Ala	C ACC	G GTK r Val	G GA'	r AAG 5 Lys 630	s Asp	AA Ly	G AGG s Se	C CAG	G CCC n Pro 63	o Asi	C AT n Me	G CT t Le	T TT u Ph	T GTT e Val 640	1920
GA	G AT	C CC	T GA	A TA	r cg	S AAC	. AA	G CA	TA T	C CG	C AC	A CC	T GT	AA A	a gtg	1968

Glu	Ile	Pro	Glu	Tyr 645	Arg	Asn	Lys	His	Ile 650	Arg	Thr	Pro	Val	Lys 655	Val	
			_							CGA Arg						2016
										ACG Thr						2064
										CAT His						2112
										GCC Ala 715						2160
										TTC Phe						2208
			_							CCA Pro						2256
_										CTG Leu						2304
										GAC Asp						2352
										TCA Ser 795						2400
				_	_					ATC Ile						2448
										GAG Glu						2496
										AGA Arg						2544
										TCC Ser						2592
										GCC Ala 875						2640

GTC AGT GAC CA Val Ser Asp Gl	AA AAG GAA ln Lys Glu 885	GTA TTA Val Leu	CCT GCG Pro Ala 890	GGG GTG A	ACC ATT Thr Ile	AAA CAG Lys Gln 895	2688
GAG CAG AAC TY Glu Gln Asn Le 90	rg GAC CAG eu Asp Gln 00	ACC TAC Thr Tyr	TTG GAT Leu Asp 905	GAT GTT A	AAT GAA Asn Glu 910	ATT ATC Ile Ile	2736
AGG AAG GAG T Arg Lys Glu Pl 915	TT TCA GGA he Ser Gly	CCT CCT Pro Pro 920	GCC AGA Ala Arg	Asn Gln	ACG AGA Thr Arg 925	ATT CTG Ile Leu	2784
CAG TCG ACG G Gln Ser Thr V 930	TA CCG CGG al Pro Arg	GCC CGG Ala Arg 935	GAT CCA Asp Pro	CCG GTC Pro Val 940	GCC ACC Ala Thr	ATG GTG Met Val	2832
AGC AAG GGC G Ser Lys Gly G 945	AG GAG CTG lu Glu Leu 950	TTC ACC Phe Thr	GGG GTG Gly Val	GTG CCC Val Pro 955	ATC CTG Ile Leu	GTC GAG Val Glu 960	2880
CTG GAC GGC G Leu Asp Gly A	AC GTA AAC Asp Val Asn 965	GGC CAC Gly His	AAG TTC Lys Phe 970	Ser Val	TCC GGC Ser Gly	GAG GGC Glu Gly 975	2928
GAG GGC GAT G Glu Gly Asp A	GCC ACC TAC Ala Thr Tyr 980	GGC AAG Gly Lys	CTG ACC Leu Thr 985	CTG AAG Leu Lys	TTC ATC Phe Ile 990	Cys Thr	2976
ACC GGC AAG C Thr Gly Lys I 995	CTG CCC GTG Leu Pro Val	CCC TGG Pro Trp 1000	Pro Thr	Leu Val	ACC ACC Thr Thr 1005	CTG ACC	3024
TAC GGC GTG C Tyr Gly Val C 1010	CAG TGC TTC Gln Cys Phe	AGC CGC Ser Arg	TAC CCC	GAC CAC Asp His 1020	ATG AAG Met Lys	CAG CAC	3072
GAC TTC TTC A Asp Phe Phe I 1025	AAG TCC GCC Lys Ser Ala 1030	a Met Pro	GAA GGG Glu Gly	TAC GTC Y TYY Val 1035	CAG GAG	G CGC ACC Arg Thr 1040	3120
ATC TTC TTC I	AAG GAC GAG Lys Asp Asp 1045	GGC AAC Gly Asr	TAC AAG Tyr Lys 105	s Thr Arg	GCC GAC Ala Glu	G GTG AAG 1 Val Lys 1055	3168
TTC GAG GGC (Phe Glu Gly 1	GAC ACC CTO Asp Thr Let 060	G GTG AAC u Val Asr	C CGC ATO n Arg Il 1065	C GAG CTG e Glu Leu	AAG GGG Lys Gly 1070	y Ile Asp	3216
TTC AAG GAG Phe Lys Glu 1075	GAC GGC AA Asp Gly As	C ATC CTC n Ile Lev 1080	u Gly Hi	C AAG CTC s Lys Lei	GAG TAG 1 Glu Ty: 1085	C AAC TAC r Asn Tyr	3264
AAC AGC CAC Asn Ser His 1090	AAC GTC TA Asn Val Ty	T ATC ATC r Ile Me 1095	G GCC GA t Ala As	C AAG CAC p Lys Glr 1100	n Lys As	C GGC ATC n Gly Ile	3312
AAG GTG AAC	TTC AAG AT	C CGC CA	C AAC AT	C GAG GAG	C GGC AG	C GTG CAG	3360

Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln 1110 1115 1105 CTC GCC GAC CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC CCC GTG 3408 Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val 1125 1130 CTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG AGC AAA 3456 Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys 1145 1140 GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC GTG ACC 3504 Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr 1165 1160 1155 GCC GCC GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TAA 3546 Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 1180 1175 1170 (2) INFORMATION FOR SEQ ID NO:133: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1181 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (v) FRAGMENT TYPE: internal (xi) SEQUENCE DESCRIPTION: SEQ ID NO:133: Met Asn Ala Pro Glu Arg Gln Pro Gln Pro Asp Gly Gly Asp Ala Pro 10 1 5 Gly His Glu Pro Gly Gly Ser Pro Gln Asp Glu Leu Asp Phe Ser Ile 25 20 Leu Phe Asp Tyr Glu Tyr Leu Asn Pro Asn Glu Glu Glu Pro Asn Ala 40 His Lys Val Ala Ser Pro Pro Ser Gly Pro Ala Tyr Pro Asp Asp Val 55 60 Met Asp Tyr Gly Leu Lys Pro Tyr Ser Pro Leu Ala Ser Leu Ser Gly 70 75 80 Glu Pro Pro Gly Arg Phe Gly Glu Pro Asp Arg Val Gly Pro Gln Lys 90 85 Phe Leu Ser Ala Ala Lys Pro Ala Gly Ala Ser Gly Leu Ser Pro Arg 105 110 100 Ile Glu Ile Thr Pro Ser His Glu Leu Ile Gln Ala Val Gly Pro Leu 120 125 Arg Met Arg Asp Ala Gly Leu Leu Val Glu Gln Pro Pro Leu Ala Gly 140 130 135 Val Ala Ala Ser Pro Arg Phe Thr Leu Pro Val Pro Gly Phe Glu Gly 145 150 155 Tyr Arg Glu Pro Leu Cys Leu Ser Pro Ala Ser Ser Gly Ser Ser Ala 165 170 175 Ser Phe Ile Ser Asp Thr Phe Ser Pro Tyr Thr Ser Pro Cys Val Ser 180 185 190

Pro Asn Asn Gly Gly Pro Asp Asp Leu Cys Pro Gln Phe Gln Asn Ile

		195					200					205			
	210	His				215					Met 220				
Ser 225	Leu	Ala	Glu	Asp	Ser 230	Cys	Leu	Gly	Arg	His 235	Ser	Pro	Val	Pro	Arg 240
Pro	Ala	Ser	Arg	Ser 245	Ser	Ser	Pro	Gly	Ala 250	Lys	Arg	Arg	His	Ser 255	Cys
Ala	Glu	Ala	Leu 260	Val	Ala	Leu		Pro 265	Gly	Ala	Ser	Pro	Gln 270	Arg	Ser
		275					280				Ala	285			
	290					295					Ser 300				
305					310					315					320
				325					330		Val			335	
			340					345			Ala		350		
		355					360				Ala	365			
	370					375					Val 380				
Ile 385		Ser	Ile	Pro	Val 390	Thr	Ala	Ser	Leu	9rc 395	Pro	Leu	Glu	Trp	Pro 400
Leu	Ser	Ser	Gln	Ser 405	Gly	Ser	Tyr	Glu	Leu 410		, Ile	Glu	Val	Gln 415	Pro
Lys	Pro	His	His	Arg		His	Tyr	Glu 425		Glu	ı Gly	Ser	Arg 430		Ala
		435	,				440					445			Tyr
	450)				455					460	,			qzA ı
465	5				470					475	5				480
				485	i				490)				49	
			500)				505	5				510)	a Thr
		519	5				520					525	1		ı Leu
	530)				535	•				540)			g Leu
545	5				550)				55	5				r Leu 560
Gli	n Thi			565	5				57	0				57	
			580	5				58	5				590	0	r Gly
		59	5				600)				605	5		r Lys
	61	0				619	5				62	0			u Met
G1 62		a Th	r Va	l Ası	p Lys 630) Lys	s Se	r Gl	n Pr 63		n Me	t Le	u Ph	e Val 640
G1	u Il	e Pr	o Gl	u Ty:	r Arg		n Ly:	s Hi	s Il 65		g Th	r Pr	o Va	l Ly 65	s Val
As	n Ph	е Ту	r Va			n Gl	y Ly:	s Ar			g Se	r Gl	n Pr	o G1	n His

			660					665					670		
		675	His		Val		680					685			
	690					695					700				
705					Gln 710					715					720
				725	Ala				730					735	
			740		Tyr			745					750		
		755			Leu		760					765			
	770				Pro	775					780				
785					Ser 790					795					800
				805	Gln				810					815	
			820		Суѕ			825					830	1	
-		835					840					845			Pro
	850					855					860)			Ile
		Glr	ı Asn	Ala	Thr 870	Ser	Gin	Arg	Alc	875		ASI	(GI)	, 110	Pro 880
865 Val	Ser	Asp	Glr.	Lys 885	Glu	Val	Leu	Pro	Ala 890	a Gly		Thr	Ile	895	Gln
			900	Asp	Gln			905					910)	e Ile
		91	5				920					925	•		e Leu
	930)				935	,				94)			val
		Gl;	y Glu	ı Glu	1 Leu 950		Thr	Gly	/ Va	1 Va 95	l Pro	o Ile	e Le	u va.	1 Glu 960
945 Leu	ASI	o Gl	y Asi	o Val 969	Asr		His	Lys	97	e Se		l Sei	c Gl	y Gl	a Gly
			986	a Thi	туг			98	5				99	U	s Thr
		99	s Le	u Pro			1000)				100	5		u Thr
	101	n				1015	5				102	0			n His
029	5				1030)				103	5				g Thr 1040
				104	5				105	0				105	
			106	0				106	5				107	0	e Asp
		107	5				108	0				108	5		n Tyr
	109	0				109	5				110	00			y Ile al Gln
10	5				111	0				111	15				al Gln 1120
Le	u Al	a As	sp Hi	з Ту	r Gl	n Gl	n As	n Th	ır Pı	0 I	le G	ly As	sp G	ly Pi	co Val

1135 1125 1130 Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys 1140 1145 1150 Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr 1155 1160 1165 Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 1175 1170 (2) INFORMATION FOR SEQ ID NO:134: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2802 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (ix) FEATURE: (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...2799 (D) OTHER INFORMATION: (xi) SEQUENCE DESCRIPTION: SEQ ID NO:134: ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG 48 Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 5 GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 35 40 TGC ACC ACC GGC AAG CTG CCC GTG CCC TGG CCC ACC CTC GTG ACC ACC 192 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 50 55 CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG 240 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 75 65 CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG 288 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 CGC ACC ATC TTC TTC AAG GAC GAC GGC AAC TAC AAG ACC CGC GCC GAG Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 105 GTG AAG TTC GAG GGC GAC ACC CTG GTG AAC CGC ATC GAG CTG AAG GGC 384 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 120 ATC GAC TTC AAG GAG GAC GGC AAC ATC CTG GGG CAC AAG CTG GAG TAC 432

Ile	Asp 130	Phe	Lys	Glu	Asp	Gly 135	Asn	Ile	Leu	Gly	His 140	Lys	Leu	Glu	Tyr	
AAC Asn 145	TAC Tyr	AAC Asn	AGC Ser	CAC His	AAC Asn 150	GTC Val	TAT Tyr	ATC Ile	ATG Met	GCC Ala 155	GAC Asp	AAG Lys	CAG Gln	AAG Lys	AAC Asn 160	480
GGC Gly	ATC Ile	AAG Lys	GTG Val	AAC Asn 165	TTC Phe	AAG Lys	ATC Ile	CGC Arg	CAC His 170	AAC Asn	ATC Ile	GAG Glu	GAC Asp	GGC Gly 175	AGC Ser	528
GTG Val	CAG Gln	CTC Leu	GCC Ala 180	GAC Asp	CAC His	TAC Tyr	CAG Gln	CAG Gln 185	AAC Asn	ACC Thr	CCC Pro	ATC Ile	GGC Gly 190	GAC Asp	GGC Gly	576
CCC Pro	GTG Val	CTG Leu 195	CTG Leu	CCC	GAC Asp	AAC Asn	CAC His 200	TAC Tyr	CTG Leu	AGC Ser	ACC Thr	CAG Gln 205	TCC Ser	GCC Ala	CTG Leu	624
AGC Ser	AAA Lys 210	GAC Asp	CCC	AAC Asn	GAG Glu	AAG Lys 215	CGC Arg	GAT Asp	CAC His	ATG Met	GTC Val 220	Leu	CTG Leu	GAG Glu	TTC Phe	672
GTG Val 225	ACC Thr	GCC Ala	GCC Ala	GGG Gly	ATC Ile 230	ACT Thr	CTC Leu	GGC Gly	ATG Met	GAC Asp 235	Glu	CTG Leu	TAC Tyr	AAG Lys	TCC Ser 240	720
GGA Gly	CTC Leu	AGA Arg	TCT Ser	CGA Arg 245	Gly	AGC Ser	ATG Met	GGC Gly	ACC Thr 250	Leu	CGG Arg	GAT Asp	TTA Leu	CAG Gln 255	TAC	768
GCG Ala	CTC Leu	CAG Gln	GAG Glu 260	Lys	ATC Ile	GAG Glu	GAG Glu	CTG Leu 265	AGG Arg	CAC Glr	CGG Arg	GAT Asp	GCT Ala 270	Leu	ATC Ile	816
GAC Asp	GAG Glu	CTC Lev 275	Glu	CTC Leu	GAG Glu	TTG Leu	GAT Asp 280	Gln	AA G	GAC Asp	GAA Glu	Lev 285	ı Ile	CAC Glr	AAG Lys	864
CTG Leu	CAG Gln 290	Asr	GAG Glu	CTC Lev	GAC Asp	AAG Lys 295	TAC Tyr	CGC Arg	TCC	GTC Val	300	e Arç	A CCA	A GCC	ACC Thr	912
CAG Gln 305	Glr	GCC Ala	G CAC	AAC Lys	G CAG Glr 310	Ser	GCG	AGC Ser	ACC Thi	Lev 315	ı Glı	G GGC	C GAG	CCC Pro	G CGC D Arg 320	960
ACC Thr	: AAC	G CGG	G CAC	G GC0 1 Ala 325	a Ile	TCC Ser	GCC Ala	GAC Glu	CCC Pro	Th:	C GCC	C TT	C GAG	2 ATG 2 Il 33	C CAG e Gln 5	1008
GAT Asp	CTC Lev	AG(1 Se:	C CAT r His 340	s Va	G ACC	CTC	CCC Pro	TTC Phe 345	Ty:	C CC	C AA	G AG s Se	c cc r Pr 35	o Gl	G TCC n Ser	1056
AAC Lys	GAS	r CT D Le	u Il	A AAG e Ly:	G GAZ	A GCT	TATO 11e 360	e Le	GA(C AA p As	T GA n As	C TT p Ph 36	e Me	G AA t Ly	G AAC s Asn	1104

									GTG Val							1152
									AAA Lys							1200
									GTT Val 410							1248
									AAA Lys							1296
									ACC Thr							1344
									TGT Cys							1392
	Thr								TAT Tyr							1440
									GAG Glu 490							1488
									Asn					Ile	AGG Arg	1536
			Arg					Phe					Gly		GTA Val	1584
		Thr					Pro					Val			AGA Arg	1632
ACT Thr 545	Leu	GGA Gly	AAA Lys	GGA Gly	GAC Asp 550	Trp	TTI Phe	GGA Gly	GAG Glu	AAA Lys 555	: Ala	TTC	G CAC	G GG(n Gly	G GAA / Glu 560	1680
GAT Asp	GTO Val	AGA Arg	ACA Thr	GCA Ala 565	Asn	GTA Val	TTA .	GCT Ala	GCA Ala 570	Gli	A GCT 1 Ala	r GTA a Val	A ACC	TGG r Cy: 57!	CTT s Leu 5	1728
				Asp					Leu					u As	r GAT p Asp	1776
GT:	r TCI	r aa t	AAA :	GC#	LAT A	GAA	GAT	r GCA	A GAA	A GC	r aa	A GC	AA A	A TA	T GAA	1824

Va1	Ser	Asn 595	Lys	Ala	Tyr	Glu	Asp 600	Ala	Glu	Ala	Lys	Ala 605	Lys	Tyr	Glu	
GCT Ala	GAA Glu 610	GCG Ala	GCT Ala	TTC Phe	TTC Phe	GCC Ala 615	AAC Asn	CTG Leu	AAG Lys	CTG Leu	TCT Ser 620	GAT Asp	TTC Phe	AAC Asn	ATC Ile	1872
ATT Ile 625	GAT Asp	ACC Thr	CTT Leu	GGA Gly	GTT Val 630	GGA Gly	GGT Gly	TTC Phe	GGA Gly	CGA Arg 635	GTA Val	GAA Glu	CTG Leu	GTC Val	CAG Gln 640	1920
TTG Leu	AAA Lys	AGT Ser	GAA Glu	GAA Glu 645	TCC Ser	AAA Lys	ACG Thr	TTT Phe	GCA Ala 650	ATG Met	AAG Lys	ATT Ile	CTC Leu	AAG Lys 655	AAA Lys	1968
CGT Arg	CAC His	ATT	GTG Val 660	GAC Asp	ACA Thr	AGA Arg	CAG Gln	CAG Gln 665	GAG Glu	CAC	ATC	CGC Arg	TCA Ser 670	Glu	AAG Lys	2016
CAG Gln	ATC Ile	ATG Met 675	Gln	GGG Gly	GCT Ala	CAT His	TCC Ser 680	GAT Asp	TTC Phe	ATA Ile	GTG Val	AGA Arg 685	Leu	TAC Tyr	AGA Arg	2064
ACA Thr	TTT Phe 690	Lys	GAC Asp	AGC Ser	AAA Lys	ТАТ Туг 695	TTG Leu	TAT Tyr	ATG Met	TTC Lev	ATC Met 700	: Glu	GCT Ala	TGI Cys	CTA Leu	2112
GGT Gly 705	Gly	GAC Glu	CTC	TGG Trp	ACC Thr 710	ATT	CTC Leu	AGG Arg	GAT Asp	AGA Arg 715	g Gly	r TCC / Ser	TTI Phe	GAÆ Glu	A GAT Asp 720	2160
TC1 Ser	ACA Thr	A ACC	C AGA	A TTI g Phe 725	Tyr	ACA Thr	GCA Ala	. TGI Cys	GTG Val	. Va	A GAI	A GC:	r TT:	r GC0 ≥ Ala 735	TAT Tyr	2208
CT(Let	G CAT	r TC(2 AA r Lys 740	s Gly	ATC	ATI	TAC Tyr	AGC Arc 745	, Asr	C CTO	C AA	G CC	A GAZ O Gli 75	ı Ası	r CTC	2256
ATC Ile	CTA E Lev	A GA' L As 75	p Hi	C CGA	GGT Gly	тАТ / Туз	GCC Ala 760	Lys	A CTO	G GT u Va	T GA l As	T TT p Ph	e Gl	C TT y Ph	T GCA e Ala	2304
AA(Ly:	G AA s Ly: 77	s Il	A GG e Gl	A TT y Phe	r GGZ e Gly	A AAG 7 Ly: 77!	s Lys	A AC	A TGO	G AC p Th	T TT I Ph 78	е Су	T GG s Gl	G AC y Th	T CCA r Pro	2352
GA G1 78	и Ту	T GT r Va	A GC 1 Al	C CC a Pr	A GA(o Gl) 79	ı Il	c ATG	C CTV	G AA u As	C AA n Ly 79	s Gl	C CA y Hi	T GA s As	C AT	T TCA e Ser 800	2400
GC Al	C GA a As	С ТА Р Ту	C TG	G TC p Se 80	r Le	G GG. u Gl	A ATO	C CT e Le	A AT u Me 81	t Ty	AT G# Ar Gl	A CI lu Le	C CI u Le	G AC eu Th 81	T GGC ir Gly	2448
AG Se	c cc r Pr	A CC	TT TT O Ph	ie Se	A GG r Gl	c cc y Pr	A GA o As	T CC p Pr 82	o Me	G AA	AA AC /s Tì	CC TA	r As	AC AT sn II	CC ATA le Ile	2496

TTG Leu	AGG Arg	GGG Gly 835	ATT Ile	GAC Asp	ATG Met	ATA Ile	GAA Glu 840	TTT Phe	CCA Pro	AAG Lys	AAG Lys	ATT Ile 845	GCC Ala	AAA Lys	AAT Asn	2544
GCT Ala	GCT Ala 850	AAT Asn	TTA Leu	ATT Ile	AAA Lys	AAA Lys 855	CTA Leu	TGC Cys	AGG Arg	GAC Asp	AAT Asn 860	CCA Pro	TCA Ser	GAA Glu	AGA Arg	2592
TTA Leu 865	GGG Gly	AAT Asn	TTG Leu	AAA Lys	AAT Asn 870	GGA Gly	GTA Val	AAA Lys	GAC Asp	ATT Ile 875	CAA Gln	AAG Lys	CAC His	AAA Lys	TGG Trp 880	2640
TTT Phe	GAG Glu	GGC Gly	TTT Phe	AAC Asn 885	TGG Trp	GAA Glu	GGC Gly	TTA Leu	AGA Arg 890	AAA Lys	GGT Gly	ACC Thr	TTG Leu	ACA Thr 895	CCT Pro	2688
CCT Pro	ATA Ile	ATA Ile	CCA Pro 900	AGT Ser	GTT Val	GCA Ala	TCA Ser	CCC Pro 905	ACA Thr	GAC Asp	ACA Thr	AGT Ser	AAT Asn 910	Phe	GAC Asp	2736
AGT Ser	TTC Phe	CCT Pro 915	Glu	GAC Asp	AAC Asn	GAT Asp	GAA Glu 920	Pro	CCA Pro	CCT	GAT Asp	GAC Asp 925	Asn	TCA Ser	GGA Gly	2784
	GAT Asp 930	Ile			TAA											2802

(2) INFORMATION FOR SEQ ID NO:135:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 933 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:135:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 45 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 60 55 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 75 70 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 90 85 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 105 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly

		115					120					125			
Ile A			Lys	Glu				Ile	Leu	Gly	His 140	Lys	Leu	Glu	Tyr
Asn 1	Tyr	Asn	Ser	His	Asn 150	Val	Tyr	Ile	Met	Ala 155	Asp	Lys	Gln	Lys	Asn 160
Gly :	Ile	Lys	Val	Asn 165	Phe	Lys	Ile	Arg	His 170	Asn	Ile	Glu	Asp	Gly 175	Ser
Val (180					185					190		
		195					His 200					205			
	210					215	Arg				220				
225					230		Leu			235					240
-				245			Met		250					255	
			260				Glu	265					270		
		275					Asp 280					285			
	290					295	Tyr				300				
305					310		Ala			315					320
				325			Ala		330					335	
			340				Pro	345					350		
_		355					Ile 360					365			
	370					375					380				
385					390		Ile			395					400
				405	,				410)				415	
			420	}				425					430	}	Ala
		435					440					445	•		Asn
	450					455	•				460)			Met
465					470)				475	5				480
				485	5				490)				499	
			500)				505	5				510)	Arg
		515	5				520)				52	5		val
	530)				535	5				540)			Arg
545					550	0				55	5				y Glu 560
				56	5				57	0				57	
Val	Ile	e Ası	o Ar	g As	p Se	r Phe	e Lys	s His	s Le	u Il	e Gl	y Gl	y Le	u As	p Asp

585 580 Val Ser Asn Lys Ala Tyr Glu Asp Ala Glu Ala Lys Ala Lys Tyr Glu 600 Ala Glu Ala Ala Phe Phe Ala Asn Leu Lys Leu Ser Asp Phe Asn Ile 620 615 Ile Asp Thr Leu Gly Val Gly Gly Phe Gly Arg Val Glu Leu Val Gln 635 630 Leu Lys Ser Glu Glu Ser Lys Thr Phe Ala Met Lys Ile Leu Lys Lys 650 645 Arg His Ile Val Asp Thr Arg Gln Gln Glu His Ile Arg Ser Glu Lys 665 660 Gln Ile Met Gln Gly Ala His Ser Asp Phe Ile Val Arg Leu Tyr Arg 680 Thr Phe Lys Asp Ser Lys Tyr Leu Tyr Met Leu Met Glu Ala Cys Leu 700 695 Gly Gly Glu Leu Trp Thr Ile Leu Arg Asp Arg Gly Ser Phe Glu Asp 715 710 Ser Thr Thr Arg Phe Tyr Thr Ala Cys Val Val Glu Ala Phe Ala Tyr 730 725 Leu His Ser Lys Gly Ile Ile Tyr Arg Asp Leu Lys Pro Glu Asn Leu 745 740 Ile Leu Asp His Arg Gly Tyr Ala Lys Leu Val Asp Phe Gly Phe Ala 765 760 Lys Lys Ile Gly Phe Gly Lys Lys Thr Trp Thr Phe Cys Gly Thr Pro 780 775 Glu Tyr Val Ala Pro Glu Ile Ile Leu Asn Lys Gly His Asp Ile Ser 790 795 Ala Asp Tyr Trp Ser Leu Gly Ile Leu Met Tyr Glu Leu Leu Thr Gly 805 810 815 Ser Pro Pro Phe Ser Gly Pro Asp Pro Met Lys Thr Tyr Asn Ile Ile 820 825 Leu Arg Gly Ile Asp Met Ile Glu Phe Pro Lys Lys Ile Ala Lys Asn 835 840 Ala Ala Asn Leu Ile Lys Lys Leu Cys Arg Asp Asn Pro Ser Glu Arg 855 860 Leu Gly Asn Leu Lys Asn Gly Val Lys Asp Ile Gln Lys His Lys Trp 875 870 Phe Glu Gly Phe Asn Trp Glu Gly Leu Arg Lys Gly Thr Leu Thr Pro 890 895 Pro Ile Ile Pro Ser Val Ala Ser Pro Thr Asp Thr Ser Asn Phe Asp 900 905 Ser Phe Pro Glu Asp Asn Asp Glu Pro Pro Pro Asp Asp Asn Ser Gly 920 915 Trp Asp Ile Asp Phe

(2) INFORMATION FOR SEQ ID NO:136:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2799 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence

(B) LOCATION: 1...2795

(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:136:

ATG Met 1	GGC Gly	ACC Thr	TTG Leu	CGG Arg 5	GAT Asp	TTA Leu	CAG Gln	TAC Tyr	GCG Ala 10	CTC Leu	CAG Gln	GAG Glu	AAG Lys	ATC Ile 15	GAG Glu	48
GAG Glu	CTG Leu	AGG Arg	CAG Gln 20	CGG Arg	GAT Asp	GCT Ala	CTC Leu	ATC Ile 25	GAC Asp	GAG Glu	CTG Leu	GAG Glu	CTG Leu 30	GAG Glu	TTG Leu	96
GAT Asp	CAG Gln	AAG Lys 35	GAC Asp	GAA Glu	CTG Leu	ATC Ile	CAG Gln 40	AAG Lys	CTG Leu	CAG Gln	AAC Asn	GAG Glu 45	CTG Leu	GAC Asp	AAG Lys	144
TAC Tyr	CGC Arg 50	TCG Ser	GTG Val	ATC Ile	CGA Arg	CCA Pro 55	GCC Ala	ACC Thr	CAG Gln	CAG Gln	GCG Ala 60	CAG Gln	AAG Lys	CAG Gln	AGC Ser	192
GCG Ala 65	AGC Ser	ACC Thr	TTG Leu	CAG Gln	GGC Gly 70	GAG Glu	CCG Pro	CGC Arg	ACC Thr	AAG Lys 75	CGG Arg	CAG Gln	GCG Ala	ATC	TCC Ser 80	240
GCC Ala	GAG Glu	CCC	ACC Thr	GCC Ala 85	TTC Phe	GAC Asp	ATC Ile	CAG Gln	GAT Asp 90	CTC	AGC Ser	CAT His	GTG Val	ACC Thr 95	CTG Leu	288
CCC Pro	TTC Phe	TAC Tyr	CCC Pro 100	AAG Lys	AGC Ser	CCA Pro	CAG Gln	TCC Ser 105	AAG Lys	GAT Asp	CTI Leu	`ATA	AAG Lys 110	Glu	GCT Ala	336
ATC Ile	CTT Leu	GAC Asp 115) Asn	GAC Asp	TTT Phe	ATG Met	AAG Lys 120	Asn	TTG Leu	GAG Glu	CTC Lev	TCG Ser 125	Gln	ATC lle	CAG Gln	384
GAG Glu	ATT Ile 130	Val	GAT Asp	TGI Cys	ATG Met	TAC Tyr 135	Pro	GTG Val	GAG Glu	TAT TYT	GG(Gl ₂ 14(/ Lys	GAC Asp	AGT Ser	TGC Cys	432
ATC Ile 145	Ile	Lys	A GAA s Glu	GGA Gly	GAC Asp 150	Val	. G1 <u>y</u>	TCA Ser	CTG	GT(Val 155	l Tyı	r GTC r Val	Met	G GAA	A GAT 1 Asp 160	480
GGT Gly	`AAG	GT Va	r GAÆ l Glu	GT1 Val 165	Thr	AAA Lys	s Glu	A GGT 1 Gly	/ Val	L Ly:	s Le	n CA	r ACC	C ATO r Met 17!	G GGT t Gly 5	528
CCA Pro	GGA Gly	A AA.	A GTC s Val	Ph•	r GGC e Gly	GAA Glu	A TTO	G GCT L Ala 185	a Ile	r CT	T TAG	C AAG	TG n Cy: 19	s Th	c ccc r Arg	576
ACA Thi	A GCC	G AC	r Vai	D AAG	G ACT	r CT	r GT. ı Va 20	l Ası	r GTZ n Val	A AA l Ly	A CT s Le	C TG u Tr 20	p Al	C AT a Il	T GAT e Asp	624
CGA	A CA	A TG	т тт	r ca	A AC	A AT	TA A	G ATY	G AG	G AC	A GG	A CT	C AT	C AA	G CAT	672

Arg	Gln 210	Cys	Phe	Gln		Ile 215	Met	Met	Arg	Thr	Gly 220	Leu	Ile	Lys	His	
ACC Thr 225	GAG Glu	TAT Tyr	ATG Met	GAA Glu	TTT Phe 230	TTA Leu	AAA Lys	AGC Ser	GTT Val	CCA Pro 235	ACA Thr	TTC Phe	CAG Gln	AGC Ser	CTT Leu 240	720
CCT Pro	GAA Glu	GAG Glu	ATC Ile	CTC Leu 245	AGC Ser	AAG Lys	CTT Leu	GCT Ala	GAT Asp 250	GTC Val	CTT Leu	GAA Glu	GAG Glu	ACC Thr 255	CAC His	768
TAT Tyr	GAA Glu	AAT Asn	GGA Gly 260	GAA Glu	TAT Tyr	ATT Ile	ATC Ile	AGG Arg 265	CAA Gln	GGT Gly	GCA Ala	AGA Arg	GGG Gly 270	GAC Asp	ACC Thr	816
TTC Phe	TTT Phe	ATC Ile 275	ATC Ile	AGC Ser	AAA Lys	GGA Gly	ACG Thr 280	GTA Val	AAT Asn	GTC Val	ACT Thr	CGT Arg 285	GAA Glu	GAC Asp	TCA Ser	864
CCG Pro	AGT Ser 290	GAA Glu	GAC Asp	CCA Pro	GTC Val	TTT Phe 295	CTT Leu	AGA Arg	ACT Thr	TTA Leu	GGA Gly 300	AAA Lys	GGA Gly	GAC Asp	TGG Trp	912
TTT Phe 305	Gly	GAG Glu	AAA Lys	GCC Ala	TTG Leu 310	CAG Gln	GGG Gly	GAA Glu	GAT Asp	GTG Val 315	Arg	ACA Thr	GCA Ala	AAC Asn	GTA Val 320	960
ATT	GCT Ala	GCA Ala	GAA Glu	GCT Ala 325	Val	ACC Thr	TGC Cys	CTT	GTG Val 330	Ile	GAC	AGA Arg	GAC Asp	TCT Ser 335	TTT Phe	1008
AAA Lys	CAT His	TTG	ATT Ile 340	Gly	GGG Gly	CTG Leu	GAT Asp	GAT Asp 345	Val	TCT Ser	AAT Asn	'AAA Lys	GCA Ala 350	Туг	GAA Glu	1056
GAT Asp	GCA Ala	GA# Glu 355	ı Ala	'AAA	GCA Ala	AAA Lys	ТАТ Туг 360	Glu	GCT Ala	GAZ Glu	A GCG	GCT Ala 365	Phe	TTC Phe	GCC Ala	1104
AAC Asr	2 CTC 1 Let 370	Lys	G CTC	TCI Ser	GAT Asp	TTC Phe 375	Asr	ATC	: ATI	GA:	Thi 380	Let	GGA Gly	A GT:	r GGA l Gly	1152
GG: Gly 385	y Phe	GG/	A CGA Y Arg	A GTA	A GAP Glu 390	ı Lev	GTC 1 Val	CAC L Glr	TTC	39!	s Sei	r GAZ c Glu	A GAZ	A TCC	C AAA r Lys 400	1200
ACC Thi	G TT:	r GC/ e Ala	A ATX a Met	AAC Lys	s Ile	r CTC	AAC Ly:	G AAA	A CG' S Arg 410	g Hi	C AT	r GTG e Val	G GAG	C AC. p Th: 41	A AGA r Arg 5	1248
CA(G CA(G GAG	G CAG u His 420	s Ile	C CGG	TC g Sei	A GAG	3 AAG 1 Lys 42!	s Gl	G AT	C ATO	G CAG	G GG n Gl 43	y Al	T CAT a His	1296
TC Se	C GA' r As	r TT p Ph 43	e Il	A GTY e Va	g AGA	A CTO	3 TA 1 Ty 44	r Ar	A AC. g Th	A TT r Ph	T AA e Ly	G GA S As 44	p Se	C AA r Ly	A TAT	1344

							TGT Cys									1392
							GAA Glu									1440
							GCC Ala									1488
							AAT Asn									1536
						_	TTT Phe 520									1584
							ACT Thr									1632
							ATT Ile									1680
							ACT Thr									1728
							ATC Ile				_				_	1776
							AAA Lys 600								AAA Lys	1824
							GAA Glu								GGA Gly	1872
							AAA Lys				_				GAA Glu 640	1920
							ACA Thr								GCA Ala	1968
															GAT Asp	2016
GAA	CCA	CCA	CCT	GAT	GAC	AAC	TCA	GGA	TGG	GAT	ATA	GAC	TTC	TCG	GAT	2064

Glu	Pro	Pro 675	Pro	Asp	Asp		Ser 680	Gly	Trp	Asp	Ile	Asp 685	Phe	Ser	Asp	
CCA Pro	CCG Pro 690	GTC Val	GCC Ala	ACC Thr	ATG Met	GTG Val 695	AGC Ser	AAG Lys	GGC Gly	GAG Glu	GAG Glu 700	CTG Leu	TTC Phe	ACC Thr	GGG Gly	2112
GTG Val 705	GTG Val	CCC Pro	ATC Ile	CTG Leu	GTC Val 710	GAG Glu	CTG Leu	GAC Asp	GGC Gly	GAC Asp 715	GTA Val	AAC Asn	GGC Gly	CAC His	AAG Lys 720	2160
TTC Phe	AGC Ser	GTG Val	TCC Ser	GGC Gly 725	GAG Glu	GGC Gly	GAG Glu	GGC Gly	GAT Asp 730	GCC Ala	ACC Thr	TAC Tyr	Gly	AAG Lys 735	CTG Leu	2208
ACC Thr	CTG Leu	AAG Lys	TTC Phe 740	ATC Ile	TGC Cys	ACC Thr	ACC Thr	GGC Gly 745	AAG Lys	CTG Leu	CCC Pro	GTG Val	CCC Pro 750	TGG Trp	CCC Pro	2256
ACC Thr	CTC Leu	GTG Val 755	ACC Thr	ACC Thr	CTG Leu	ACC Thr	TAC Tyr 760	GGC Gly	GTG Val	CAG Gln	TGC Cys	TTC Phe 765	AGC Ser	CGC Arg	TAC Tyr	2304
CCC Pro	GAC Asp 770	CAC	ATG Met	AAG Lys	CAG Gln	CAC His 775	GAC Asp	TTC Phe	TTC Phe	AAG Lys	TCC Ser 780	Ala	ATG Met	CCC Pro	GAA Glu	2352
GGC Gly 785	Tyr	GTC Val	CAG Gln	GAG Glu	CGC Arg 790	ACC Thr	ATC Ile	TTC Phe	TTC Phe	AAG Lys 795	Asp	GAC Asp	GGC	AAC Asn	TAC Tyr 800	2400
AAG Lys	ACC Thr	CGC Arg	GCC Ala	GAG Glu 805	Val	AAG Lys	TTC	GAG Glu	GGC Gly 810	Asp	: ACC	CTG Leu	GTC Val	AAC Asn 815	CGC Arg	2448
ATC Ile	GAG Glu	CTG Leu	AAG Lys 820	Gly	TATC	GAC	TTC Phe	AAG Lys 825	Glu	GAC Asp	GGC Gly	AAC Asn	TATC 110 830	e Leu	G GGG	2496
CAC His	AAG Lys	6 CTC 6 Lev 835	Glu	TAC Tyr	AAC Asn	TAC Tyr	AAC Asr 840	Ser	CAC His	AAC Asr	GTC n Val	TAT 1 Tyr 845	: 11	TA C	G GCC Ala	2544
GAC Asp	AAC Lys	Glr	AAC Lys	AAC Asr	GGC Gly	11e	Lys	G GTG S Val	AAC Asr	TTO	AAC 2 Ly: 860	s Ile	e Arg	C CAG	C AAC s Asn	2592
AT0 116 869	e Glu	GAC 1 Asp	GG(AGC Sei	GTC Val	Glr	CTC	C GCC	GAG ASI	C CAG P Hi: 87	s Ty:	C CAG	G CA	G AA n As:	C ACC n Thr 880	2640
CC0 Pro	TA C	C GG(e Gly	C GAG Y Asi	GG(Gl ₂ 88!	y Pro	GTC Val	G CTO	G CTY	G CCG 2 Pro 89	o As	C AA p As:	C CA	С ТА s Ту	C CT r Le 89	G AGC u Ser 5	2688
AC(C CAC	G TCC	C GCC r Ala 90	a Le	G AGG	C AA	A GA	C CCC p Pro 90!	aA c	C GA n Gl	G AA u Ly	G CG s Ar	C GA g As 91	рHi	C ATG s Met	2736

GTC CTG GAG TTC GTG ACC GCC GGC GGG ATC ACT CTC GGC ATG GAC 278

Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp
915 920 925

GAG CTG TAC AA GTAA Glu Leu Tyr Lys 930 2799

(2) INFORMATION FOR SEQ ID NO:137:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 932 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:137:

Met Gly Thr Leu Arg Asp Leu Gln Tyr Ala Leu Gln Glu Lys Ile Glu 10 1 5 Glu Leu Arg Gln Arg Asp Ala Leu Ile Asp Glu Leu Glu Leu Glu Leu 25 Asp Gln Lys Asp Glu Leu Ile Gln Lys Leu Gln Asn Glu Leu Asp Lys 40 Tyr Arg Ser Val Ile Arg Pro Ala Thr Gln Gln Ala Gln Lys Gln Ser 55 Ala Ser Thr Leu Gln Gly Glu Pro Arg Thr Lys Arg Gln Ala Ile Ser 75 70 65 Ala Glu Pro Thr Ala Phe Asp Ile Gln Asp Leu Ser His Val Thr Leu 90 85 Pro Phe Tyr Pro Lys Ser Pro Gln Ser Lys Asp Leu Ile Lys Glu Ala 105 110 Ile Leu Asp Asn Asp Phe Met Lys Asn Leu Glu Leu Ser Gln Ile Gln 125 120 115 Glu Ile Val Asp Cys Met Tyr Pro Val Glu Tyr Gly Lys Asp Ser Cys 140 135 Ile Ile Lys Glu Gly Asp Val Gly Ser Leu Val Tyr Val Met Glu Asp 155 150 Gly Lys Val Glu Val Thr Lys Glu Gly Val Lys Leu Cys Thr Met Gly 165 170 Pro Gly Lys Val Phe Gly Glu Leu Ala Ile Leu Tyr Asn Cys Thr Arg 190 185 Thr Ala Thr Val Lys Thr Leu Val Asn Val Lys Leu Trp Ala Ile Asp 200 205 Arg Gln Cys Phe Gln Thr Ile Met Met Arg Thr Gly Leu Ile Lys His 210 215 220 Thr Glu Tyr Met Glu Phe Leu Lys Ser Val Pro Thr Phe Gln Ser Leu 230 235 Pro Glu Glu Ile Leu Ser Lys Leu Ala Asp Val Leu Glu Glu Thr His 250 245 Tyr Glu Asn Gly Glu Tyr Ile Ile Arg Gln Gly Ala Arg Gly Asp Thr 270 265 260 Phe Phe Ile Ile Ser Lys Gly Thr Val Asn Val Thr Arg Glu Asp Ser

		275					280					285			
	290					295					300				
305	=				Leu 310					315					320
Ile				325	Val				330					335	
_			340		Gly			345					350		
		355			Ala		360					365			
	370				Asp	375					380				
385					Glu 390					395					400
				405	Ile				410					415	
			420		Arg			425					430		
		435			Arg		440					445			
	450				Glu	455					460				
465					Ser 4 70					475					480
				485					490					495	
			500	1	Pro			505					510		
		515	5				520					525			Lys
	530)				535					540)			Ile
545	,				550					555	,				Gly 560
				565	<u>, </u>				570)				575	
			580)				585					590)	Ile
		595	5				600					605	5		Lys
	61)				615	ı				620)			Gly
625	5				630)				63	5				Glu 640
				645	5				650	C				65	
			66	0				665	5				670	3	n Asp
		67	5				680)				68!	5		r Asp
	69	0				695	5				70	0			r Gly s Lys
70	5				710)				71	5				720
				72	5				73	0				73	
Th	r Le	u Ly	s Ph	e Il	e Cy:	s Th:	r Ini	L GT.	у гу	ച പല	u PI	o va	T LT	J 11	p Pro

			740					745					750			
Thr	Leu	Val 755	Thr	Thr	Leu	Thr	Tyr 760	Gly	Val	Gln	Суѕ	Phe 765	Ser	Arg	Tyr	
Pro	Asp 770	His	Met	Lys	Gln	His 775		Phe	Phe	Lys	Ser 780		Met	Pro	Glu	
Gly 785	Tyr	Val	Gln	Glu	Arg 790	Thr	Ile	Phe	Phe	Lys 795	Asp	Asp	Gly	Asn	Tyr 800	
Lys	Thr	Arg	Ala	Glu 805		Lys	Phe	Glu	Gly 810		Thr	Leu	Val	Asn 815		
Ile	Glu	Leu	Lys 820		Ile	Asp	Phe	Lys 825		Asp	Gly	Asn	Ile 830	_	Gly	
His	Lys	Leu 835	Glu	Tyr	Asn	Tyr	Asn 840		His	Asn	Val	Tyr 845		Met	Ala	
Asp			Lys	Asn	Gly			Val	Asn	Phe			Arg	His	Asn	
	850 Glu	Asp	Gly	Ser		855 Gln	Leu	Ala	Asp		860 Tyr	Gln	Gln	Asn	Thr	
865	710	C1	7.55	C1	870	1101	T	T 011	D	875		,,,,,	m	•	880	
			Asp	885					890					895		
Thr	Gln	Ser	Ala 900	Leu	Ser	Lys	Asp	Pro 905	Asn	Glu	Lys	Arg	Asp 910	His	Met	
Val	Leu	Leu 915	Glu	Phe	Val	Thr	Ala 920	Ala	Gly	Ile	Thr	Leu 925	Gly	Met	Asp	
Glu	Leu 930	Tyr	Lys									3_3				
	(:	(C) (D) (ii) N (ix) N (A) (B) (D)	TYPI STRA TOPO MOLEC FEATT NAI LOC OTE	ANDEI DLOGY CULE JRE: - TE/KI CATIC HER I	ONESS (: 1: TYPE EY: (ON: 1 INFOR	S: siinear E: cI Codir	ingle DNA ng Se 2181	equer		NO:1	138:					
									-							
			AAG Lys													4.8
			GAC Asp 20													96
			GGC Gly													144
			GGC Gly													192

CTG . Leu '	ACC Thr	TAC Tyr	GGC Gly	GTG Val	CAG Gln 70	TGC Cys	TTC Phe	AGC Ser	Arg	TAC Tyr 75	CCC Pro	GAC Asp	CAC His	ATG Met	AAG Lys 80	240
CAG Gln	CAC His	GAC Asp	TTC Phe	TTC Phe 85	AAG Lys	TCC Ser	GCC Ala	ATG Met	CCC Pro 90	GAA Glu	GGC Gly	TAC Tyr	GTC Val	CAG Gln 95	GAG Glu	288
CGC Arg	ACC Thr	ATC Ile	TTC Phe 100	TTC Phe	AAG Lys	GAC Asp	GAC Asp	GGC Gly 105	AAC Asn	TAC Tyr	AAG Lys	ACC Thr	CGC Arg 110	GCC Ala	GAG Glu	336
GTG Val	AAG Lys	TTC Phe 115	GAG Glu	GGC Gly	GAC Asp	ACC Thr	CTG Leu 120	GTG Val	AAC Asn	CGC Arg	ATC Ile	GAG Glu 125	CTG Leu	AAG Lys	GGC Gly	384
ATC Ile	GAC Asp 130	TTC Phe	AAG Lys	GAG Glu	GAC Asp	GGC Gly 135	AAC Asn	ATC Ile	CTG Leu	GGG Gly	CAC His 140	AAG Lys	CTG Leu	GAG Glu	TAC Tyr	432
AAC Asn 145	TAC Tyr	AAC Asn	AGC Ser	CAC His	AAC Asn 150	GTC Val	TAT Tyr	ATC	ATG Met	GCC Ala 155	GAC Asp	AAG Lys	CAG Gln	AAG Lys	AAC Asn 160	480
GGC Gly	ATC Ile	AAG Lys	GTG Val	AAC Asn 165	Phe	AAG Lys	ATC	CGC Arg	CAC His 170	AAC Asn	ATC Ile	GAG Glu	GAC Asp	GGC Gly 175	AGC Ser	528
GTG Val	CAG Gln	CTC	GCC Ala 180	Asp	CAC His	TAC Tyr	CAG Gln	CAG Gln 185	Asn	ACC	CCC	ATC Ile	GGC Gly 190	Asp	GGC Gly	576
CCC Pro	GTG Val	Lev 195	Leu	CCC Pro	GAC Asp	AAC Asn	CAC His 200	Tyr	CTG Leu	AGC Ser	ACC Thr	CAC Glr 205	Ser	GCC Ala	CTG Leu	624
AGC Ser	Lys 210	Ası	CCC Pro	AAC Asr	GAG Glu	AAG Lys 215	Arg	GAT Asp	CAC His	: ATC	GTC Val 220	Leu	CTC	G GAG	TTC Phe	672
GTG Val 225	Thr	GCC Ala	a Ala	GGC Gly	/ Ile	Thr	Leu	ı Gly	ATC	: Asp	Glu	CTC	TAC Ty:	C AAG	G TCC s Ser 240	720
GGA Gly	A CTO	AG Arg	A TCT g Ser	CGA Arg 245	g Gly	ACC Thr	Met	S AGO	GAC Asr 250	va:	G GCT l Ala	T ATT	r GTM e Va	G AA 1 Ly 25	G GAG s Glu 5	768
GGT Gly	r TGO 7 Tri	G CT	G CAC u His 260	Ly:	A CGA	A GG(g Gly	G GAC	TAC 1 Ty1 265	: Ile	C AAG e Ly:	G ACC	TG(G CG p Ar 27	g Pr	A CGC o Arg	816
TAC Tyi	r Ph	C CT e Le 27	u Lei	C AAG	G AA' s Ası	T GAT	r GG(5 Gl) 28	y Thi	r Phe	C AT e Il	T GG(e Gl	C TA y Ty 28	r Ly	G GA s Gl	G CGG u Arg	864
CCC	G CA	G GA	T GT	G GA	C CA	A CG	r ga	G GC	r cc	C CT	C AA	C AA	C TI	C IC	T GTG	912

Pro	Gln 290	Asp	Val	Asp	Gln	Arg 295	Glu	Ala	Pro	Leu	Asn 300	Asn	Phe	Ser	Val	
					ATG Met 310											960
					CAG Gln											1008
					GAG Glu											1056
					AAG Lys											1104
	_				GAC Asp											1152
					CAC His 390											1200
					GGC Gly											1248
	_		_		TAC Tyr											1296
_	_	_			GAG Glu											1344
					CAC His											1392
					CTC Leu 470											1440
_					CTG Leu											1488
					GCT Ala											1536
					GTG Val											1584

CTG Leu	GAC Asp 530	AAG Lys	GAC Asp	GGG Gly	His	ATT Ile 535	AAG Lys	ATC Ile	ACA Thr	GAC Asp	TTC Phe 540	GGG Gly	CTG Leu	TGC Cys	AAG Lys	1632
GAG Glu 545	GGG Gly	ATC Ile	AAG Lys	GAC Asp	GGT Gly 550	GCC Ala	ACC Thr	ATG Met	AAG Lys	ACC Thr 555	TTT Phe	TGC Cys	GGC Gly	ACA Thr	CCT Pro 560	1680
GAG Glu	TAC Tyr	CTG Leu	GCC Ala	CCC Pro 565	GAG Glu	GTG Val	CTG Leu	GAG Glu	GAC Asp 570	AAT Asn	GAC Asp	TAC Tyr	GGC Gly	CGT Arg 575	GCA Ala	1728
GTG Val	GAC Asp	TGG Trp	TGG Trp 580	GGG Gly	CTG Leu	GGC Gly	GTG Val	GTC Val 585	ATG Met	TAC Tyr	GAG Glu	ATG Met	ATG Met 590	TGC Cys	GGT Gly	1776
CGC Arg	CTG Leu	CCC Pro 595	Phe	TAC Tyr	AAC Asn	CÁG Gln	GAC Asp 600	CAT His	GAG Glu	AAG Lys	CTT Leu	TTT Phe 605	GAG Glu	CTC Leu	ATC Ile	1824
CTC Leu	ATG Met 610	GAG Glu	GAG Glu	ATC Ile	CGC Arg	TTC Phe 615	CCG Pro	CGC Arg	ACG Thr	CTT Leu	GGT Gly 620	Pro	GAG Glu	GCC Ala	AAG Lys	1872
TCC Ser 625		CTT Leu	TCA Ser	GGG Gly	CTG Leu 630	CTC Leu	AAG Lys	AAG Lys	GAC Asp	CCC Pro 635	Lys	CAG Gln	AGG Arg	CTT Leu	GGC Gly 640	1920
GGG	GGC Gly	TCC Ser	GAG Glu	GAC Asp 645	Ala	AAG Lys	GAG Glu	ATC Ile	ATG Met	Gln	CAT	CGC Arg	TTC Phe	Phe	GCC Ala	1968
GGT Gly	'ATC	GTC Va]	TGG Trp	Glr	CAC His	GTG Val	ТАС Туг	GAG Glu 665	Lys	AAG Lys	CTC Lev	AGC Ser	Pro 670	Pro	TTC Phe	2016
AAC Lys	CCC Pro	CAC Glr 675	ı Val	ACC Thr	TCG Ser	GAC Glu	ACT Thr	Asp	C ACC	AGC Arg	TA:	r TTT c Phe 685	Asr	GAC	G GAG	2064
TTC Phe	ACC Thr	Ala	CAC	ATC	∶ll∈	ACC Thr 695	116	C ACA	A CCA	A CCT	GAG ASI 70	o Glr	GAT	r GAG o Asi	AGC Ser	2112
ATO Met	: Glv	TG' Cy:	r GTC s Val	GAC LASI	2 AGC 5 Ser 710	Glu	G CG(C AGG g Arg	g CC	C CAC O His 71!	s Ph	c cco	CAC O Gli	g TTO	TCC e Ser 720	2160
	C TCC				r Thr			A								2184

- (2) INFORMATION FOR SEQ ID NO:139:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 727 amino acids

- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:139:

1			Lys	5					10					15	
Val	Glu	Leu	Asp 20	Gly	Asp	Val		Gly 25	His	Lys	Phe	Ser	Val 30	Ser	Gly
Glu	Gly	Glu 35	Gly	Asp	Ala	Thr	Tyr 40	Gly	Lys	Leu	Thr	Leu 45	Lys	Phe	Ile
Cys	Thr 50	Thr	Gly	Lys	Leu	Pro 55	Val	Pro	Trp	Pro	Thr 60	Leu	Val	Thr	Thr
65	Thr		Gly		70					75					80
			Phe	85					90					95	
			Phe 100					105					110		
		115	Glu				120					125			
	130		Lys			135					140				
145			Ser		150					155					160
			Val	165					170					1/5	
			Ala 180					185					190		
		195					200					205			Leu
	210					215					220				Phe
225					230					235					Ser 240
				245					250)				255	
			260					265	,				270		Arg
		275	5				280					285)		Arg
	290)				295	,				300)			val
Ala 305		ı Çys	s Glr	Let	Met 310		Thr	Glu	ı Arç	3 Pro	Arg	g Pro) Asr	ı Tnı	2 Phe 320
Il€	, E Ile	e Arg	g Cys	Lei			Thr	Thr	. Val	l Ile	e Glu	ı Arg	y Thi	Phe	e His
	- C1.	. Th	- חיי	325	. Gl	. Arc	, Gli	. Gli	330 Tra		r Thi	r Ala	a Ile	339 Gl:	n Thr
			340)				345	5				350	נ	
		35.	5				360)				36	5		e Arg
	370	C				37!	5				38	0			l Ser
Le	ı Ala	a Ly	s Pro	o Ly:	s His	s Ar	g Vai	l Thi	r Me	t As	n Gl	u Ph	e Gl	и Ту	r Leu

385					390					395					400
Lys	Leu	Leu	Gly	Lys 405	Gly	Thr	Phe		Lys 410	Val	Ile	Leu	Val	Lys 415	Glu
Lys	Ala	Thr	Gly 420	Arg	Tyr	Tyr	Ala	Met 425	Lys	Ile	Leu	Lys	Lys 430	Glu	Val
Ile	Val	Ala 435	Lys	Asp	Glu	Val	Ala 440	His	Thr	Leu	Thr	Glu 445	Asn	Arg	Val
Leu	Gln 450		Ser	Arg	His	Pro	Phe	Leu	Thr	Ala	Leu 460	Lys	Tyr	Ser	Phe
		His	Asp	Arg	Leu 470	Cys	Phe	Val	Met	Glu 475	Tyr	Ala	Asn	Gly	Gly 480
465 Glu	Leu	Phe	Phe	His	Leu	Ser	Arg	Glu	Arg 490		Phe	Ser	Glu	Asp	Arg
Ala	Arg	Phe	Tyr 500		Ala	Glu	Ile	Val		Ala	Leu	Asp	Tyr 510	Leu	His
Ser	Glu	Lys 515	Asn	Val	Val	Tyr	Arg 520		Leu	Lys	Leu	Glu 525	Asn	Leu	Met
Leu	Asp		Asp	Gly	His	Ile 535	Lys	Ile	Thr	Asp	Phe 540	Gly	Leu	Cys	Lys
Glu 545		Ile	Lys	Asp	Gly 550	Ala	Thr	Met	Lys	Thr 555	Phe	Cys	Gly	Thr	Pro 560
	Tyr	Leu	Ala	Pro 565	Glu	Val	Leu	Glu	Asp 570		Asp	Tyr	Gly	Arg 575	Ala
Val	Asp	Trp	Trp 580	Gly	Leu	Gly	Val	Val 585	Met	Tyr	Glu	Met	Met 590	Суѕ	Gly
Arg	Leu	Pro		Tyr	Asn	Gln	Asp 600	His	Glu	Lys	Leu	Phe 605	Glu	Leu	Ile
Leu	Met 610		Glu	Ile	Arg	Phe 615		Arg	Thr	Leu	Gly 620		Glu	Ala	Lys
Ser 625		Leu	Ser	Gly	Leu 630		Lys	Lys	Asp	Pro 635		Glr	Arg	Leu	640
Gly	Gly	Ser	Glu	Asp 645		Lys	Glu	Ile	Met 650		His	Arg	p Phe	Ph∈ 655	Ala
_			660)				665					670)	Phe
		675	5				680					685	5		ı Glu
	690)				695	i				700)			Ser
705	5				Ser 710 Thr)		Arg	Pro	715		e Pro	o Glr	n Ph€	720
тух	. sei	. Alā	1 561	729		. Ald	•								

- (2) INFORMATION FOR SEQ ID NO:140:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2394 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...2391
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:140:

			ATC Ile						48
			ATT Ile						96
			GGG Gly 40						144
			AAG Lys						192
			GTG Val						240
			CAC His						288
			CTC Leu						336
			TGT Cys 120						384
			ACC Thr						432
			TAC Tyr						480
	Thr	Arg	CCA Pro	Gly	Arg				528
								GCC Ala	576
								GGT Gly	624
								ATT Ile	672

GAG (Glu ' 225	GTG Val	тат Туг	TTC Phe	ACG Thr	GGA Gly 230	CCA Pro	GGC Gly	TGG Trp	GAG Glu	GCC Ala 235	CGA A rg	GGC Gly	TCC Ser	TTT Phe	TCC Ser 240	r	720
CAA Gln	GCT Ala	GAT Asp	GTC Val	CAC His	Arg	CAA Gln	GTG Val	GCC Ala	ATT Ile 250	GTG Val	TTC Phe	CGG Arg	ACC Thr	CCT Pro 255	CCC Pr	C 0	768
TAC Tyr	GCA Ala	GAC Asr	260	Ser	CTG Leu	CAG Gln	GCT Ala	CCT Pro 265	GTG Val	CGT Arg	GTC Val	TCC Ser	ATG Met 270	CAG Gln	CT Le	G u	816
CGG Arg	CGG Arg	CCT Pro	Se:	GAC Asp	CGG Arg	GAG Glu	CTC Leu 280	AGT Ser	GAG Glu	CCC	ATG Met	GAA Glu 285	TTC Phe	CAG Gln	TA Ty	C T	864
CTG Leu	CCA Pro	As	r ac. o Th	A GAC r Asp	GAT Asp	CGT Arg 295	His	CGG Arg	ATT	GAG Glu	GAG Glu 300	Lys	CGT Arg	AAA Lys	A AC	G g	912
ACA Thr 305	ТАТ Тут	GA	G AC u Th	C TIV	2 AAC 2 Lys 310	Ser	ATC	ATG Met	AAG Lys	AAG Lys 315	Ser	CCT Pro	TTC Phe	AGC Sei	r G	GA ly 20	960
CCC Pro	ACC Thr	GA As	C CC p Pr	C CGG O Arg	g Pro	r CCA o Pro	A CCT	r CGA o Arg	CGC Arg	Ile	GCT Ala	r GTC a Val	CCT Pro	TC0 Se:	r A	GC rg	1008
AGC Ser	TC/ Sei	A GC	T TC a Se	r Va	C CCO	C AAC o Lys	G CCA	A GCA D Ala 345	Pro	CAC Gli	G CCC	C TAT	r CCC r Pro 350	o Ph	T A	CG hr	1056
TCA Ser	TC(C CI r Le	eu Se	C AC	C AT	C AA(e Ası	TA' 1 Ty: 36	T GAT r Asp 0	GAC	F TT	T CC	C ACC O Th: 36	r Me	G GT t Va	G T .1 P	TT he	1104
CCT	TC Se 37	r G	G CA	AG AT In Il	C AG e Se	C CAG r Gli 37	n Al	C TCC	GCC Ala	TT a Le	G GC u Al 38	a Pr	G GC o Al	C CC a Pr	T C	cc	1152
CAA Glr 385	ı Va	C C'	rg Co eu P:	cc ca ro Gl	AG GC .n Al	a Pr	A GC o Al	C CC a Pro	r GCO	C CC a Pr 39	O Al	T CC a Pr	A GC	C AT	י ספ	GTA Val 100	1200
TC? Ser	A GC r Al	T C a L	TG G eu A	la G	AG GC ln Al	C CC a Pr	A GC o Al	c cc a Pr	r GT o Va 41	l Pr	A GT to Va	C CI	'A GC eu Al	a Pi	CA (ro (15	GC Gly	1248
CC'	T CC	T C	ln A	CT G' la V 20	rg go al Al	CC CC	A CC	T GC to Al 42	a Pr	C A.	AG CC /s Pi	CC AC	r G	AG G In A 30	CT (GG Gly	1296
GA G1	A GC u Gl	уТ	CG Chr L	TG T eu S	CA GA er G	AG GC lu Al	a Le	NG CT eu Le 10	G CA	G Ci	rg CA eu Gi	ln Fl	MT GA ne As 45	AT G sp A	AT (GAA Glu	1344
GA	.c c1	rg c	GG C	CC T	TG C'	rt GO	BC A	AC AG	C AC	A G	AC C	CA G	CT G	TG I	TC	ACA	1392

Asp	Leu 450	Gly	Ala	Leu		Gly . 455	Asn	Ser	Thr	Asp	Pro 460	Ala	Val	Phe	Thr	
GAC Asp 465	CTG Leu	GCA Ala	TCC Ser	GTC Val	GAC Asp 470	AAC Asn	TCC Ser	GAG Glu	TTT Phe	CAG Gln 475	CAG Gln	CTG Leu	CTG Leu	AAC Asn	CAG Gln 480	1440
GGC Gly	ATA Ile	CCT Pro	GTG Val	GCC Ala 485	CCC Pro	CAC His	ACA Thr	ACT Thr	GAG Glu 490	CCC Pro	ATG Met	CTG Leu	ATG Met	GAG Glu 495	TAC Tyr	1488
CCT Pro	GAG Glu	GCT Ala	ATA Ile 500	ACT Thr	CGC Arg	CTA Leu	GTG Val	ACA Thr 505	GGG Gly	GCC Ala	CAG Gln	AGG Arg	CCC Pro 510	CCC Pro	GAC Asp	1536
CCA Pro	GCT Ala	CCT Pro 515	GCT Ala	CCA Pro	CTG Leu	GGG Gly	GCC Ala 520	CCG Pro	GGG Gly	CTC Leu	CCC Pro	AAT Asn 525	GGC Gly	CTC Leu	CTT Leu	1584
TCA Ser	GGA Gly 530	GAT Asp	GAA Glu	GAC Asp	TTC Phe	TCC Ser 535	TCC Ser	ATT Ile	GCG Ala	GAC Asp	ATG Met 540	GAC Asp	TTC Phe	TCA Ser	GCC Ala	1632
CTG Leu 545	Leu	AGT Ser	CAG Gln	ATC Ile	AGC Ser 550	TCC Ser	TTG Leu	GAT Asp	CCA Pro	CCG Pro 555	Val	GCC Ala	ACC Thr	: ATG : Met	GTG Val 560	1680
AGC Ser	AAG Lys	GGC Gly	GAG Glu	GAG Glu 565	CTG Leu	TTC Phe	ACC Thr	GGG Gly	GTG Val 570	GTG Val	CCC Pro	TATC	CTC Lev	GTC Val	GAG Glu	1728
CTG Leu	GAC Asp	GGC GGC	GAC Asp 580	Val	AAC Asn	GGC Gly	CAC	AAG Lys 585	Phe	AGC Ser	GTC Val	TCC Ser	GG(G1 ₂ 59(/ Glu	GGC Gly	1776
GAC Glu	GGC Gly	GAT Asp 595	Ala	ACC Thr	TAC Tyr	GGC	AAG Lys 600	Leu	ACC Thr	CTC	AAC Lys	FTTC FPhe 605	e Ile	C TGC	ACC Thr	1824
ACC Thi	GGC Gly 610	Lys	CTC	CCC Pro	GTG Val	CCC Pro 615	Trp	CCC Pro	ACC Thr	CTC	C GTY L Val 620	l Thi	C ACC	C CTO	ACC Thr	1872
ТАС Туз 625	c Gly	GT(G CAC	TGC	TTC Phe	Ser	CG(TAC TYI	C CCC	GAG Ası 63	p Hi	C ATG	G AA	G CAG	G CAC n His 640	1920
GA(As ₁	TTC p Phe	TTO	C AAG e Lys	5 TCC 5 Ser 645	: Ala	ATG Met	CCC Pro	GAZ Gli	A GG(L Gl) 65(у Ту	C GT r Va	C CA	G GA n Gl	G CG u Ar 65	C ACC g Thr 5	1968
ATY Il	C TTO e Pho	C TTG ∋ Ph	C AA0 e Ly:	s Ası	C GAC	GGC Gly	AA(Asi	TA n Ty: 66	r Ly:	G AC s Th	C CG r Ar	C GC g Al	C GA a G1 67	u Va	G AAG 1 Lys	2016
TT Ph	C GAG e Gl	G GG u Gl 67	y As	C ACC	CTX	G GTC u Val	AA L As 68	n Ar	C AT	C GA e Gl	G CT u Le	G AA u Ly 68	s Gl	C AT	C GAC e Asp	2064

	AAG Lys 690	GAG Glu	GAC Asp	GGC Gly	AAC Asn	ATC Ile 695	CTG Leu	GGG Gly	CAC His	AAG Lys	CTG Leu 700	GAG Glu	TAC Tyr	AAC Asn	TAC Tyr	2112
AAC Asn 705	AGC Ser	CAC His	AAC Asn	GTC Val	тат тут 710	ATC Ile	ATG Met	GCC Ala	GAC Asp	AAG Lys 715	CAG Gln	AAG Lys	AAC Asn	GGC Gly	ATC 11e 720	2160
AAG Lys	GTG Val	AAC Asn	TTC Phe	AAG Lys 725	ATC Ile	CGC Arg	CAC His	AAC Asn	ATC Ile 730	GAG Glu	GAC Asp	GGC Gly	AGC Ser	GTG Val 735	CAG Gln	2208
CTC Leu	GCC Ala	GAC Asp	CAC His 740	TAC Tyr	CAG Gln	CAG Gln	AAC Asn	ACC Thr 745	CCC Pro	ATC Ile	GGC Gly	GAC Asp	GGC Gly 750	CCC	GTG Val	2256
CTG Leu	CTG Leu	CCC Pro 755	GAC Asp	AAC Asn	CAC His	TAC Tyr	CTG Leu 760	Ser	ACC Thr	CAG Gln	TCC Ser	GCC Ala 765	Leu	AGC Ser	AAA Lys	2304
GAC Asp	CCC Pro 770	Asn	GAG Glu	AAG Lys	CGC	GAT Asp 775	His	ATG Met	GTC Val	CTG Leu	CTG Leu 780	Glu	TTC Phe	GTG Val	ACC Thr	2352
	Ala					Gly			GAG		Туг					2394

(2) INFORMATION FOR SEQ ID NO:141:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 797 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:141:

Met Asp Glu Leu Phe Pro Leu Ile Phe Pro Ala Glu Pro Ala Gln Ala 10 Ser Gly Pro Tyr Val Glu Ile Ile Glu Gln Pro Lys Gln Arg Gly Met 20 25 Arg Phe Arg Tyr Lys Cys Glu Gly Arg Ser Ala Gly Ser Ile Pro Gly 40 Glu Arg Ser Thr Asp Thr Thr Lys Thr His Pro Thr Ile Lys Ile Asn 55 Gly Tyr Thr Gly Pro Gly Thr Val Arg Ile Ser Leu Val Thr Lys Asp 70 Pro Pro His Arg Pro His Pro His Glu Leu Val Gly Lys Asp Cys Arg 90 85 Asp Gly Phe Tyr Glu Ala Glu Leu Cys Pro Asp Arg Cys Ile His Ser 105 100 Phe Gln Asn Leu Gly Ile Gln Cys Val Lys Lys Arg Asp Leu Glu Gln

		115					120					125			
Ala	Ile 130	Ser	Gln	Arg	Ile	Gln 135	Thr	Asn	Asn	Asn	Pro 140	Phe	Gln	Val	Pro
11e 145	Glu	Glu	Gln	Arg	Gly 150	Asp	Tyr	Asp	Leu	Asn 155	Ala	Val	Arg	Leu	Cys 160
Phe	Gln	Val	Thr	Val 165	Arg	Asp	Pro	Ser	Gly 170	Arg	Pro	Leu	Arg	Leu 175	Pro
Pro	Val	Leu	Pro 180	His	Pro	Ile	Phe	Asp 185	Asn	Arg	Ala	Pro	Asn 190	Thr	Ala
Glu	Leu	Lys 195	Ile	Суѕ	Arg	Val	Asn 200	Arg	Asn	Ser	Gly	Ser 205	Cys	Leu	Gly
Gly	Asp 210	Glu	Ile	Phe	Leu	Leu 215	Суѕ	Asp	Lys	Val	Gln 220	Lys	Glu	Asp	Ile
Glu 225	Val	Tyr	Phe	Thr	Gly 230	Pro	Gly	Trp	Glu	Ala 235	Arg	Gly	Ser	Phe	Ser 240
Gln	Ala	Asp	Val	His 245	Arg	Gln	Val	Ala	Ile 250	Val	Phe	Arg	Thr	Pro 255	Pro
_			260					265	Val				270		
_		275					280		Glu			285			_
	290	_				295			Ile		300			_	
305	•				310				Lys	315					320
				325				-	Arg 330					335	_
			340			_		345	Pro				350		
		355					360		Glu			365			
	370	_				375			Ala		380				
385					390				Ala	395					400
				G1n 405	Ala	Pro	Ala	Pro	Val	Pro	Val	Leu	Ala	Pro	GIY
	Pro	GIn				_	_		410	_	_		-1	415	~-
GIU	~3	(77)	420					425	Pro				430	Ala	_
3	_	435	420 Leu	Ser	Glu	Ala	Leu 440	425 Leu	Pro Gln	Leu	Gln	Phe 445	430 Asp	Ala Asp	Glu
_	Leu 450	435 Gly	420 Leu Ala	Ser Leu	Glu Leu	Ala Gly 455	Leu 440 Asn	425 Leu Ser	Pro Gln Thr	Leu Asp	Gln Pro 460	Phe 445 Ala	430 Asp Val	Ala Asp Phe	Glu
Asp 465	Leu 450 Leu	435 Gly Ala	420 Leu Ala Ser	Ser Leu Val	Glu Leu Asp 470	Ala Gly 455 Asn	Leu 440 Asn Ser	425 Leu Ser Glu	Pro Gln Thr Phe	Leu Asp Gln 475	Gln Pro 460 Gln	Phe 445 Ala Leu	430 Asp Val Leu	Ala Asp Phe Asn	Glu Thr Gln 480
Asp 465 Gly	Leu 450 Leu Ile	435 Gly Ala Pro	420 Leu Ala Ser Val	Ser Leu Val Ala 485	Glu Leu Asp 470 Pro	Ala Gly 455 Asn	Leu 440 Asn Ser Thr	425 Leu Ser Glu Thr	Pro Gln Thr Phe Glu 490	Leu Asp Gln 475 Pro	Gln Pro 460 Gln Met	Phe 445 Ala Leu Leu	430 Asp Val Leu Met	Ala Asp Phe Asn Glu 495	Glu Thr Gln 480 Tyr
Asp 465 Gly Pro	Leu 450 Leu Ile Glu	435 Gly Ala Pro	420 Leu Ala Ser Val Ile 500	Ser Leu Val Ala 485 Thr	Glu Leu Asp 470 Pro	Ala Gly 455 Asn His Leu	Leu 440 Asn Ser Thr	425 Leu Ser Glu Thr Thr 505	Pro Gln Thr Phe Glu 490 Gly	Leu Asp Gln 475 Pro	Gln Pro 460 Gln Met Gln	Phe 445 Ala Leu Leu	430 Asp Val Leu Met Pro 510	Ala Asp Phe Asn Glu 495 Pro	Glu Thr Gln 480 Tyr
Asp 465 Gly Pro	Leu 450 Leu Ile Glu Ala	435 Gly Ala Pro Ala Pro 515	420 Leu Ala Ser Val Ile 500 Ala	Ser Leu Val Ala 485 Thr	Glu Leu Asp 470 Pro Arg Leu	Ala Gly 455 Asn His Leu Gly	Leu 440 Asn Ser Thr Val Ala 520	425 Leu Ser Glu Thr Thr 505 Pro	Pro Gln Thr Phe Glu 490 Gly Gly	Leu Asp Gln 475 Pro Ala Leu	Gln Pro 460 Gln Met Gln Pro	Phe 445 Ala Leu Leu Arg Asn 525	430 Asp Val Leu Met Pro 510 Gly	Ala Asp Phe Asn Glu 495 Pro	Glu Thr Gln 480 Tyr Asp
Asp 465 Gly Pro Pro	Leu 450 Leu Ile Glu Ala Gly 530	Ala Pro Ala Pro 515 Asp	Ala Ser Val Ile 500 Ala Glu	Ser Leu Val Ala 485 Thr Pro	Glu Leu Asp 470 Pro Arg Leu	Ala Gly 455 Asn His Leu Gly Ser 535	Leu 440 Asn Ser Thr Val Ala 520 Ser	425 Leu Ser Glu Thr Thr 505 Pro	Pro Gln Thr Phe Glu 490 Gly Gly Ala	Leu Asp Gln 475 Pro Ala Leu Asp	Gln Pro 460 Gln Met Gln Pro Met 540	Phe 445 Ala Leu Leu Arg Asn 525 Asp	430 Asp Val Leu Met Pro 510 Gly	Ala Asp Phe Asn Glu 495 Pro Leu Ser	Glu Thr Gln 480 Tyr Asp Leu
Asp 465 Gly Pro Pro Ser Leu 545	Leu 450 Leu Ile Glu Ala Gly 530 Leu	A1a Pro A1a Pro 515 Asp	Ala Ser Val Ile 500 Ala Glu Gln	Ser Leu Val Ala 485 Thr Pro Asp Ile	Glu Leu Asp 470 Pro Arg Leu Phe Ser 550	Ala Gly 455 Asn His Leu Gly Ser 535 Ser	Leu 440 Asn Ser Thr Val Ala 520 Ser Leu	425 Leu Ser Glu Thr Thr 505 Pro Ile Asp	Pro Gln Thr Phe Glu 490 Gly Gly Ala Pro	Leu Asp Gln 475 Pro Ala Leu Asp Pro 555	Gln Pro 460 Gln Met Gln Pro Met Val	Phe 445 Ala Leu Leu Arg Asn 525 Asp	430 Asp Val Leu Met Pro 510 Gly Phe Thr	Ala Asp Phe Asn Glu 495 Pro Leu Ser Met	Glu Thr Gln 480 Tyr Asp Leu Ala Val 560
Asp 465 Gly Pro Pro Ser Leu 545 Ser	Leu 450 Leu Ile Glu Ala Gly 530 Leu Lys	435 Gly Ala Pro Ala Pro 515 Asp Ser Gly	420 Leu Ala Ser Val Ile 500 Ala Glu Gln	Ser Leu Val Ala 485 Thr Pro Asp Ile Glu 565	Glu Leu Asp 470 Pro Arg Leu Phe Ser 550 Leu	Ala Gly 455 Asn His Leu Gly Ser 535 Ser	Leu 440 Asn Ser Thr Val Ala 520 Ser Leu	425 Leu Ser Glu Thr Thr 505 Pro Ile Asp	Pro Gln Thr Phe Glu 490 Gly Gly Ala Pro Val 570	Leu Asp Gln 475 Pro Ala Leu Asp Pro 555 Val	Gln Pro 460 Gln Met Gln Pro Met 540 Val	Phe 445 Ala Leu Leu Arg Asn 525 Asp Ala Ile	430 Asp Val Leu Met Pro 510 Gly Phe Thr	Ala Asp Phe Asn Glu 495 Pro Leu Ser Met Val 575	Glu Thr Gln 480 Tyr Asp Leu Ala Val 560

			580					585					590			
		595	Ala			Gly	600					605				
	610					Pro 615					620					
	Gly	Val	Gln	Cys		Ser	Arg	Tyr	Pro	Asp 635	His	Met	Lys	Gln	His 640	
625 Asp	Phe	Phe	Lys	Ser 645	630 Ala	Met	Pro	Glu	Gly 650		Val	Gln	Glu	Arg 655		
Ile	Phe	Phe	Lys 660		Asp	Gly	Asn	Tyr 665		Thr	Arg	Ala	Glu 670	Val	Lys	
Phe	Glu	Gly 675		Thr	Leu	Val	Asn 680		Ile	Glu	Leu	Lys 685	Gly	Ile	Asp	
	690	Glu				Ile 695					700					
705	Ser				710	Ile				715					720	
Lys				725					730					735		
Leu	Ala	Asp	His 740	Tyr	Gln	Gln	Asn	Thr 745	Pro	Ile	Gly	Asp	Gly 750		Val	
Leu	Leu	Pro 755		Asn	His	Tyr	Leu 760		Thr	Gln	Ser	Ala 765		Ser	Lys	
Asp	Pro	Asn	Glu	Lys	Arg	Asp 775	His	Met	Val	Leu	Leu 780		Ph€	val	Thr	
Ala 785		Gly	Ile	Thr	Leu 790	Gly	Met	. Asp	Glu	Leu 795		Lys				
	((A) (B) (C) (D) (ii) (ix) (A) (I) (I) (I)	LEN TYF STR TOF MOLE FEAT A) NA 3) LC	GTH: PE: n LANDE POLOG CULE TURE: AME/I CATI	239 Sucle EDNES SY: 1 E TYPE SEY: SON: INFO	ACTE 4 ba ic a S: s inea PE: c Codi	se r cid ingl r DNA	oairs le Seque l	ence							
						SCRII										
ATO Med 1	G GTC	G AGO	C AAC	G GG(S Gly 5	GA(G GAG u Glu	ı Le	G TTO u Pho	e Th	C GG r Gl	G GTY y Va	G GT 1 Va	G CC 1 Pr	C AT o Il 15	C CTG e Lei	5 48 1
GT(Va)	C GAG	G CTY u Let	G GAG u Ası 20	C GG G G1	C GAG	C GTA	A AA l As	C GG n Gl _j 25	C CA	C AA s Ly	G TT s Ph	C AG e Se	C GT r Va 30	l Se	c GGG r Gly	96
GA ⁽	G GGG u Gl	C GA y Gl	G GG u Gl	y As	r GC p Al	C ACC	C TA r Ty 40	r Gl	C AA y Ly	G CT s Le	G AC u Th	C CT r Le 45	u Ly	AG TI 's Ph	C ATO	144 e
TG	C AC	C AC	c GG	c aa	G CT	G CC	C GI	e cc	C TG	G CC	C AC	C CI	'C G7	rg ac	C AC	C 192

Cys	Thr 50	Thr	Gly	Lys	Leu	Pro 55	Val	Pro	Trp	Pro	Thr 60	Leu	Val	Thr	Thr		
				_	CAG Gln 70											:	240
					AAG Lys						_			_	_		288
					AAG Lys							_			_		336
					GAC Asp												384
					GAC Asp												432
					AAC Asn 150												480
					TTC Phe												528
					CAC His										GGC Gly		576
															CTG Leu		624
															TTC Phe		672
															TCC Ser 240		720
															CCG Pro		768
														Glu	CAG Gln		816
													Gly		TCC Ser		864

GCG Ala	GGC Gly 290	AGC Ser	ATC Ile	CCA Pro	Gly	GAG Glu 295	AGG Arg	AGC Ser	ACA Thr	GAT Asp	ACC Thr 300	ACC Thr	AAG Lys	ACC Thr	CAC His	912
CCC Pro 305	ACC Thr	ATC Ile	AAG Lys	ATC Ile	AAT Asn 310	GGC Gly	TAC Tyr	ACA Thr	GGA Gly	CCA Pro 315	GGG Gly	ACA Thr	GTG Val	CGC Arg	ATC Ile 320	960
TCC Ser	CTG Leu	GTC Val	ACC Thr	AAG Lys 325	GAC Asp	CCT Pro	CCT Pro	CAC His	CGG Arg 330	CCT Pro	CAC His	CCC Pro	CAC His	GAG Glu 335	CTT Leu	1009
GTA Val	GGA Gly	AAG Lys	GAC Asp 340	TGC Cys	CGG Arg	GAT Asp	GGC Gly	TTC Phe 345	TAT Tyr	GAG Glu	GCT Ala	GAG Glu	CTC Leu 350	TGC Cys	CCG Pro	1056
GAC Asp	CGC Arg	TGC Cys 355	ATC Ile	CAC His	AGT Ser	TTC Phe	CAG Gln 360	AAC Asn	CTG Leu	GGA Gly	ATC Ile	CAG Gln 365	TGT Cys	GTG Val	AAG Lys	1104
AAG Lys	CGG Arg 370	Asp	CTG Leu	GAG Glu	CAG Gln	GCT Ala 375	ATC Ile	AGT Ser	CAG Gln	CGC Arg	ATC Ile 380	Gln	ACC Thr	AAC Asn	AAC Asn	1152
AAC Asn 385	Pro	TTC Phe	CAA Gln	GTT Val	CCT Pro 390	ATA Ile	GAA Glu	GAG Glu	CAG Gln	CGT Arg 395	Gly	GAC Asp	TAC Tyr	GAC Asp	CTG Leu 400	1200
AAT Asn	GCT Ala	GTG Val	CGG Arg	CTC Leu 405	Cys	TTC Phe	CAG Gln	GTG Val	ACA Thr 410	Val	CGG Arg	GAC JASP	CCA Pro	TCA Ser 415	GGC Gly	1248
AGG Arg	CCC Pro	CTC	CGC Arg 420	Leu	CCG Pro	CCT Pro	GTC Val	CT1 Leu 425	Pro	CAT His	CCC Pro	TATC	TT1 Phe 430	a Ası	AAT Asn	1296
CGT Arg	GCC Ala	2 CCC 2 Pro 435	Asn	ACT Thr	GCC Ala	GAG Glu	CTC Let 440	ı Lys	ATC	TG(C CGA	GTC G Val	l Ası	C CG	A AAC g Asn	1344
TCT Ser	GGC Gl ₂ 450	/ Sei	TGC Cys	CTC Lev	GGI Gly	GGG Gly 455	Asī	r GAC	G ATO	TTY Pho	CTZ e Lev 460	u Lev	ICY:	T GA	C AAG p Lys	1392
GTC Val	l Glr	G AA	A GAC	G GAC	AT1 0 Ile 470	e Glu	GT(G TAT	r TTC	e Th	r Gl	A CC	A GG o Gl	C TG y Tr	G GAG p Glu 480	1440
GC(C CGA	A GGG	C TCC y Se:	TT:	e Sei	G CAR	A GC'	T GA' a Ası	T GT p Va 49	l Hi	C CG s Ar	A CA g Gl:	A GT n Va	G GC 1 Al 49	C ATT a Ile	1488
GT Va	G TT	c cc e Ar	G AC g Th 50	r Pro	r CC0	С ТАС Э Туз	GC.	A GA a As 50	p Pr	C AG o Se	C CT	G CA u Gl	.G GC n Al 51	a Pr	CT GTG	1536
CG	T GT	C TC	C AT	G CA	G CT	G CG(G CG	G CC	T TC	C GA	'C CG	G GA	G CI	C AC	GT GAG	1584

Arg V		Ser 515	Met	Gln	Leu	Arg	Arg 520	Pro	Ser	Asp	Arg	Glu 525	Leu	Ser	Glu	
CCC A Pro M 5																1632
GAG G Glu G 545																1680
AAG A Lys S																1728
ATT G	_	_														1776
CAG C	Pro															1824
TTT C Phe P 6																1872
TTG G Leu A 625	_															1920
CCT G Pro A	_															1968
CCA G Pro V	_															2016
AAG C	ro '															2064
CTG C. Leu G																2112
GAC CO Asp P 705																2160
CAG C. Gln G	_															2208
CCC A																2256

GCC CAG AGG CCC CCC GAC CCA GCT CCT GCT CCA CTG GGG GCC CCG GGG

Ala Gln Arg Pro Pro Asp Pro Ala Pro Ala Pro Leu Gly Ala Pro Gly
755

CTC CCC AAT GGC CTC CTT TCA GGA GAT GAA GAC TTC TCC ATT GCG

Leu Pro Asn Gly Leu Leu Ser Gly Asp Glu Asp Phe Ser Ile Ala
770

GAC ATG GAC TTC TCA GCC CTG CTG AGT CAG ATC AGC TCC TAA

Asp Met Asp Phe Ser Ala Leu Leu Ser Gln Ile Ser Ser
790

2304

2304

2304

2304

(2) INFORMATION FOR SEQ ID NO:143:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 797 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:143:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 1 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 60 55 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 75 70 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 90 85 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 110 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 115 120 125 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 135 140 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 150 155 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 170 165 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 185 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 200 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 220 210 215 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 230 235 Gly Leu Arg Ser Arg Ala Met Asp Glu Leu Phe Pro Leu Ile Phe Pro

				245					250					255	
Ala	Glu	Pro	Ala 260	Gln	Ala	Ser	Gly	Pro 265	Tyr	Val	Glu	Ile	Ile 270	Glu	Gln
Pro	Lys	Gln 275	Arg	Gly	Met	Arg	Phe 280	Arg	Tyr	Lys	Cys	Glu 285	Gly	Arg	Ser
Ala	Gly 290	Ser	Ile	Pro	Gly	Glu 295	Arg	Ser	Thr	Asp	Thr 300	Thr	Lys	Thr	His
Pro 305	Thr	Ile	Lys	Ile	Asn 310	Gly	Tyr	Thr	Gly	Pro 315	Gly	Thr	Val	Arg	Ile 320
Ser	Leu	Val	Thr	Lys 325	Asp	Pro	Pro	His	Arg 330	Pro	His	Pro	His	Glu 335	Leu
Val	Gly	Lys	Asp 340	Cys	Arg	Asp	Gly	Phe	Tyr	Glu	Ala	Glu	Leu 350	Суѕ	Pro
		355					360			_		365	-	Val	-
_	370					375				_	380			Asn	
385					390					395				Asp	400
				405	_				410			•		Ser 415	-
			420					425					430	Asp	
		435					440	-		_	_	445		Arg	
	450					455	_				460		_	Asp	_
465					470					475	_		_	Trp Ala	480
Ald	Arg	GIY	ser	485	Ser	GIII	Ala	Asp	490	HIS	Arg	GIII	vaı	495	шe
1757	Dhe	ara	Thr		Dro	T 12	7 l a	700	Pro	502	T 011	Cln	77.		1107
			500	Pro				505					510	Pro	
Arg	Val	Ser 515	500 Met	Pro Gln	Leu	Arg	Arg 520	505 Pro	Ser	Asp	Arg	Glu 525	510 Leu	Pro Ser	Glu
Arg Pro	Val Met 530	Ser 515 Glu	500 Met Phe	Pro Gln Gln	Leu Tyr	Arg Leu 535	Arg 520 Pro	505 Pro Asp	Ser Thr	Asp Asp	Arg Asp 540	Glu 525 Arg	510 Leu His	Pro Ser Arg	Glu Ile
Arg Pro Glu 545	Val Met 530 Glu	Ser 515 Glu Lys	500 Met Phe Arg	Pro Gln Gln Lys	Leu Tyr Arg 550	Arg Leu 535 Thr	Arg 520 Pro Tyr	505 Pro Asp Glu	Ser Thr Thr	Asp Asp Phe 555	Arg Asp 540 Lys	Glu 525 Arg Ser	510 Leu His Ile	Pro Ser Arg Met	Glu Ile Lys 560
Arg Pro Glu 545 Lys	Val Met 530 Glu Ser	Ser 515 Glu Lys Pro	500 Met Phe Arg Phe	Pro Gln Gln Lys Ser 565	Leu Tyr Arg 550 Gly	Arg Leu 535 Thr	Arg 520 Pro Tyr	505 Pro Asp Glu Asp	Ser Thr Thr Pro 570	Asp Asp Phe 555 Arg	Arg Asp 540 Lys Pro	Glu 525 Arg Ser Pro	510 Leu His Ile Pro	Pro Ser Arg Met Arg 575	Glu Ile Lys 560 Arg
Arg Pro Glu 545 Lys Ile	Val Met 530 Glu Ser Ala	Ser 515 Glu Lys Pro Val	500 Met Phe Arg Phe Pro 580	Pro Gln Gln Lys Ser 565 Ser	Leu Tyr Arg 550 Gly Arg	Arg Leu 535 Thr Pro	Arg 520 Pro Tyr Thr	505 Pro Asp Glu Asp Ala 585	Ser Thr Thr Pro 570 Ser	Asp Asp Phe 555 Arg Val	Arg Asp 540 Lys Pro	Glu 525 Arg Ser Pro	510 Leu His Ile Pro Pro 590	Pro Ser Arg Met Arg 575 Ala	Glu Ile Lys 560 Arg
Arg Pro Glu 545 Lys Ile Gln	Val Met 530 Glu Ser Ala Pro	Ser 515 Glu Lys Pro Val Tyr 595	500 Met Phe Arg Phe Pro 580 Pro	Pro Gln Gln Lys Ser 565 Ser Phe	Leu Tyr Arg 550 Gly Arg	Arg Leu 535 Thr Pro Ser Ser	Arg 520 Pro Tyr Thr Ser Ser 600	505 Pro Asp Glu Asp Ala 585 Leu	Ser Thr Thr Pro 570 Ser Ser	Asp Phe 555 Arg Val	Arg Asp 540 Lys Pro Pro	Glu 525 Arg Ser Pro Lys Asn 605	510 Leu His Ile Pro Pro 590 Tyr	Pro Ser Arg Met Arg 575 Ala	Glu Ile Lys 560 Arg Pro
Arg Pro Glu 545 Lys Ile Gln Phe	Val Met 530 Glu Ser Ala Pro Pro 610	Ser 515 Glu Lys Pro Val Tyr 595 Thr	500 Met Phe Arg Phe Pro 580 Pro Met	Pro Gln Gln Lys Ser 565 Ser Phe Val	Leu Tyr Arg 550 Gly Arg Thr	Arg Leu 535 Thr Pro Ser Ser Pro 615	Arg 520 Pro Tyr Thr Ser 600 Ser	S05 Pro Asp Glu Asp Ala 585 Leu Gly	Ser Thr Thr Pro 570 Ser Ser	Asp Phe 555 Arg Val Thr	Arg Asp 540 Lys Pro Pro Ile Ser 620	Glu 525 Arg Ser Pro Lys Asn 605 Gln	510 Leu His Ile Pro 590 Tyr	Pro Ser Arg Met Arg 575 Ala Asp	Glu Ile Lys 560 Arg Pro Glu
Arg Pro Glu 545 Lys Ile Gln Phe Leu 625	Val Met 530 Glu Ser Ala Pro Pro 610 Ala	Ser 515 Glu Lys Pro Val Tyr 595 Thr	500 Met Phe Arg Phe Pro 580 Pro Met Ala	Pro Gln Gln Lys Ser 565 Ser Phe Val	Leu Tyr Arg 550 Gly Arg Thr Phe Pro 630	Arg Leu 535 Thr Pro Ser Pro 615 Gln	Arg 520 Pro Tyr Thr Ser 600 Ser Val	505 Pro Asp Glu Asp Ala 585 Leu Gly Leu	Ser Thr Thr Pro 570 Ser Ser Gln Pro	Asp Phe 555 Arg Val Thr Ile Gln 635	Arg Asp 540 Lys Pro Ile Ser 620 Ala	Glu 525 Arg Ser Pro Lys Asn 605 Gln	510 Leu His Ile Pro 590 Tyr Ala Ala	Pro Ser Arg Met Arg 575 Ala Asp Ser Pro	Glu Ile Lys 560 Arg Pro Glu Ala Ala 640
Arg Pro Glu 545 Lys Ile Gln Phe Leu 625 Pro	Val Met 530 Glu Ser Ala Pro 610 Ala	Ser 515 Glu Lys Pro Val Tyr 595 Thr	500 Met Phe Arg Phe Pro 580 Pro Met Ala	Pro Gln Gln Lys Ser 565 Ser Phe Val Pro Met 645	Leu Tyr Arg 550 Gly Arg Thr Phe Pro 630 Val	Arg Leu 535 Thr Pro Ser Ser Pro 615 Gln Ser	Arg 520 Pro Tyr Thr Ser 600 Ser Val	505 Pro Asp Glu Asp Ala 585 Leu Gly Leu	Ser Thr Thr Pro 570 Ser Gln Pro Ala 650	Asp Phe 5555 Arg Val Thr Ile Gln 635 Gln	Arg Asp 540 Lys Pro Ile Ser 620 Ala	Glu 525 Arg Ser Pro Lys Asn 605 Gln Pro	510 Leu His Ile Pro 590 Tyr Ala Ala	Pro Ser Arg Met Arg 575 Ala Asp Ser Pro 655	Glu Ile Lys 560 Arg Pro Glu Ala Ala 640 Val
Arg Pro Glu 545 Lys Ile Gln Phe Leu 625 Pro	Val Met 530 Glu Ser Ala Pro 610 Ala Ala Val	Ser 515 Glu Lys Pro Val Tyr 595 Thr Pro Pro	500 Met Phe Arg Phe Pro 580 Pro Met Ala Ala Ala 660	Pro Gln Gln Lys Ser 565 Ser Phe Val Pro Met 645 Pro	Leu Tyr Arg 550 Gly Arg Thr Phe Pro 630 Val	Arg Leu 535 Thr Pro Ser Pro 615 Gln Ser Pro	Arg 520 Pro Tyr Thr Ser 600 Ser Val Ala Pro	505 Pro Asp Glu Asp Ala 585 Leu Gly Leu Leu Gln 665	Ser Thr Thr Pro 570 Ser Gln Pro Ala 650 Ala	Asp Phe 555 Arg Val Thr Ile Gln 635 Gln Val	Arg Asp 540 Lys Pro Ile Ser 620 Ala Ala	Glu 525 Arg Ser Pro Lys Asn 605 Gln Pro Pro	510 Leu His Ile Pro 590 Tyr Ala Ala Ala Pro 670	Pro Ser Arg Met Arg 575 Ala Asp Ser Pro 655 Ala	Glu Ile Lys 560 Arg Pro Glu Ala 640 Val
Arg Pro Glu 545 Lys Ile Gln Phe Leu 625 Pro Pro	Val Met 530 Glu Ser Ala Pro 610 Ala Ala Val	Ser 515 Glu Lys Pro Val Tyr 595 Thr Pro Leu Thr 675	500 Met Phe Arg Phe Pro 580 Pro Met Ala Ala 660 Gln	Pro Gln Gln Lys Ser 565 Ser Phe Val Pro Met 645 Pro Ala	Leu Tyr Arg 550 Gly Arg Thr Phe Pro 630 Val Gly	Arg Leu 535 Thr Pro Ser Pro 615 Gln Ser Pro Glu	Arg 520 Pro Tyr Thr Ser 600 Ser Val Ala Pro 630 680	505 Pro Asp Glu Asp Ala 585 Leu Gly Leu Gln 665 Thr	Ser Thr Thr Pro 570 Ser Gln Pro Ala 650 Ala Leu	Asp Phe 5555 Arg Val Thr Ile Gln 635 Gln Val Ser	Arg Asp 540 Lys Pro Ile Ser 620 Ala Ala Glu	Glu 525 Arg Ser Pro Lys Asn 605 Gln Pro Pro Ala 685	510 Leu His Ile Pro 590 Tyr Ala Ala Pro 670 Leu	Pro Ser Arg Met Arg 575 Ala Asp Ser Pro 655 Ala Leu	Glu Ile Lys 560 Arg Pro Glu Ala 640 Val Pro Gln
Arg Pro Glu 545 Lys Ile Gln Phe Leu 625 Pro Pro Lys Leu	Val Met 530 Glu Ser Ala Pro 610 Ala Ala Val Pro Gln 690	Ser 515 Glu Lys Pro Val Tyr 595 Thr Pro Leu Thr 675 Phe	500 Met Phe Arg Phe Pro 580 Pro Met Ala Ala 660 Gln Asp	Pro Gln Gln Lys Ser 565 Ser Phe Val Pro Met 645 Pro Ala Asp	Leu Tyr Arg 550 Gly Arg Thr Phe Pro 630 Val Gly Gly Glu	Arg Leu 535 Thr Pro Ser Ser Pro 615 Gln Ser Pro Glu Asp 695	Arg 520 Pro Tyr Thr Ser 600 Ser Val Ala Pro 680 Leu	505 Pro Asp Glu Asp Ala 585 Leu Gly Leu Gln 665 Thr	Ser Thr Thr Pro 570 Ser Gln Pro Ala 650 Ala Leu Ala	Asp Phe 5555 Arg Val Thr Ile Gln 635 Gln Val Ser Leu	Arg Asp 540 Lys Pro Ile Ser 620 Ala Ala Glu Leu 700	Glu 525 Arg Ser Pro Lys Asn 605 Gln Pro Pro Ala 685 Gly	510 Leu His Ile Pro 590 Tyr Ala Ala Pro 670 Leu Asn	Pro Ser Arg Met Arg 575 Ala Asp Pro 655 Ala Leu Ser	Glu Ile Lys 560 Arg Pro Glu Ala 640 Val Pro Gln

705					710					715					720
Gln	Gln	Leu	Leu	Asn 725	Gln	Gly	Ile		Val 730	Ala	Pro	His	Thr	Thr 735	Glu
Pro	Met	Leu	Met 740	Glu	Tyr	Pro	Glu	Ala 745	Ile	Thr	Arg	Leu	Val 750	Thr	Gly
Ala	Gln	Arg 755	Pro	Pro	Asp	Pro	Ala 760	Pro	Ala	Pro	Leu	Gly 765	Ala	Pro	Gly
Leu	Pro 770	Asn	Gly	Leu	Leu	Ser 775	Gly	Asp	Glu	Asp	Phe 780	Ser	Ser	Ile	Ala
Asp 785	Met	Asp	Phe	Ser	Ala 790	Leu	Leu	Ser	Gln	Ile 795	Ser	Ser			

(2) INFORMATION FOR SEQ ID NO:144:

151	SECTENCE	CHARACTERISTICS:
(1)	SECUENCE	CHARACIERISITOS.

- (A) LENGTH: 3381 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...3378
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:144:

			AGC Ser						4	8
			CAG Gln						9	6
			GAC Asp						14	4
			AAT Asn 55						19	92
			GGT Gly						24	10
								ACA Thr	28	88

90

85

290

295

CCA ACC AGG AAA ATC TCT GCC TCT GAA TTT GAC CGG CCT CTT AGA CCC Pro Thr Arg Lys Ile Ser Ala Ser Glu Phe Asp Arg Pro Leu Arg Pro 100 105 ATT GTT GTC AAG GAT TCT GAG GGA ACT GTG AGC TTC CTC TCT GAC TCA 384 Ile Val Val Lys Asp Ser Glu Gly Thr Val Ser Phe Leu Ser Asp Ser 115 120 GAA AAG AAG GAA CAG ATG CCT CTA ACC CCT CCA AGG TTT GAT CAT GAT 432 Glu Lys Lys Glu Gln Met Pro Leu Thr Pro Pro Arg Phe Asp His Asp 130 135 GAA GGG GAC CAG TGC TCA AGA CTC TTG GAA TTA GTG AAG GAT ATT TCT 480 Glu Gly Asp Gln Cys Ser Arg Leu Leu Glu Leu Val Lys Asp Ile Ser 155 AGT CAT TTG GAT GTC ACA GCC TTA TGT CAC AAA ATT TTC TTG CAT ATC 528 Ser His Leu Asp Val Thr Ala Leu Cys His Lys Ile Phe Leu His Ile 165 170 CAT GGA CTG ATA TCT GCT GAC CGC TAT TCC CTG TTC CTT GTC TGT GAA 576 His Gly Leu Ile Ser Ala Asp Arg Tyr Ser Leu Phe Leu Val Cys Glu 180 185 GAC AGC TCC AAT GAC AAG TTT CTT ATC AGC CGC CTC TTT GAT GTT GCT 624 Asp Ser Ser Asn Asp Lys Phe Leu Ile Ser Arg Leu Phe Asp Val Ala 195 200 GAA GGT TCA ACA CTG GAA GAA GTT TCA AAT AAC TGT ATC CGC TTA GAA 672 Glu Gly Ser Thr Leu Glu Glu Val Ser Asn Asn Cys Ile Arg Leu Glu 210 TGG AAC AAA GGC ATT GTG GGA CAT GTG GCA GCG CTT GGT GAG CCC TTG 720 Trp Asn Lys Gly Ile Val Gly His Val Ala Ala Leu Gly Glu Pro Leu AAC ATC AAA GAT GCA TAT GAG GAT CCT CGG TTC AAT GCA GAA GTT GAC Asn Ile Lys Asp Ala Tyr Glu Asp Pro Arg Phe Asn Ala Glu Val Asp CAA ATT ACA GGC TAC AAG ACA CAA AGC ATT CTT TGT ATG CCA ATT AAG 816 Gln Ile Thr Gly Tyr Lys Thr Gln Ser Ile Leu Cys Met Pro Ile Lys 260 265 AAT CAT AGG GAA GAG GTT GTT GGT GTA GCC CAG GCC ATC AAC AAA 864 Asn His Arg Glu Glu Val Val Gly Val Ala Gln Ala Ile Asn Lys Lys 275 280 TCA GGA AAC GGT GGG ACA TTT ACT GAA AAA GAT GAA AAG GAC TTT GCT Ser Gly Asn Gly Gly Thr Phe Thr Glu Lys Asp Glu Lys Asp Phe Ala

GCT Ala 305	TAT Tyr	TTG Leu	GCA Ala	TTT Phe	TGT Cys 310	GGT Gly	ATT Ile	GTT Val	CTT Leu	CAT His 315	AAT Asn	GCT Ala	CAG Gln	CTC Leu	TAT Tyr 320	960
									AAT Asn 330							1008
									TCA Ser							1056
									ATG Met							1104
												Ser			TTT Phe	1152
	Met										Asp				AGG Arg 400	1200
					Lys					Тут					: AAA . Lys	1248
AAT Asr	ACT Thr	ATC Met	GAA Glu 420	Pro	CTT Leu	AAT Asn	ATC	CCA Pro 425	Asp	GTC Val	AGT Ser	AAC Lys	G GAT S Asp 430	Lys	A AGA S Arg	1296
			Thr					c Gly					n Glr		C ATT	1344
		Le					Ile					s Ly			A GTT s Val	1392
AT: 110 46	e Gly	GT' / Va	T TG(l Cy:	C CAA	A CTC 1 Let 470	ı Val	AA' Asi	T AAG	G ATY	G GA(t Gl) 47!	u Gl	G AA u As	T AC n Th	T GG r Gl	C AAG y Lys 480	1440
GT Va	T AAG	G CC	T TTO	C AA(e As: 48!	n Arg	AAA BAA E	r Gad	C GAI	A CAG u Gli 49	n Ph	T CT e Le	G GA u Gl	A GC u Al	T TT a Ph 49	T GTC e Val 5	1488
AT Il	C TT e Ph	r TG e Cy	T GG s Gl 50	y Le	G GGG u Gl	G ATY	C CA	G AA n As 50	n Th	G CA r Gl	G AT n Me	G TA	T GA T Gl 51	u Al	A GTG a Val	1536
GA G1	G AG u Ar	A GC g Al	C AT a Me	G GC	C AA a Ly	G CA s Gl:	A AT n Me	G GT t Va	C AC	A TT	rG GA eu Gl	kG GT Lu Vā	T CI	G TO	G TAT	1584

520 525 515 CAT GCT TCA GCA GCA GAG GAA GAA ACA AGA GAG CTA CAG TCG TTA GCG His Ala Ser Ala Ala Glu Glu Glu Thr Arg Glu Leu Gln Ser Leu Ala 535 530 GCT GCT GTG GTG CCA TCT GCC CAG ACC CTT AAA ATT ACT GAC TTT AGC 1680 Ala Ala Val Val Pro Ser Ala Gln Thr Leu Lys Ile Thr Asp Phe Ser 550 555 TTC AGT GAC TTT GAG CTG TCT GAT CTG GAA ACA GCA CTG TGC ACA ATT 1728 Phe Ser Asp Phe Glu Leu Ser Asp Leu Glu Thr Ala Leu Cys Thr Ile 565 CGG ATG TTT ACT GAC CTC AAC CTT GTG CAG AAC TTC CAG ATG AAA CAT 1776 Arg Met Phe Thr Asp Leu Asn Leu Val Gln Asn Phe Gln Met Lys His 585 GAG GTT CTT TGC AGA TGG ATT TTA AGT GTT AAG AAG AAT TAT CGG AAG 1824 Glu Val Leu Cys Arg Trp Ile Leu Ser Val Lys Lys Asn Tyr Arg Lys 595 600 AAT GTT GCC TAT CAT AAT TGG AGA CAT GCC TTT AAT ACA GCT CAG TGC 1872 Asn Val Ala Tyr His Asn Trp Arg His Ala Phe Asn Thr Ala Gln Cys 615 610 ATG TTT GCT GCT CTA AAA GCA GGC AAA ATT CAG AAC AAG CTG ACT GAC Met Phe Ala Ala Leu Lys Ala Gly Lys Ile Gln Asn Lys Leu Thr Asp 630 CTG GAG ATA CTT GCA TTG CTG ATT GCT GCA CTA AGC CAC GAT TTG GAT 1968 Leu Glu Ile Leu Ala Leu Leu Ile Ala Ala Leu Ser His Asp Leu Asp 650 645 CAC CGT GGT GTG AAT AAC TCT TAC ATA CAG CGA AGT GAA CAT CCA CTT 2016 His Arg Gly Val Asn Asn Ser Tyr Ile Gln Arg Ser Glu His Pro Leu 665 GCC CAG CTT TAC TGC CAT TCA ATC ATG GAA CAC CAT CAT TTT GAC CAG 2064 Ala Gln Leu Tyr Cys His Ser Ile Met Glu His His His Phe Asp Gln 680 675 TGC CTG ATG ATT CTT AAT AGT CCA GGC AAT CAG ATT CTC AGT GGC CTC 2112 Cys Leu Met Ile Leu Asn Ser Pro Gly Asn Gln Ile Leu Ser Gly Leu 700 695 690 TCC ATT GAA GAA TAT AAG ACC ACG TTG AAA ATA ATC AAG CAA GCT ATT 2160

Ser Ile Glu Glu Tyr Lys Thr Thr Leu Lys Ile Ile Lys Gln Ala Ile

TTA GCT ACA GAC CTA GCA CTG TAC ATT AAG AGG CGA GGA GAA TTT TTT Leu Ala Thr Asp Leu Ala Leu Tyr Ile Lys Arg Arg Gly Glu Phe Phe

730

710

725

					AAT Asn											2256
					ATG Met											2304
					ATT Ile											2352
					GGA Gly 790											2400
					AAC Asn											2448
														Ala	CTG Leu	2496
			Ser												Lys	2544
		Gln										Glu			CTG Leu	2592
Ile 865	Asn	Gly	Glu	Ser	Gly 870	Gln	Ala	Lys	Arg	875	Trp	Val	Pro	Arg	G GCC g Ala 880	2640
Arg	Asp	Pro	Pro	Val 885	Ala	Thr	Met	. Val	Ser 890	Lys	s Gly	, Glu	ı Glu	1 Let 895		2688
Thi	: Gly	/ Val	900	Pro) Ile	e Leu	ı Val	. Glu 905	Leu S	ı Ası	ο Gly	/ Asp	910	l Ası O	G GGC n Gly	2736
			e Ser					Gl)					a Th		c GGC r Gly	2784
		נלT נ					e Cys					s Le			G CCC 1 Pro	2832
															C AGC e Ser	2880

945					950					955					960	
						AAG										2928
Arg	Tyr	Pro	Asp	His 965	Met	Lys	Gln	His	Asp 970	Phe	Phe	Lys	Ser	Ala 975	Met	
						GAG										2976
Pro	Glu	Gly	7yr 980	Val	Gln	Glu	Arg	Thr 985	Ile	Phe	Phe	Lys	Asp 990	Asp	Gly	
						GAG										3024
Asn	Tyr	Lys 995	Thr	Arg	Ala	Glu	Val 1000	Lys	Phe	Glu		qzA 2001	Thr	Leu	Val	
		,,,,				•	.000					1003				
						GGC										3072
	Arg 1010	Ile	Glu	Leu		Gly 1015	Ile	Asp	Phe			Asp	Gly	Asn	Ile	
	1010					1013				•	1020					
CTG	GGG	CAC	AAG	CTG	GAG	TAC	AAC	TAC	AAC	AGC	CAC	AAC	GTC	TAT	OTA	3120
	Gly	His	Lys			Tyr	Asn	Tyr	Asn	Ser	His	Asn	Val	Tyr	Ile	
1025					1030				1	1035					1040	
ATG	GCC	GAC	AAG	CAG	AAG	AAC	GGC	ATC	AAG	GTG	AAC	TTC	AAG	ATC	CGC	3168
						Asn										
			1	1045				1	1050				:	1055		
CAC	AAC	OTA	GAG	GAC	GGC	AGC	GTG	CAG	CTC	GCC	GAC	CAC	TAC	CAG	CAG	3216
						Ser										5525
		1	060				1	1065				1	070			
AAC	ACC	CCC	ATC	GGC	GAC	GGC	CCC	GTG	CTG	CTG	CCC	GAC	AAC	CAC	TAC	3264
						Gly										
	-	1075				1	080				1	.085				
CTG	AGC	ACC	CAG	TCC	GCC	CTG	AGC	AAA	GAC	CCC	AAC	GAG	AAG	CGC	GAT	3312
						Leu										
1	1090				3	1095				1	100			,		
CAC	ATG	GTC	CTG	CTG	GAG	TTC	GTG	ACC	GCC	GCC	GGG	ATC	ACT	CTC	GGC	3360
						Phe										
1105				1	110				1	115				J	120	
ATG	GAC	GAG	CTG	TAC	AAG	TAA										3381
		Glu														2201
			1	125												

(2) INFORMATION FOR SEQ ID NO:145:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1126 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein
 (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:145:

														_	
Met	Glu	Arg	Ala	Gly 5	Pro	Ser	Phe	Gly	Gln 10	Gln	Arg	Gln	Gln	Gln 15	Gln
1 Pro	Gln	Gln	Gln		Gln	Gln	Gln	Ara		Gln	Asp	Ser	Val		Ala
FIC	GIII	GIII	20	טעם	0111	01	01	25					30		
Trp	Leu	Asp		His	Trp	Asp	Phe	Thr	Phe	Ser	Tyr	Phe	Val	Arg	Lys
•		35	=		_		40					45			
Ala	Thr	Arg	Glu	Met	Val	Asn	Ala	Trp	Phe	Ala	Glu	Arg	Val	His	Thr
	50					55					60				
Ile	Pro	Val	Cys	Lys		Gly	Ile	Arg	Gly	His	Thr	Glu	Ser	Cys	
65	_	_		-1	70	D	3		>	75	C~~	1101	Dro	C114	80 Thr
Cys	Pro	Leu	Gin	61n 85	Ser	Pro	Arg	Ala	90	Asn	Set	vai	PIO	95	1111
Pro	Thr	Δra	T.vs		Ser	Δla	Ser	Glu		Asp	Ara	Pro	Leu		Pro
FIU	1111	n g	100	110	001			105					110	_	
Ile	Val	Val		Asp	Ser	Glu	Gly		Val	Ser	Phe	Leu	Ser	Asp	Ser
		115	-	_			120					125			
Glu	Lys	Lys	Glu	Gln	Met	Pro	Leu	Thr	Pro	Pro	Arg	Phe	Asp	His	Asp
	130					135					140				
Glu	Gly	Asp	Gln	Суѕ			Leu	Leu	Glu	Leu	Val	Lys	Asp	Ile	
145		_			150			۵	114.0	155	T10	Dho	Lou	n; c	160
Ser	His	Leu	Asp	165		Ala	Leu	Cys	170	Lys	116	PHE	Leu	175	116
uic	Gly	Leu	Tle			Asn	Ara	Tyr			Phe	Leu	Val		Glu
nis	Gly	Deu	180		7.14	ı	9	185			•		190	-	
Asp	Ser	Ser			Lys	Phe	Leu	Ilė	Ser	Arg	Leu	Phe	Asp	Val	Ala
		195					200					205			
Glu	Gly	Ser	Thr	Leu	Glu	Glu	Val	Ser	Asn	Asn	Cys	Ile	Arg	Leu	Glu
	210					215		_			220		01	5	•
		Lys	Gly	Ile			His	: Val	Ala			G17	GIU	Pro	Leu 240
225		7	7.00	. או	230		700	Dro) N T C	235		ء ۵۱	Glu	Val	Asp
ASN	116	Lys	Asp	245		GIU	ASL	FIC	250		. ASI	7,20	. 010	255	
Gln	Ile	Thr	Glv			Thr	Gln	. Ser			Cys	Met	Pro		Lys
02			260		-			265					270		
Asn	His	Arg	Glu	Glu	Va]	Val	. Gly	/ Val	Ala	a Glr	. Ala	Ile	e Asn	Lys	Lys
		275					280					285			
Ser	Gly	Asr	Gly	, Gl	Thi	Phe	Thr	Glu	ı Lys	s Asp			Asp	Ph∈	Ala
	290					295			_		300		- G1		
		Leu	ı Ala	a Ph∈			/ Ile	∘ Val	Let			1 Ala	a Gir	Leu	320
305					310			- N×0	, her	315		Lei	ı T.ei	ı Asr	
GIU	Tnr	ser	. Let	1 Let 325		ASI	ı riys	· WIC	33(. ve.	. 100	ع عاد د	335	Leu
212	Ser	T.et	1 716			ıGlı	ıGlr	n Glr			ı Glı	ı Vai	l Ile		ı Lys
VIG			340					345					350		-
Lys	: Ile	a Ala			: Ile	e Ile	e Sei			t Glr	n Vai	Gl:	n Lys	суя	5 Thr
		355	5				360	О				36	5		
Il∈	Phe	e Ile	e Val	l Ası	o Glu	ı Ası	o Cys	s Se	r Ası	p Se	r Phe	e Se	r Sei	r Val	l Phe
	370)				375	5				380)			

His 385	Met	Glu	Cys	Glu	Glu 390	Leu	Glu	Lys	Ser	Ser 395	Asp	Thr	Leu	Thr	Arg 400
	His	Asp	Ala	Asn 405		Ile	Asn	Tyr	Met 410	Tyr	Ala	Gln	Tyr	Val 415	Lys
Asn	Thr	Met	Glu 420		Leu	Asn	Ile	Pro	Asp	Val	Ser	Lys	Asp 430	Lys	Arg
Phe	Pro	Trp		Thr	Glu	Asn	Thr	Gly	Asn	Val	Asn	Gln 445	Gln	Суѕ	Ile
Arg	Ser		Leu	Cys	Thr	Pro	Ile	Lys	Asn	Gly	Lys 460	Lys	Asn	Lys	Val
Ile 465		Val	Суѕ	Gln	Leu 470		Asn	Lys	Met	Glu 475	Glu	Asn	Thr	Gly	Lys 480
	Lys	Pro	Phe	Asn 485	Arg	Asn	Asp	Glu	Gln 490	Phe	Leu	Glu	Ala	Fhe 495	Val
Ile	Phe	Cys	Gly 500	Leu	Gly	Ile	Gln	Asn 505	Thr	Gln	Met	Tyr	Glu 510	Ala	Val
Glu	Arg	Ala 515	Met	Ala	Lys	Gln	Met 520	Val	Thr	Leu	Glu	Val 525	Leu	Ser	Tyr
His	Ala 530		Ala	Ala	Glu	Glu 535	Glu	Thr	Arg	Glu	Leu 540	Gln	Ser	Leu	Ala
545					550		Gln			555					560
				565			Asp		570					575	
			580				Leu	585					590		
		595					Leu 600					605			
	610					615					620				
625					630					635					Asp 640
				645					650					655	
			660					665					670)	Leu
		675	•				680					685	i		Gln
	690)				695	,				700				Leu
705	·)				710	1				715					720 Phe
				725	,				730)				735	
			740)				745	•				750)	a Ile
		755	5				760)				765	5		a Thr
	770)				775	5				780)			e Glu
785	5				790)				795	·				800 r Met
P.T.(للللار	, wal	اعبدر	805			, 010		810					81	

Gln Val Gly Phe Ile Asp Ala Ile Cys Leu Gln Leu Tyr Glu Ala Leu 820 825 830 Thr His Val Ser Glu Asp Cys Phe Pro Leu Leu Asp Gly Cys Arg Lys 840 845 Asn Arg Gln Lys Trp Gln Ala Leu Ala Glu Gln Gln Glu Lys Met Leu 850 855 Ile Asn Gly Glu Ser Gly Gln Ala Lys Arg Asn Trp Val Pro Arg Ala 865 870 875 880 Arg Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu Glu Leu Phe 890 885 Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly 905 900 His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly 925 920 Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro 930 935 Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser 945 950 955 Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met 965 970 Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly 980 985 Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val 995 1000 1005 Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile 1010 1015 1020 Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile 1030 1035 1040 Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Fhe Lys Ile Arg 1045 1050 1055 His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln 1065 1070 1060 Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr 1080 1085 1075 Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp 1090 1095 1100 His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly 1110 1115 Met Asp Glu Leu Tyr Lys 1125

(2) INFORMATION FOR SEQ ID NO:146:

- (i) SEOUENCE CHARACTERISTICS:
 - (A) LENGTH: 2760 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...2757

(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:146:

_		_					GAG Glu 15	 48	
							GTG Val	96	
							CCC Pro	144	
							CAG Gln	192	
							CAT His	240	
							TCC Ser 95	288	
							AGC Ser	 336	
							CAC His	384	
							TGC Cys	432	
							CGC Arg	480	
							GTC Val 175	528	
							CTG Leu	576	
							GAG Glu	624	:

200 195 AAA CAG AAG ACC AAA ACC ATC AAA TGC TCC CTC AAC CCT GAG TGG AAT 672 Lys Gln Lys Thr Lys Thr Ile Lys Cys Ser Leu Asn Pro Glu Trp Asn 215 210 720 GAG ACA TTT AGA TTT CAG CTG AAA GAA TCG GAC AAA GAC AGA AGA CTG Glu Thr Phe Arg Phe Gln Leu Lys Glu Ser Asp Lys Asp Arg Arg Leu 235 230 TCA GTA GAG ATT TGG GAT TGG GAT TTG ACC AGC AGG AAT GAC TTC ATG 768 Ser Val Glu Ile Trp Asp Trp Asp Leu Thr Ser Arg Asn Asp Phe Met GGA TCT TTG TCC TTT GGG ATT TCT GAA CTT CAG AAG GCC AGT GTT GAT 816 Gly Ser Leu Ser Phe Gly Ile Ser Glu Leu Gln Lys Ala Ser Val Asp 864 GGC TGG TTT AAG TTA CTG AGC CAG GAG GAA GGC GAG TAC TTC AAT GTG Gly Trp Phe Lys Leu Leu Ser Gln Glu Glu Gly Glu Tyr Phe Asn Val 280 275 912 CCT GTG CCA CCA GAA GGA AGT GAG GCC AAT GAA GAA CTG CGG CAG AAA Pro Val Pro Pro Glu Gly Ser Glu Ala Asn Glu Glu Leu Arg Gln Lys 295 290 TIT GAG AGG GCC AAG ATC AGT CAG GGA ACC AAG GTC CCG GAA GAA AAG 960 Phe Glu Arg Ala Lys Ile Ser Gln Gly Thr Lys Val Pro Glu Glu Lys 310 315 305 ACG ACC AAC ACT GTC TCC AAA TTT GAC AAC AAT GGC AAC AGA GAC CGG 1008 Thr Thr Asn Thr Val Ser Lys Phe Asp Asn Asn Gly Asn Arg Asp Arg 325 ATG AAA CTG ACC GAT TTT AAC TTC CTA ATG GTG CTG GGG AAA GGC AGC 1056 Met Lys Leu Thr Asp Phe Asn Phe Leu Met Val Leu Gly Lys Gly Ser TTT GGC AAG GTC ATG CTT TCA GAA CGA AAA GGC ACA GAT GAG CTC TAT 1104 Phe Gly Lys Val Met Leu Ser Glu Arg Lys Gly Thr Asp Glu Leu Tyr 360 355 GCT GTG AAG ATC CTG AAG AAG GAC GTT GTG ATC CAA GAT GAT GAC GTG Ala Val Lys Ile Leu Lys Lys Asp Val Val Ile Gln Asp Asp Val 375 GAG TGC ACT ATG GTG GAG AAG CGG GTG TTG GCC CTG CCT GGG AAG CCG 1200 Glu Cys Thr Met Val Glu Lys Arg Val Leu Ala Leu Pro Gly Lys Pro 395 390 385 CCC TTC CTG ACC CAG CTC CAC TCC TGC TTC CAG ACC ATG GAC CGC CTG Pro Phe Leu Thr Gln Leu His Ser Cys Phe Gln Thr Met Asp Arg Leu

410

									CTC Leu					1296
									GTA Val					1344
									AAG Lys 460					1392
									TCT Ser					1440
									ATC Ile					1488
													ATA Ile	1536
											Ala		GGA Gly	1584
										Phe			GAG Glu	1632
Glu									His				TAT Tyr 560	1680
				Lys				Ile					ATG Met	1728
			Gly				Cys					/ Glu	A CGT 1 Arg	1776
		Glu				e Arg					o Glu		A CTT s Leu	1824
	Lys				Pro					s Ala			G CGA y Arg	1872
													C CTA l Leu	1920

625				630					635					640	
							AGG Arg								1968
							GAA Glu 665								2016
							ATG Met								2064
							GTC Val								2112
							GAG Glu								2160
							TGC Cys							GTG Val	2208
													Cys	TTC Phe	2256
							Gln					Lys		GCC	2304
						Glu					Phe			GAC Asp	2352
	Asn				Ala					e Glu				CTG Leu 800	2400
				Leu					Phe					AAC Asn	2448
			Lys					Туг					ı Val	TAT L Tyr	2496
		Asp					n Gly					n Phe		G ATC	2544

				GTG Val					2592
				CCC Pro					2640
				AGC Ser					2688
								CTC Leu	2736
		TAC Tyr	TAA						2760

(2) INFORMATION FOR SEQ ID NO:147:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 919 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:147:

Met Ala Asp Pro Ala Ala Gly Pro Pro Pro Ser Glu Gly Glu Glu Ser 10 5 Thr Val Arg Phe Ala Arg Lys Gly Ala Leu Arg Gln Lys Asn Val His Glu Val Lys Asn His Lys Phe Thr Ala Arg Phe Phe Lys Gln Pro Thr 40 Phe Cys Ser His Cys Thr Asp Phe Ile Trp Gly Phe Gly Lys Gln Gly 55 Phe Gln Cys Gln Val Cys Cys Phe Val Val His Lys Arg Cys His Glu 75 70 Phe Val Thr Phe Ser Cys Pro Gly Ala Asp Lys Gly Pro Ala Ser Asp 85 90 Asp Pro Arg Ser Lys His Lys Phe Lys Ile His Thr Tyr Ser Ser Pro 100 105 110 Thr Phe Cys Asp His Cys Gly Ser Leu Leu Tyr Gly Leu Ile His Gln 125 120 Gly Met Lys Cys Asp Thr Cys Met Met Asn Val His Lys Arg Cys Val 135 140 Met Asn Val Pro Ser Leu Cys Gly Thr Asp His Thr Glu Arg Arg Gly 160 150 155

Arg	Ile	Tyr	Ile	Gln 165	Ala	His	Ile	Asp	Arg 170	Asp	Val	Leu	Ile	Val 175	Leu
Val .	Arg	qzA	Ala 180		Asn	Leu	Val	Pro 185		Asp	Pro	Asn	Gly 190	Leu	Ser
Asp	Pro	Tyr 195	Val	Lys	Leu	Lys	Leu 200	Ile	Pro	Asp	Pro	Lys 205	Ser	Glu	Ser
	210		Thr			215					220				
Glu 225	Thr	Phe	Arg	Phe	Gln 230	Leu	Lys	Glu	Ser	Asp 235	Lys	Asp	Arg	Arg	Leu 240
	Val	Glu	Ile	Trp 245		Trp	Asp	Leu	Thr 250	Ser	Arg	Asn	Asp	Phe 255	Met
Gly	Ser	Leu	Ser 260		Gly	Ile	Ser	Glu 265	Leu	Gln	Lys	Ala	Ser 270	Val	Asp
Gly	Trp	Phe 275	Lys	Leu	Leu	Ser	Gln 280	Glu	Glu	Gly	Glu	Tyr 285	Phe	Asn	Val
Pro	Val 290	Pro	Pro	Glu	Gly	Ser 295	Glu	Ala	Asn	Glu	Glu 300	Leu	Arg	Gln	Lys
Phe 305	Glu	Arg	Ala	Lys	Ile 310	Ser	Gln	Gly	Thr	Lys 315	Val	Pro	Glu	Glu	Lys 320
Thr	Thr	Asn	Thr	Val 325		Lys	Phe	Asp	Asn 330	Asn	Gly	Asn	Arg	Asp 335	Arg
Met	Lys	Leu	Thr 340	Asp	Phe	Asn	Phe	Leu 345	Met	Val	Leu	Gly	Lys 350		Ser
Phe	Gly	Lys 355	Val	Met	Leu	Ser	Glu 360	Arg	Lys	Gly	Thr	Asp 365		Leu	Tyr
Ala	Val 370		Ile	Leu	Lys	Lys 375		Val	Val	Ile	Gln 380	Asp	Asp	Asp	Val
Glu 385	Cys	Thr	Met	Val	Glu 390	Lys	Arg	Val	Leu	Ala 395	Leu	Pro	Gly	Lys	Pro 400
	Phe	Leu	Thr	Gln 405		His	Ser	Суs	Phe		Thr	Met	Asp	Arg 415	Leu
Tyr	Phe	Val	Met 420		Tyr	Val	Asn	Gly 425		Asp	Leu	Met	430		Ile
Gln	Gln	Val		Arg	Phe	Lys	Glu 440		His	Ala	Val	Ph∈		Ala	Ala
Glu	Ile 450		ılle	Gly	Leu	Phe 455		Leu	Gln	Ser	Lys 460		' Ile	e Ile	Tyr
Arg 465		Leu	Lys	Leu	470		val	Met	Leu	475		Glu	ı Gly	/ His	11e 480
		Ala	a Asp	Phe 485	e Gly		: Суя	Lys	Glu 490		Ile	Tr	Ası	Gl _y	/ Val
Thr	Thr	Lys	Thr	Phe		Gly	/ Thr	Pro	Asp		Ile	e Ala	9 Pro		ılle
Ile	Ala	тул 515		n Pro	туг	Gly	/ Lys		Va]	l Asp	Tr	525		a Phe	e Gly
Val	Let 530	ı Lev		Glu	ı Met	: Lev 535		a Gly	/ Glr	n Ala	Fro 540		e Gl	u Gly	y Glu
Asp 545	Glu		o Glu	ı Lev	Phe ب 550		n Sei	: Ile	e Met	: Glv 555		s Ası	n Va	l Al	a Tyr 560
		s Sei	r Met	Se: 56!	r Lys		ı Ala	a Val	L Ala	a Ile		s Ly	s Gl	y Le [.] 57	u Met
Thr	Lys	s His	5 Pro	Gl _y		s Arg	g Le	u Gly 585	y Cys		y Pro	o Gl	u G1 59		u Arg

Asp Ile Lys Glu His Ala Phe Phe Arg Tyr Ile Asp Trp Glu Lys Leu 600 Glu Arg Lys Glu Ile Gln Pro Pro Tyr Lys Pro Lys Ala Cys Gly Arg 615 620 Asn Ala Glu Asn Phe Asp Arg Phe Phe Thr Arg His Pro Pro Val Leu 635 630 Thr Pro Pro Asp Gln Glu Val Ile Arg Asn Ile Asp Gln Ser Glu Phe 645 650 Glu Gly Phe Ser Phe Val Asn Ser Glu Phe Leu Lys Pro Glu Val Lys 665 670 Ser Ser Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu Glu Leu 680 685 Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn 695 Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr 710 715 720 Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val 725 730 Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe 745 740 Ser Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala 760 Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp 775 Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu 790 795 Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn 805 810 Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr 825 Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe Lys Ile 840 Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln 855 Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His 870 875 Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg 885 890 Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu 905 Gly Met Asp Glu Leu Tyr Lys 915

(2) INFORMATION FOR SEQ ID NO:148:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3009 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:

(A) NAME/KEY: Coding Sequence

(B) LOCATION: 1...3006

(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:148:

									TTA Leu 10							48
									AAC Asn							96
									AAG Lys							144
									ATC Ile							192
									AGC Ser							240
									GAT Asp 90							288
GCG Ala	GGA Gly	CGG Arg	S AGT Ser 100	Pro	TTG	GAT Asp	CCC Pro	ATG Met 105	Thr	AGC Ser	CCA Pro	. GGA Gly	TCC Ser 110	Gly	CTA Leu	336
			Ala					Ser					sei		CTG Leu	384
ТАТ Туг	CGA Arg	Ser	GAC Asp	AGC Ser	GAT Asp	TAT Tyr 135	Asp	CTC Lev	TCT Ser	CC;	A AAC Lys	Sei	T ATC	TCC Ser	CGG Arg	432
	ı Ser					: Asp					Ası				ACT Thr 160	480
					l Le					Th:					TTT n Phe	528
				r Ası					g Ala					g Se	A CCC r Pro	576

ATG Met	TGC Cys	AAC Asn 195	CAA Gln	CCA Pro	TCC Ser	Ile	AAC Asn 200	AAA Lys	GCC . Ala	ACC Thr	ATA Ile	ACA (Thr (GAG (GAG (GCC Ala	624
TAC Tyr	CAG Gln 210	AAA Lys	CTG Leu	GCC Ala	AGC Ser	GAG Glu 215	ACC Thr	CTG Leu	GAG Glu	GAG Glu	CTG Leu 220	GAC Asp	TGG Trp	TGT Cys	CTG Leu	672
GAC Asp 225	CAG Gln	CTA Leu	GAG Glu	ACC Thr	CTA Leu 230	CAG Gln	ACC Thr	AGG Arg	CAC His	TCC Ser 235	GTC Val	AGT Ser	GAG Glu	ATG Met	GCC Ala 240	720
TCC Ser	AAC Asn	AAG Lys	TTT Phe	AAA Lys 245	AGG Arg	ATG Met	CTT Leu	AAT Asn	CGG Arg 250	GAG Glu	CTC Leu	ACC Thr	CAT His	CTC Leu 255	TCT Ser	768
GAA Glu	ATG Met	AGT Ser	CGG Arg 260	TCT Ser	GGA Gly	TAA Asn	CAA Gln	GTG Val 265	TCA Ser	GAG Glu	TTT Phe	ATA Ile	TCA Ser 270	AAC Asn	ACA Thr	816
TTC Phe	TTA Leu	GAT Asp 275	Lys	CAA Gln	CAT His	GAA Glu	GTG Val 280	Glu	ATT	CCT	TCT	CCA Pro 285	ACT Thr	CAG Gln	AAG Lys	864
GAA Glu	AAG Lys 290	Glu	AAA Lys	AAG Lys	AAA Lys	AGA Arg 295	CCA Pro	ATG Met	TCT	CAG	ATC Ile 300	AGT Ser	GGA Gly	GTC Val	AAG Lys	912
AAA Lys 305	Leu	ATC Met	CAC His	: AGC Ser	Ser 310	Ser	Leu	ACT Thr	`AAT Asn	TCA Ser 315	Ser	r ATC	CCA Pro	AGG Arg	Phe 320	960
GGA Gly	A GTM / Val	r AA/ Lys	A ACT	GAA Glu 325	ı Glr	A GAA	GAT Asp	r GTC o Val	CTT Leu 330	Ala	AA(G GAA S Glu	CTA Leu	GAA Glu 335	A GAT	1008
Va:	l Ası	ı Ly:	340	o Gly	y Let	ı His	va:	1 Phe 349	e Arg	j Il∙	e Ala	a Glu	350	ser	GGT Gly	1056
As:	n Ar	g Pr	o Le	u Th	r Vai	l Ile	36	t His	s Thi	r Il	e Ph	e Gli 369	n Glu	ı Arç	G GAT g Asp	1104
Le	u Le 37	u Ly 0	s Th	r Ph	e Ly	37	e Pr 5	o Vai	l As _l	p Th	r Le 38	u Il	e Th:	r Ty	T CTT r Leu	1152
Me 38	t Th	r Le	eu Gl	u As	р Ні 39	s Ту 0	r Hi	s Al	a As	p Va 39	1 Al	а Ту	r Hi	s As	C AAT n Asn 400	1200
TA Il	C CA e Hi	T GC .s Al	T GC .a Al	A GA a As	T GT p Va	T GT l Va	C CA 1 Gl	G TC n Se	T AC	T CA r Hi	T GT s Vā	NG CT	A TT u Le	A TO u Se	T ACA Thr	1248

415

405 CCT GCT TTG GAG GCT GTG TTT ACA GAT TTG GAG ATT CTT GCA GCA ATT Pro Ala Leu Glu Ala Val Phe Thr Asp Leu Glu Ile Leu Ala Ala Ile 425 TTT GCC AGT GCA ATA CAT GAT GTA GAT CAT CCT GGT GTG TCC AAT CAA 1344 Phe Ala Ser Ala Ile His Asp Val Asp His Pro Gly Val Ser Asn Gln 435 TTT CTG ATC AAT ACA AAC TCT GAA CTT GCC TTG ATG TAC AAT GAT TCC 1392 Phe Leu Ile Asn Thr Asn Ser Glu Leu Ala Leu Met Tyr Asn Asp Ser 455 TCA GTC TTA GAG AAC CAT CAT TTG GCT GTG GGC TTT AAA TTG CTT CAG 1440 Ser Val Leu Glu Asn His His Leu Ala Val Gly Phe Lys Leu Leu Gln 475 470 GAA GAA AAC TGT GAC ATT TTC CAG AAT TTG ACC AAA AAA CAA AGA CAA 1488 Glu Glu Asn Cys Asp Ile Phe Gln Asn Leu Thr Lys Lys Gln Arg Gln 490 485 TCT TTA AGG AAA ATG GTC ATT GAC ATC GTA CTT GCA ACA GAT ATG TCA 1536 Ser Leu Arg Lys Met Val Ile Asp Ile Val Leu Ala Thr Asp Met Ser 505 500 AAA CAC ATG AAT CTA CTG GCT GAT TTG AAG ACT ATG GTT GAA ACT AAG 1584 Lys His Met Asn Leu Leu Ala Asp Leu Lys Thr Met Val Glu Thr Lys 520 515 AAA GTG ACA AGC TCT GGA GTT CTT CTT GAT AAT TAT TCC GAT AGG Lys Val Thr Ser Ser Gly Val Leu Leu Leu Asp Asn Tyr Ser Asp Arg 530 535 ATT CAG GTT CTT CAG AAT ATG GTG CAC TGT GCA GAT CTG AGC AAC CCA Ile Gln Val Leu Gln Asn Met Val His Cys Ala Asp Leu Ser Asn Pro 550 ACA AAG CCT CTC CAG CTG TAC CGC CAG TGG ACG GAC CGG ATA ATG GAG 1728 Thr Lys Pro Leu Gln Leu Tyr Arg Gln Trp Thr Asp Arg Ile Met Glu 570 565 GAG TTC TTC CGC CAA GGA GAC CGA GAG AGG GAA CGT GGC ATG GAG ATA 1776 Glu Phe Phe Arg Gln Gly Asp Arg Glu Arg Glu Arg Gly Met Glu Ile 580 AGC CCC ATG TGT GAC AAG CAC AAT GCT TCC GTG GAA AAA TCA CAG GTG 1824 Ser Pro Met Cys Asp Lys His Asn Ala Ser Val Glu Lys Ser Gln Val 600 595 GGC TTC ATA GAC TAT ATT GTT CAT CCC CTC TGG GAG ACA TGG GCA GAC Gly Fhe Ile Asp Tyr Ile Val His Pro Leu Trp Glu Thr Trp Ala Asp 615

CTC (Leu ' 625								1920
CGT (1968
GAT (2016
GAA (2064
GC A								2112
TGT Z Cys '								2160
GAA (2208
GTC A								2256
GTA (2304
GAG (2352
GAC (Asp \ 785								2400
GCC A								2448
CTG (2496
CAG '								2544

AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG CGC ACC ATC TTC TTC 2592 Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe 850 855 AAG GAC GAC GGC AAC TAC AAG ACC CGC GCC GAG GTG AAG TTC GAG GGC Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly 870 GAC ACC CTG GTG AAC CGC ATC GAG CTG AAG GGC ATC GAC TTC AAG GAG 2688 Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu 890 GAC GGC AAC ATC CTG GGG CAC AAG CTG GAG TAC AAC TAC AAC AGC CAC 2736 Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His AAC GTC TAT ATC ATG GCC GAC AAG CAG AAG AAC GGC ATC AAG GTG AAC 2784 Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn 920 915 TTC AAG ATC CGC CAC AAC ATC GAG GAC GGC AGC GTG CAG CTC GCC GAC 2832 Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp 940 930 935 CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC CCC GTG CTG CCC 2880 His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro 955 945 950 GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG AGC AAA GAC CCC AAC 2928 Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn 965 970 GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC GTG ACC GCC GCC GGG 2976 Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly 985 980 3009 ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TAA Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 995 1000

840

835

(2) INFORMATION FOR SEQ ID NO:149:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1002 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
 (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:149:

Met 1	Ala	Gln	Gln	Thr 5	Ser	Pro	Asp	Thr	Leu 10	Thr	Val	Pro	Glu	Val 15	Asp
Asn	Pro	His	Суs 20	Pro	Asn	Pro	Trp	Leu 25	Asn	Glu	Asp	Leu	Val 30	Lys	Ser
Leu	Arg	Glu 35	Asn	Leu	Leu	Gln	His 40	Glu	Lys	Ser	Lys	Thr 45	Ala	Arg	Lys
Ser	Val 50	Ser	Pro	Lys	Leu	Ser 55	Pro	Val	Ile	Ser	Pro 60	Arg	Asn	Ser	Pro
Arg 65	Leu	Leu	Arg	Arg	Met 70	Leu	Leu	Ser	Ser	Asn 75	Ile	Pro	Lys	Gln	Arg 80
Arg	Phe	Thr	Val	Ala 85	His	Thr	Cys	Phe	Asp 90	Val	Asp	Asn	Gly	Thr 95	Ser
			100			-		105				-	110	Gly	
Ile	Leu	Gln 115	Ala	Asn	Phe	Val	His 120	Ser	Gln	Arg	Arg	Glu 125	Ser	Phe	Leu
	130					135	_				140			Ser	
145					150					155				Val	160
				165					170					Asn 175	
			180					185					190	Ser	
		195					200					205		Glu	
	210					215					220	-	-	Cys	
225					230			_		235				Met	240
				245					250					Leu 255	
			260					265					270	Asn	
		275					280					285		Gln	•
	290					295					300			Val	
305	Leu	Met	His	Ser	310	Ser	Leu	Thr	Asn	Ser 315	Ser	Ile	Pro	Arg	Phe 320
Gly	Val	Lys	Thr	Glu 325	Gln	Glu	Asp	Val	Leu 330	Ala	Lys	Glu	Leu	Glu 335	Asp
Val	Asn	Lys	Trp 340	Gly	Leu	His	Val	Phe 345	Arg	Ile	Ala	Glu	Leu 350	Ser	Gly
Asn	Arg	Pro 355	Leu	Thr	Val	Ile	Met 360	His	Thr	Ile	Phe	Gln 365	Glu	Arg	qzA
Leu	Leu 370	Lys	Thr	Phe	Lys	Ile 375	Pro	Val	Asp	Thr	Leu 380	Ile	Thr	Tyr	Leu
Met 385	Thr	Leu	Glu	Asp	His 390	Tyr	His	Ala	Asp	Val 395	Ala	Tyr	His	Asn	Asn 400
Ile	His	Ala	Ala	Asp 405	Val	Val	Gln	Ser	Thr 410	His	Val	Leu	Leu	Ser 415	Thr

Pro	Ala	Leu	Glu 420	Ala	Val	Phe		Asp 425	Leu	Glu	Ile	Leu	Ala 430	Ala	Ile
Phe	Ala	Ser 435		Ile	His.		Val 440	Asp	His	Pro	Gly	Val 445	Ser	Asn	Gln
Phe	Leu 450	Ile	Asn	Thr	Asn	Ser 455	Glu	Leu	Ala	Leu	Met 460	Tyr	Asn	Asp	Ser
Ser	Val	Leu	Glu	Asn	His	His	Leu	Ala	Val	Gly	Phe	Lys	Leu	Leu	Gln
465					470					475					480
				485	Ile				490					495	
			500		Val			505					510		
Lys	His	Met 515	Asn	Leu	Leu	Ala	Asp 520	Leu	Lys	Thr	Met	Val 525	Glu	Thr	Lys
Lys	Val 530	Thr	Ser	Ser	Gly	Val 535	Leu	Leu	Leu	Asp	Asn 540	Tyr	Ser	Asp	Arg
Ile 545	Gln	Val	Leu	Gln	Asn 550	Met	Val	His	Cys	Ala 555	Asp	Leu	Ser	Asn	Pro 560
Thr	Lys	Pro	Leu	Gln 565	Leu	Tyr	Arg	Gln	Trp 570	Thr	Asp	Arg	Ile	Met 575	Glu
Glu	Phe	Phe	Arg 580		Gly	Asp	Arg	Glu 585	Arg	Glu	Arg	Gly	Met 590	Glu	Ile
Ser	Pro	Met 595		Asp	Lys	His	Asn 600	Ala	Ser	Val	Glu	Lys 605	Ser	Gln	Val
Gly	Phe	Ile		Tyr	Ile	Val 615	His	Pro	Leu	Trp	Glu 620	Thr	Trp	Ala	Asp
Leu			Pro	Asp	Ala		Asp	Ile	Leu	Asp	Thr	Leu	Glu	Asp	Asn
625					630					635					640
				645					650					655	
			660					665					670	H	Phe
		675)				680					685	·		Ser
	690)				695					700	ı			Leu
705	•				710					715	5				720
Glu	Glu	Glu	ı Ala	Val 725		Glu	Glu	Glu	Glu 730		Glr	Pro	o Gli	a Ala 735	Cys
Val	. Ile	e Asp	Asp 740	Arg		Pro	Asp	Thr 745		Gly	/ Ile	e Lev	3 Glr 750		Thr
Va]	Pro	755		a Arg	, Asp	Pro	Pro 760		Alā	a Thi	r Met	769		r Ly:	s Gly
Glu	າ Glu 770		ı Phe	e Thr	Gly	Val 775		Pro	Ile	e Lei	u Val 780		ı Lei	ı Ası	o Gly
Asp 785	o Val		Gly	/ His	Lys 790		Ser	· Val	Sei	c Gl ₃		ı Gly	y Gli	u Gl	qzA y 008
		с Ту:	r Gly	7 Lys 809	s Leu		Leu	Lys	Phe 810	e Il		s Th	r Th	r Gl; 81	y Lys 5
Le	ı Pro	o Vā	1 Fro 820	Tr		Thr	Leu	. Val	Th		r Le	u Th	r Ty 83	r Gl	y Val
Glı	n Cv:	s Ph			ı Tyr	Pro	Asp			t Ly	s Gl	n Hi			e Phe
		83					840					84			

Lys	Ser 850	Ala	Met	Pro	Glu	Gly 855	Tyr	Val	GIn	GIu	860	Thr	11e	Pne	Phe	
Lys		Asp	Gly	Asn		Lys	Thr	Arg	Ala		Val	Lys	Phe	Glu	Gly 880	
865	Thr	Len	V-1	λen	870 Ara	Tle	Glu	Leu	Lvs	875 Glv	Ile	Asp	Phe	Lys		
ASP	1111	Leu	Val	885	A. g	110	Old	Dea	890	017				895		
Asp	Gly	Asn	Ile 900	Leu	Gly	His	Lys	Leu 905	Glu	Tyr	Asn	Tyr	Asn 910	Ser	His	
		915					Lys 920					925				
	930					935	Glu				940					
His	Tyr	Gln	Gln	Asn		Pro	Ile	Gly	Asp		Pro	Val	Leu	Leu	Pro 960	
945		II.	m	Lou	950	ሞb r	Gln	Ser	Ala	955 Leu	Ser	Lvs	Asp	Pro		
ASP	ASI	nıs	туг	965	Sei	1111	GIII	Ser	970	Dea	Der	210		975		
Glu	Lys	Arg	Asp 980		Met	Val	Leu	Leu 985	Glu	Phe	Va1	Thr	Ala 990		Gly	
Ile	Thr	Leu 995		Met	Asp		Leu 1000	Тут	Lys							
								0.75	NO	150.						
		(2) IN	FORM	ATIC	N FO	R SE	Ŏ ID	NO:	150:						
	(i) S	EQUE	NCE	CHAR	ACTE	RIST	ICS:								
							se p	airs								
			TYF					_								
			STR				ingl r	e								
		(D)	101	ODOC	,	1	-									
	(ii)	MOLE	CULI	E TYP	E: c	DNA									
	(ix)	FEAT	TURE :												
		()	A) NA	AME/I	ŒY:	Codi	.ng S	Seque	ence							
							3198									
		(1	0) 07	THER	INFO	RMAT	: NOI									
		(xi)	SEOL	IENC!	E DES	SCRIE	401T	1: SI	EO II	ON C	: 150	:				
ΥTA	G GAG	GC.	A GAG	G GGG	AG(AG(GCC	CCC	G GC	C CG	G GC	G GG(AG	C GG	A GAG	48
	t Glu	ı Al	a Glu		y Se:	r Sei	c Ala	a Pro		a Ar	g Al	a GI	/ Se	r GI 15	y Glu	
1				5					10					13		
GG	C AG	G GA	C AG	C GC	C GG	C GG	G GCC	C ACC	G CT	C AA	A GC	c cc	C AA	G CA	T CTC	96
) Ly	s Hi	s Leu	
			20					25					30			
m~-	C 7 C	3 C2	C GM	G CA	G CA	C C A	C CAC	TA	a ca	G CT	c ca	G CA	G CC	C CA	G TTC	144
Tr	o Ar	g Hi	s Gl	u Gl	n Hi	s Hi	s Gli	n Ty	r Pr	o Le	u Ar	g Gl	n Pr	o Gl	n Phe	
	-	35					40					45				
_				m	a a :	m ~:	C C :	C C	c cc	C CC	.c. cc	ים רר	ھ در	י אַ ריר	C TCG	192
CG	C CT a Le	C CT	G CA u Hi	r CC s Pr	c ca o Hi	r CA s Hi	s Hi	s Le	u Pr	o Pr	o Pr	o Pr	o Pr	o Pr	o Ser	
Λı	9 De 50		- 111			55			_		60					

CCC Pro 65	CAG Gln	CCC Pro	CAG Gln	CCC Pro	CAG Gln 70	TGT Cys	CCG Pro	CTA Leu	CAG Gln	CCG Pro 75	CCG Pro	CCG Pro	CCG Pro	CCC Pro	CCC Pro 80	240
CTG Leu	CCG Pro	CCG Pro	CCC Pro	CCG Pro 85	CCG Pro	CCG Pro	CCC Pro	GGG Gly	GCT Ala 90	GCC Ala	CGC Arg	GGC Gly	CGC Arg	TAC Tyr 95	GCC Ala	288
TCG Ser	AGC Ser	GGG Gly	GCC Ala 100	ACC Thr	GGC Gly	CGC Arg	GTC Val	CGG Arg 105	CAT His	CGC Arg	GGC Gly	TAC Tyr	TCG Ser 110	GAC Asp	ACC Thr	336
GAG Glu	CGC Arg	TAC Tyr 115	CTG Leu	TAC Tyr	TGT Cys	CGC Arg	GCC Ala 120	ATG Met	GAC Asp	CGC Arg	ACC Thr	TCC Ser 125	TAC Tyr	GCG Ala	GTG Val	384
									AAA Lys			Met				432
TCC Ser 145	Ser	TIC	CAG Gln	GGA Gly	CTC Leu 150	Arg	CGT Arg	TTT	GAT Asp	GTG Val 155	Asp	AAT Asn	GGC Gly	ACA Thr	TCT Ser 160	480
GCG Ala	GGA Gly	CGC	AGT Ser	CCC Pro 165	Leu	GAT Asp	CCC Pro	ATG Met	ACC Thr 170	Ser	Pro	GGA Gly	TCC Ser	: GGC : Gly 175	CTA Leu	528
ATT Ile	CTC	CA/	A GCA n Ala 180	Asn	TTT Phe	GTC Val	CAC His	AGT Ser 185	Gln	CGA Arg	Yrā	G GAC	TCC Sei 190	r Phe	C CTG e Leu	576
TAT Tyt	r CGA	A TCC g Se: 19!	r Asp	AGC Ser	GAT Asp	PAT T	GAC Asp 200	Lei	TCT 1 Ser	CCA Pro	A AAC	TC: Ser 209	r Me	G TC	c CGG r Arg	624
		r Se					o Ile					p Le			G ACT 1 Thr	
CC. Pr 22	o Ph	T GC e Al	T CA	G GTY	230	u Ala	AGʻ a Se:	r Le	G CGA	A AC' Thi 23:	r Va	A CG 1 A r	A AA g As	C AA n As	C TTT in Phe 240	2
GC Al	T GC. a Al	A TT a Le	A AC u Th	T AA' r As: 24	n Le	G CA u Gl	A GA' n As	T CG p Ar	A GCZ g Ala 25	a Pr	T AG o Se	C AA	A AG	A TO	CA CCC er Pro	768
AT Me	G TG t Cy	C AA	C CA n Gl 26	n Pr	A TC o Se	C AT	C AA e As	C AA n Ly 26	s Al	C AC a Th	C AT	'A AC .e Th	A GA ir Gl 27	lu G	AG GCC lu Ala	2 816 a
TA T)	C CA	G AA L' n.	A CI	G GC	C AG	C GA	G AC u Th	C CI	rg GA eu Gl	G GA u Gl	G CI u Le	rG GA eu As	AC TO sp Ti	G TY CP C	GT CT ys Le	G 864 u

275 280 285 GAC CAG CTA GAG ACC CTA CAG ACC AGG CAC TCC GTC AGT GAG ATG GCC 912 Asp Gln Leu Glu Thr Leu Gln Thr Arg His Ser Val Ser Glu Met Ala 295 TCC AAC AAG TTT AAA AGG ATG CTT AAT CGG GAG CTC ACC CAT CTC TCT 960 Ser Asn Lys Phe Lys Arg Met Leu Asn Arg Glu Leu Thr His Leu Ser 315 310 GAA ATG AGT CGG TCT GGA AAT CAA GTG TCA GAG TTT ATA TCA AAC ACA 1008 Glu Met Ser Arg Ser Gly Asn Gln Val Ser Glu Phe Ile Ser Asn Thr 330 325 TTC TTA GAT AAG CAA CAT GAA GTG GAA ATT CCT TCT CCA ACT CAG AAG 1056 Phe Leu Asp Lys Gln His Glu Val Glu Ile Pro Ser Pro Thr Gln Lys 345 340 GAA AAG GAG AAA AAG AAA AGA CCA ATG TCT CAG ATC AGT GGA GTC AAG 1104 Glu Lys Glu Lys Lys Lys Arg Pro Met Ser Gln Ile Ser Gly Val Lys 360 355 AAA TTG ATG CAC AGC TCT AGT CTG ACT AAT TCA AGT ATC CCA AGG TTT 1152 Lys Leu Met His Ser Ser Ser Leu Thr Asn Ser Ser Ile Pro Arg Phe 375 CGA GTT AAA ACT GAA CAA GAA GAT GTC CTT GCC AAG GAA CTA GAA GAT 1200 Gly Val Lys Thr Glu Gln Glu Asp Val Leu Ala Lys Glu Leu Glu Asp 395 GTG AAC AAA TGG GGT CTT CAT GTT TTC AGA ATA GCA GAG TTG TCT GGT 1248 Val Asn Lys Trp Gly Leu Eis Val Phe Arg Ile Ala Glu Leu Ser Gly 410 405 AAC CGG CCC TTG ACT GTT ATC ATG CAC ACC ATT TTT CAG GAA CGG GAT 1296 Asn Arg Pro Leu Thr Val Ile Met His Thr Ile Phe Gln Glu Arg Asp 425 420 TTA TTA AAA ACA TTT AAA ATT CCA GTA GAT ACT TTA ATT ACA TAT CTT Leu Leu Lys Thr Phe Lys Ile Pro Val Asp Thr Leu Ile Thr Tyr Leu 440 435 ATG ACT CTC GAA GAC CAT TAC CAT GCT GAT GTG GCC TAT CAC AAC AAT Met Thr Leu Glu Asp His Tyr His Ala Asp Val Ala Tyr His Asn Asn 455 450 ATC CAT GCT GCA GAT GTT GTC CAG TCT ACT CAT GTG CTA TTA TCT ACA Ile His Ala Ala Asp Val Val Gln Ser Thr His Val Leu Leu Ser Thr 470 CCT GCT TTG GAG GCT GTG TTT ACA GAT TTG GAG ATT CTT GCA GCA ATT 1488 Pro Ala Leu Glu Ala Val Phe Thr Asp Leu Glu Ile Leu Ala Ala Ile

490

								CAT His							1536
								GCC Ala							1584
								GTG Val							1632
								TTG Leu							1680
								GTA Val 570							1728
								AAG Lys					Thr		1776
		Ser					Leu	CTT Leu							1824
	Val					Val					Leu			CCA Pro	1872
Lys					Tyr					Asp				GAG Glu 640	1920
				Gly					Glu					ATA lle	1968
			: Asr					a Ser					Glr	G GTG n Val	2016
		e Asp					s Pro					Tr		A GAC a Asp	2064
	l Hi					As;					r Le			C AAT p Asn	2112
														A CCT a Pro	2160

705					710					715					720	
								GG T Gly								2208
								GAG Glu 745								2256
								AGC Ser								2304
TGT Cys	ACT Thr 770	CAA Gln	GAC Asp	TCA Ser	GAG Glu	TCT Ser 775	ACT Thr	GAA Glu	ATT Ile	CCC Pro	CTT Leu 780	GAT Asp	GAA Glu	CAG Gln	GTT Val	2352
								GAG Glu								2400
GTC Val	ATA Ile	GAT Asp	GAT Asp	CGT Arg 805	TCT Ser	CCT Pro	GAC Asp	ACG Thr	ACG Thr 810	GGA Gly	ATT Ile	CTG Leu	CAG Gln	TCG Ser 815	ACG Thr	2448
				Arg										Lys	GGC	2496
GAG Glu	GAG Glu	CTG Leu 835	Phe	ACC Thr	GGG Gly	GTG Val	GTG Val 840	Pro	ATC Ile	CTG	GTC Val	GAG Glu 845	. Lev	GAC Asp	GGC Gly	2544
GAC Asp	GTA Val 850	Asr	GGC Gly	CAC His	AAG Lys	TTC Phe 855	Ser	GTG Val	TCC Ser	GGC	GAG Glu 860	Gly	GAC	GG(GAT / Asp	2592
	Thr					Thr					э Сув				AAG Y Lys 880	2640
CTC Let	CCC Pro	GT(G CCC	TGC Trp 885	Pro	ACC Thr	CTC	GTG Ual	Thr 890	Thu	CTC	ACC 1 Thi	TAC Ty:	c GG r Gl; 89	C GTG y Val 5	2688
CA(Glr	TGC	TTO	2 AG(e Sei 90(: Arg	ТАС Э Туг	CCC Pro	GAC Asi	CAC His 905	Met	AA(G CAC	G CAG	GA(S As; 91	p Ph	C TTC e Phe	2736
			a Me					r Val					r Il		C TTC e Phe	2784

AAG GAC GAC GGC AAC TAC AAG ACC CGC C Lys Asp Asp Gly Asn Tyr Lys Thr Arg 2 930 935													
GAC ACC CTG GTG AAC CGC ATC GAG CTG Asp Thr Leu Val Asn Arg Ile Glu Leu 1945													
GAC GGC AAC ATC CTG GGG CAC AAG CTG Asp Gly Asn Ile Leu Gly His Lys Leu 965													
AAC GTC TAT ATC ATG GCC GAC AAG CAG Asn Val Tyr Ile Met Ala Asp Lys Gln 980 985													
TTC AAG ATC CGC CAC AAC ATC GAG GAC Phe Lys Ile Arg His Asn Ile Glu Asp 995 1000													
CAC TAC CAG CAG AAC ACC CCC ATC GGC His Tyr Gln Gln Asn Thr Pro Ile Gly 1010 1015													
GAC AAC CAC TAC CTG AGC ACC CAG TCC Asp Asn His Tyr Leu Ser Thr Gln Ser 1025													
GAG AAG CGC GAT CAC ATG GTC CTG CTG Glu Lys Arg Asp His Met Val Leu Leu 1045													
ATC ACT CTC GGC ATG GAC GAG CTG TAC Ile Thr Leu Gly Met Asp Glu Leu Tyr 1060 1065													
(2) INFORMATION FOR SEQ ID NO:151:													
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 1066 amino acids(B) TYPE: amino acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear													
<pre>(ii) MOLECULE TYPE: protein (v) FRAGMENT TYPE: internal</pre>													
(xi) SEQUENCE DESCRIPTION: SE	EQ ID NO:151:												

Met Glu Ala Glu Gly Ser Ser Ala Pro Ala Arg Ala Gly Ser Gly Glu

Gly Ser Asp Ser Ala Gly Gly Ala Thr Leu Lys Ala Pro Lys His Leu

25

1 5 10

Trp A		His 35	Glu	Gln	His	His	Gln 40	Tyr	Pro	Leu	Arg	Gln 45	Pro	Gln	Phe
Arg I			His	Pro		His 55	His	Leu	Pro	Pro	Pro 60	Pro	Pro	Pro	Ser
Pro C	3ln	Pro	Gln	Pro	Gln	Cys	Pro	Leu	Gln	Pro	Pro	Pro	Pro	Pro	
65					70				_	75		_,		_	80
Leu I				85					90					95	
Ser S	Ser	Gly	Ala 100	Thr	Gly	Arg	Val	Arg 105	His	Arg	Gly	Tyr	Ser 110	Asp	Thr
Glu A	Arg	Tyr 115	Leu	Tyr	Cys	Arg	Ala 120	Met	Asp	Arg	Thr	Ser 125	Tyr	Ala	Val
Glu '	Thr 130	Gly	His	Arg	Pro	Gly 135	Leu	Lys	Lys	Ser	Arg 140	Met	Ser	Trp	Pro
Ser :	Ser	Phe	Gln	Gly	Leu	Arg	Arg	Phe	Asp	Val	Asp	Asn	Gly	Thr	Ser
145					150					155					160
Ala				165					170					175	
			180				His	185					190		
Tyr	Arg	Ser 195		Ser	Asp	Tyr	Asp 200	Leu	Ser	Pro	Lys	Ser 205	Met	Ser	Arg
	Ser 210	Ser	Ile	Ala	Ser	Asp 215	Ile	His	Gly	Asp	220		Ile	Val	Thr
		Ala	Gln	Val	Leu	Ala	Ser	Leu	Arg	Thr	Val	Arg	Asn	Asn	Phe
225					230					235)				240
				245					250)				255	
			260)				265					270)	Ala
		275					280					285	·		Leu
	290					295	,				300)			Ala
	Asn	Lys	Ph€	. Lys			Leu	Asn	Arg	g Glv 31		ı Thi	His	s Lev	Ser 320
305	110 6	Cox	- 7	, 661	310		Gln	Val	Set			2 Ile	e Sei	r Ası	n Thr
				325	5				330)				335	5
			340)				345	·				35	0	Lys
		355	5				360)				36	5		l Lys
	370)				375	5				386	0			g Phe
Gly	Va]	l Ly:	s Th	r Glu	ı Glr	ı Glı	ı Asp	Va:	Le	u Al	a Ly:	s Gl	u Le	u Gl	u Asp
385					390					39					400
				40	5				41	0				41	
			42	0				42	5				43	0	g Asp
Leu	Le	ц Ly 43		r Ph	e Ly:	s Il	e Pro		l As	p Th	r Le	u I1 44		r Ty	r Leu
Met	Th	r Le	u Gl	u As	p Hi	з Ту	r His	s Al	a As	p Va	l Al	а Ту	r Hi	s As	n Asn
	45					45					46				

	His	Ala	Ala	Asp	Val	Val	Gln	Ser	Thr	His	Val	Leu	Leu	Ser	Thr 480
465		.	C3	71-	470 Val	Dho	ጥኮሎ	y c.c.	Len		Tlo	T en	Ma	Δla	
				485					490					495	
Phe	Ala	Ser	Ala 500	Ile	His	Asp	Val	Asp 505	His	Pro	Gly	Val	Ser 510	Asn	Gln
Phe	Leu	Ile 515	Asn	Thr	Asn	Ser	Glu 520	Leu	Ala	Leu	Met	Tyr 525	Asn	Asp	Ser
Ser	Val 530	Leu	Glu	Asn	His	His 535	Leu	Ala	Val	Gly	Phe 540	Lys	Leu	Leu	Gln
Glu		Asn	Cvs	Asp	Ile		Gln	Asn	Leu	Thr	Lys	Lys	Gln	Arg	Gln
545	010		-1-		550					555	-				560
	Leu	Arg	Lys	Met 565	Val	Ile	Asp	Ile	Val 570	Leu	Ala	Thr	Asp	Met 575	Ser
Lys	His	Met	Asn 580		Leu	Ala	Asp	Leu 585	Lys	Thr	Met	Val	Glu 590	Thr	Lys
Lys	Val	Thr		Ser	Gly	Val	Leu 600	Leu	Leu	Asp	Asn	Tyr 605	Ser	Asp	Arg
Ile	Gln 610		Leu	Gln	Asn	Met 615	Val	His	Cys	Ala	Asp 620	Leu	Ser	Asn	Pro
Thr		Pro	Leu	Gln	Leu		Arg	Gln	Trp	Thr	Asp	Arg	Ile	Met	Glu
625					630					635					640
Glu	Phe	Phe	Arg	Gln 645	Gly	Asp	Arg	Glu	Arg 650		Arg	Gly	Met	Glu 655	
Ser	Pro	Met	Суѕ	Asp	Lys	His	Asn	Ala	Ser	Val	Glu	Lys	Ser	Gln	Val
			660					665					670		
		675					680					685			Asp
	690					695					700) Asn
Arg 705		Trp	туг	Gln	Ser 710		Ile	Pro	Glr	n Ser 715		Ser	Pro	Ala	720
		Pro	Glu	Glu 725	Gly		Gln	Gly	Glr	ı Thı		Lys	: Phe	Glr 735	n Phe
Glu	Leu	Thi		Glu		Asp	Gly	Glu 745	Ser		Thr	Glu	ı Lys 750	s Asp	Ser
Gly	/ Ser				ı Glu	Asp		Ser		s Sei	Asp	Ser 765	c Lys		Leu
<u> </u>	. mb.s	759		. 501	- Glu	Sar	760		, T]	o Pro	n Lei			ı Glr	n Val
	770					775	·				780)			
		ı Git	ı Ala	a Val	790		1 610	l Gil	1 611	u se. 79!		1 110	0 01	u Ai	a Cys 800
785		7.51	n Acr) Arc			. Asr	Thi	r Th			e Lei	u Gl:	n Se:	r Thr
va.	1 116	- ASI	o voř	809			, ,,,,,		81		,			81	
Va:	l Pro	Arq	g Ala 820	a Arg		Pro	Pro	Val 825		a Th	r Me	t Va	1 Se 83		s Gly
Glv	ı Glv	Lei 83	ı Phe		r Gly	/ Val	l Val	Pro		e Le	u Va	1 G1: 84		u As	p Gly
Asj	p Va:	l As:		y Hi:	s Lys	5 Phe 859	e Ser		l Se	r Gl	y Gl	u Gl		u Gl	y Asp
	a Th		r Gl	y Ly:		ı Thi		ı Ly:	s Ph	e Il 87	е Су		r Th	r Gl	y Lys 880
86		o 17-	1 n~	~ ~~	870 n Pro		rte	, 1/2	ገ ጥኮ			ս ԴԴ	r T∨	r Gl	y Val
re.	u PT	υ va	ı PI	88		1111 ر	r ne	, va	89		20	- • • • •	1	89	

GIII	Cys	Phe	Ser 900	Arg	Tyr	Pro	Asp	His	Met	Lys	Gln	His	Asp 910	Phe	Phe	
Lys	Ser	Ala 915	Met	Pro	Glu	Gly	Tyr 920		Gln	Glu	Arg	Thr 925		Phe	Phe	
Lvs	Asp	Asp	Glv	Asn	Tvr	Lvs		Ara	Ala	Glu	Val		Phe	Glu	Glv	
-	930	•				935		3			940	-,, 0		010	011	
Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile	Asp	Phe	Lys	Glu	
945					950					955					960	
qzA	Gly	Asn	Ile	Leu 965	Gly	His	Lys	Leu	Glu 970	Tyr	Asn	Tyr	Asn	Ser 975	His	
Asn	Val	Tyr	Ile 980	Met	Ala	Asp	Lys	Gln 985	Lys	Asn	Gly	Ile	Lys 990	Val	Asn	
Phe	Lys	Ile	Arg	His	Asn	Ile	Glu	Asp	Gly	Ser	Val	Gln	Leu	Ala	Asp	
		995					1000					1005				
		Gln	Gln	Asn			Ile	Gly	Asp			Val	Leu	Leu	Pro	
	1010 Asn	His	ጥረት	T.e.u		1015	CJn	S07	פות		1020	TAG	Acn.	Dro	700	
025	VSII	1115	ıyı		1030	1111	Gill	ser		1035	ser	rys	Asp		1040	
	Lys	Arg	Asp			Val	Leu	Leu			Val	Thr	Ala			
			1	1045				:	1050				:	1055	_	
Ile	Thr	Leu		Met	Asp	Glu	Leu	Tyr	Lys							
			1060					1065								
		(2)	TNT	ישמט.	TOITE	T EO	0 000	ת דר	NO.	150.						
		(2)	, 2112	Oran	11101	• • •	(DL	2 110	110	152.						
	(:	i) SI	EQUE	CE (CHARA	ACTE	RIST	ics:								
		(A)	LENC	GTH:	3024	1 bas	se pa	airs								
		(B)	my DE				- i d									
				E: nu												
		(C)	STRA	ANDEI	ONESS	S: si	ingle	e								
		(C)	STRA	ANDEI		S: si	ingle	e								
	(:	(C) (D)	STRA	ANDEI DLOG!	ONESS 7: li	S: si inear	ingle	÷								
		(C)	STRA TOPO	ANDEI OLOGY TULE	ONESS 7: li	S: si inear	ingle	è								
		(C) (D) ii) 1	STRA TOPO	ANDEI OLOGY TULE	ONESS 7: li	S: si inear	ingle	÷								
		(C) (D) ii) M ix) H	STRATOPO	ANDEI DLOGY CULE JRE:	ONESS Y: li TYPE	S: si inear E: cI	ingle		nce							
		(C) (D) ii) N ix) H (A) (B)	STRATOPO MOLEO FEATU NAM LOO	ANDEI DLOGY TULE JRE: ME/KI	ONESS Y: li TYPE EY: C	S: si inear E: cI Codir	ingle ONA ag Se 3021		nce							
		(C) (D) ii) N ix) H (A) (B)	STRATOPO MOLEO FEATU NAM LOO	ANDEI DLOGY TULE JRE: ME/KI	ONESS Y: li TYPE	S: si inear E: cI Codir	ingle ONA ag Se 3021		nce							
	(:	(C) (D) ii) N ix) N (A) (B) (D)	STRATOPO	ANDEI DLOGY TULE JRE: JE/KH CATIC HER J	ONESS TYPE TYPE EY: CON: I	S: si inear E: cI Codir	ingle	equer		NO:	152:					
	(:	(C) (D) ii) N ix) H (A) (B)	STRATOPO	ANDEI DLOGY TULE JRE: JE/KH CATIC HER J	ONESS TYPE TYPE EY: CON: I	S: si inear E: cI Codir	ingle	equer		NO::	152:					
DTA	(:	(C) (D) ii) N ix) N (A) (B) (D)	STRATOPO HOLEO FEATU NAN LOO OTH	ANDEI DLOGY TULE JRE: TE/KI CATIC HER J	ONESS Y: li TYPE EY: C ON: l INFOR	S: siinear E: cI Codir	ingle	equer : SEÇ	Q ID			GGG	GCC	TGG	GAA	48
	(; (s) AGC	(C) (D) ii) 1 (A) (B) (D)	STRATOPO MOLECTERATURE NAME LOCUMENT OTHER SEQUE TCA	ANDEI DLOGY TULE JRE: ME/KH CATIC HER I	DNESS Y: li TYPE EY: CON: I INFOE DESC	S: sinear Codir Codir CMATI	ingle ONA ag Se BO21 EON: TION:	equer : SEX ACG	Q ID CAG	ACA	TGT					48
	(; (s) AGC	(C) (D) ii) N ix) H (A) (B) (D) ci) S	STRATOPO MOLECTERATURE NAME LOCUMENT OTHER SEQUE TCA	ANDEI DLOGY TULE JRE: ME/KH CATIC HER I	DNESS Y: li TYPE EY: CON: I INFOE DESC	S: sinear Codir Codir CMATI	ingle ONA ag Se BO21 EON: TION:	equer : SEX ACG	Q ID CAG	ACA	TGT					48
Met 1	(; AGC Ser	(C) (D) (iii) M (A) (B) (D) (D) TGG	STRATOPO	ANDEI CULE JRE: JRE/KI LCATIC CATIC CATIC CCT Pro 5	TYPE TYPE TYPE TYPE TYPE TYPE TYPE TYPE	S: sinean inean inean inean codir inean inean codir co	ong Se 3021 CON:	equer SEÇ ACG Thr	CAG Gln 10	ACA Thr	TGT Cys	Gly	Ala	Trp 15	Glu	
Met 1 ATG	(; AGC Ser	(C) (D) (D) (A) (A) (A) (A) (B) (D) (C) (C) (C) (C) (C) (C) (C) (C) (C) (C	STRATOPO IOLEO IOL	ANDEIDLOGY TULE THE CATIO THE CATIO TO THE C	ONESS (: li TYPP TYPP (NFOF TCC Ser GGG	S: siinean inean i	ODNA Second Sec	equer SEC ACG Thr	CAG Gln 10	ACA Thr GGA	TGT Cys	Gly GTC	Ala ATC	Trp 15 CGA	Glu TGG	4 8
Met 1 ATG	(; AGC Ser	(C) (D) (iii) M (A) (B) (D) (D) TGG	STRIVE TOPO IOLEO IOL	ANDEIDLOGY TULE THE CATIO THE CATIO TO THE C	ONESS (: li TYPP TYPP (NFOF TCC Ser GGG	S: siinean inean i	ODNA Second Sec	equer SE(ACG Thr GGA Gly	CAG Gln 10	ACA Thr GGA	TGT Cys	Gly GTC	Ala ATC Ile	Trp 15 CGA	Glu TGG	
Met 1 ATG	(; AGC Ser	(C) (D) (D) (A) (A) (A) (A) (B) (D) (C) (C) (C) (C) (C) (C) (C) (C) (C) (C	STRATOPO IOLEO IOL	ANDEIDLOGY TULE THE CATIO THE CATIO TO THE C	ONESS (: li TYPP TYPP (NFOF TCC Ser GGG	S: siinean inean i	ODNA Second Sec	equer SEC ACG Thr	CAG Gln 10	ACA Thr GGA	TGT Cys	Gly GTC	Ala ATC	Trp 15 CGA	Glu TGG	
Met 1 ATG Met	(; AGC Ser AAA Lys	(C) (D) (D) (A) (A) (A) (A) (B) (D) (C) (C) (C) (C) (C) (C) (C) (C) (C) (C	TOPC IOLEC NAN LOC OTF TCA Ser CGC Arg 20	ANDEI CULE IRE: IE/KH CATIC CCT Pro 5 CTT Leu	ONESS (: li TYPE EY: (ON: li CNFOF DESC TCC Ser GGG GJy	S: sinear inear in	ng Se 3021 CON: ACA Thr	SEÇ ACG Thr GGA G1y 25	CAG Gln 10 TTT Phe	ACA Thr GGA Gly	TGT Cys AAT Asn	Gly GTC Val	ATC Ile 30	Trp 15 CGA Arg	Glu TGG Trp	
Met 1 ATG Met	(; AGC Ser AAA Lys	(C) (D) (ii) 1 (A) (B) (D) (C) TGG Trp	STRATOPOOLOGICAL TOPOOLOGICAL TOPOOLOGICA TOPOOLOGICAL TOPOOLOGICA TOPOOLOGICA TOPOOLOGICA TOPOOLOGICA TOPOOLOGICA TOPOOLO	ANDEI	ONESS (: li TYPE EY: (C ON: li INFOF DESC TCC Ser GGG Gly GGT	S: sinear inear inear Codir Codir Codir CTG Leu ACA Thr	ng Se 3021 CON: ACA Thr	SEQ ACG Thr GGA Gly 25	CAG Gln 10 TTT Phe	ACA Thr GGA Gly	TGT Cys AAT Asn	Gly GTC Val	Ala ATC Ile 30	Trp 15 CGA Arg	Glu TGG Trp CAG	96
Met 1 ATG Met	(; AGC Ser AAA Lys	(C) (D) (ii) I (A) (B) (D) (D) TGG Trp GAG GAU	STRATOPOOLOGICAL TOPOOLOGICAL TOPOOLOGICA TOPOOLOGICAL TOPOOLOGICA TOPOOLOGICA TOPOOLOGICA TOPOOLOGICA TOPOOLOGICA TOPOOLO	ANDEI	ONESS (: li TYPE EY: (C ON: li INFOF DESC TCC Ser GGG Gly GGT	S: sinear inear inear Codir Codir Codir CTG Leu ACA Thr	ng Se 3021 CON: ACA Thr	SEQ ACG Thr GGA Gly 25	CAG Gln 10 TTT Phe	ACA Thr GGA Gly	TGT Cys AAT Asn	Gly GTC Val	Ala ATC Ile 30	Trp 15 CGA Arg	Glu TGG Trp CAG	96
Met 1 ATG Met CAC His	(; AGC Ser AAA Lys AAT ASn	(C) (D) (D) (A) (A) (B) (D) (C) (C) (C) (C) (C) (C) (C) (C) (C) (C	STRATOPO IOLEO IOL	ANDEI	ONESS (: li TYPE TYPE TYPE TYPE TYPE TYPE GON: 1 TCC Ser GGG Gly GGT Gly	S: si inear inear Codir L3 MATI CTG Leu ACA Thr	ONA ag Se 3021 ON: TION: ACA Thr GGG Gly CAG GGIn 40	SEQ ACG Thr GGA Gly 25 ATT Ile	CAG Gln 10 TTT Phe	ACA Thr GGA Gly ATC	TGT Cys AAT Asn AAG Lys	Gly GTC Val CAG Gln 45	Ala ATC Ile 30 TGC Cys	Trp 15 CGA Arg CGG Arg	Glu TGG Trp CAG Gln	96 144
Met 1 ATG Met CAC His	(): AGC Ser AAAA Lys AAT Asn	(C) (D) (D) (A) (A) (B) (D) (C) (C) (C) (C) (C) (C) (C) (C) (C) (C	STRATOPO MOLEC FEATURE NAM LOC OTF TCA Ser CGC Arg 20 GAA Glu CCC	ANDEIDLOGY TULE TRE: TE/KE TE/KE TE/KE TE/KE TO T	ONESSON (: 1) TYPE Y: (ON: 1) (ONFOR TCC Ser GGG Gly GGT GAAC	S: sinear inear inear inear Codir Codir Codir CTG Leu ACA Thr GAG Glu CGA	ona Section Se	SEQ ACG Thr GGA Gly 25 ATT Ile	CAG Gln 10 TTT Phe GCC Ala	ACA Thr GGA Gly ATC Ile	TGT Cys AAT Asn AAG Lys	Gly GTC Val CAG Gln 45	Ala ATC Ile 30 TGC Cys	Trp 15 CGA Arg CGG Arg	Glu TGG Trp CAG Gln	96

Ser Ser Leu Pro Tyr Pro Asn Asn Leu Asn Ser Val Leu Ala Glu Arg 265

260

							ATG Met 280									864
							CCC Pro									912
							GTT Val									960
							ACA Thr									1008
							ACG Thr									1056
							GCG Ala 360									1104
							TTA Leu									1152
							AAC Asn									1200
							GAA Glu									1248
							TTC Phe									1296
Val	Trp	His 435	Ser	Ile	Gln	Thr	CTG Leu 440	Lys	Glu	Asp	Cys	Asn 445	Arg	Leu	Gln	1344
							ATG Met									1392
							ATG Met									1440
							ACC Thr									1488

TAC AGC GAG CAA ACC GAG TTT GGG ATC ACA TCA GAT AAA CTG CTG CTG Tyr Ser Glu Gln Thr Glu Phe Gly Ile Thr Ser Asp Lys Leu Leu GCC TGG AGG GAA ATG GAG CAG GCT GTG GAG CTC TGT GGG CGG GAG AAC Ala Trp Arg Glu Met Glu Gln Ala Val Glu Leu Cys Gly Arg Glu Asn GAA GTG AAA CTC CTG GTA GAA CGG ATG ATG GCT CTG CAG ACC GAC ATT Glu Val Lys Leu Leu Val Glu Arg Met Met Ala Leu Gln Thr Asp Ile GTG GAC TTA CAG AGG AGC CCC ATG GGC CGG AAG CAG GGG GGA ACG CTG Val Asp Leu Gln Arg Ser Pro Met Gly Arg Lys Gln Gly Gly Thr Leu GAC GAC CTA GAG GAG CAA GCA AGG GAG CTG TAC AGG AGA CTA AGG GAA Asp Asp Leu Glu Glu Gln Ala Arg Glu Leu Tyr Arg Arg Leu Arg Glu AAA CCT CGA GAC CAG CGA ACT GAG GGT GAC AGT CAG GAA ATG GTA CGG Lys Pro Arg Asp Gln Arg Thr Glu Gly Asp Ser Gln Glu Met Val Arg CTG CTG CTT CAG GCA ATT CAG AGC TTC GAG AAG AAA GTG CGA GTG ATC Leu Leu Gln Ala Ile Gln Ser Phe Glu Lys Lys Val Arg Val Ile TAT ACG CAG CTC AGT AAA ACT GTG GTT TGC AAG CAG AAG GCG CTG GAA Tyr Thr Gln Leu Ser Lys Thr Val Val Cys Lys Gln Lys Ala Leu Glu CTG TTG CCC AAG GTG GAA GAG GTG GTG AGC TTA ATG AAT GAG GAT GAG Leu Leu Pro Lys Val Glu Glu Val Val Ser Leu Met Asn Glu Asp Glu AAG ACT GTT GTC CGG CTG CAG GAG AAG CGG CAG AAG GAG CTC TGG AAT Lys Thr Val Val Arg Leu Gln Glu Lys Arg Gln Lys Glu Leu Trp Asn CTC CTG AAG ATT GCT TGT AGC AAG GTC CGT GGT CCT GTC AGT GGA AGC Leu Leu Lys Ile Ala Cys Ser Lys Val Arg Gly Pro Val Ser Gly Ser CCG GAT AGC ATG AAT GCC TCT CGA CTT AGC CAG CCT GGG CAG CTG ATG Pro Asp Ser Met Asn Ala Ser Arg Leu Ser Gln Pro Gly Gln Leu Met TOT CAG COO TOO ACG GOO TOO AAC AGO TTA COT GAG COA GOO AAG AAG Ser Gln Pro Ser Thr Ala Ser Asn Ser Leu Pro Glu Pro Ala Lys Lys

					CAT His				2160
					GAA Glu				2208
_				_	GAA Glu 745				2256
_					GCC Ala				2304
					TTC Phe				2352
_	_		_		GGC Gly				2400
					GGC Gly				2448
					CCC Pro 825				2496
					AGC Ser				2544
					ATG Met				2592
					GGC Gly				2640
					GTG Val				2688
_					ATC Ile 905				2736
					ATC Ile				2784

925

GGC ATC AAG GTG AAC TTC AAG ATC CGC CAC AAC ATC GAG GAC GGC AGC 2832 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 930 935 GTG CAG CTC GCC GAC CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 945 950 CCC GTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG 2928 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 970 965 AGC AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG GAG TTC 2976 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 985

920

GTG ACC GCC GCG GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TAA 3024 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 1000 995

(2) INFORMATION FOR SEQ ID NO:153:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1007 amino acids
- (B) TYPE: amino acid

980

915

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:153:

Met Ser Trp Ser Pro Ser Leu Thr Thr Gln Thr Cys Gly Ala Trp Glu 10 Met Lys Glu Arg Leu Gly Thr Gly Gly Phe Gly Asn Val Ile Arg Trp 25 His Asn Gln Glu Thr Gly Glu Gln Ile Ala Ile Lys Gln Cys Arg Gln 40 Glu Leu Ser Pro Arg Asn Arg Glu Arg Trp Cys Leu Glu Ile Gln Ile 55 Met Arg Arg Leu Thr His Pro Asn Val Val Ala Ala Arg Asp Val Pro 70 Glu Gly Met Gln Asn Leu Ala Pro Asn Asp Leu Pro Leu Leu Ala Met 90 Glu Tyr Cys Gln Gly Gly Asp Leu Arg Lys Tyr Leu Asn Gln Phe Glu 105 Asn Cys Cys Gly Leu Arg Glu Gly Ala Ile Leu Thr Leu Leu Ser Asp 120 125 115 Ile Ala Ser Ala Leu Arg Tyr Leu His Glu Asn Arg Ile Ile His Arg 135 140 130

Asp 1	Leu	Lys	Pro	Glu	Asn 150	Ile	Val :	Leu	Gln	Gln 155	Gly	Glu	Gln	Arg	Leu 160
Ile	His	Lys	Ile	Ile 165		Leu	Gly	Tyr	Ala 170	Lys	Glu	Leu	Asp	Gln 175	Gly
Ser :	Leu	Cys	Thr 180	Ser	Phe	Val		Thr 185	Leu	Gln	Tyr	Leu	Ala 190	Pro	Glu
Leu :	Leu	Glu 195	Gln	Gln	Lys	Tyr	Thr 200	Val	Thr	Val	Asp	Tyr 205	Trp	Ser	Phe
	210				Glu	215					220				
Asn	Trp	Gln	Pro	Val	Gln	Trp	His	Ser	Lys		Arg	Gln	Lys	Ser	
225					230					235	_,			5 1	240
				245	Ser				250					255	
			260		Pro			265					270		
		275			Gln		280					285			
-	290				Tyr	295					300				
305					Lys 310					315					320
				325					330					335	
_			340		Gln			345					350		
		355			Gly		360					365			
	370				Gly	375					380				
385					390					395	•				Gln 400
				405	,				410)				415	
			420)				425	i				430)	Gln
		435	5				440					445	5		ı Gln
	450)				455					460)			c Cys
465					470)				479	5				1 Lys 480
				485	5				49	0				49	
			500	Э				505	5				51	0	ı Leu
		51	5				520)				52	5		u Asn
	53	0				535	5				54	0			p Ile
Val	As	p Le	u Gl	n Ar			o Met	Gl ₂	y Ar			n Gl	y Gl	y Th	r Leu
545			- 2		550			- 63	• -	55 m		~ n~	a 10	11 A×	560 a Glu
Asp	As;	p Le	u Gl	u Gl 56		n Ala	a Arg	المانى تى	u Le 57		ı Ar	y MI	a re	u A1 57	g Glu 5

Lys F	Pro Z	Arg	Asp 580	Gln	Arg	Thr		Gly 585	Asp	Ser	Gln		Met 590	Val	Arg
Leu I		Leu 595		Ala	Ile	Gln			Glu	Lys	Lys	Val 605	Arg	Val	Ile
Tyr 7			Leu	Ser		Thr 615		Val	Cys	Lys	Gln 620	Lys	Ala	Leu	Glu
Leu I		Pro	Lys	Val			Val	Val	Ser	Leu	Met	Asn	Glu	qaA	Glu
625					630					635					640
Lys '	Thr	Val	Val	Arg 645	Leu	Glr.	Glu	Lys	Arg 650	Gln	Lys	Glu	Leu	77p	Asn
Leu 1	Leu	Lys	Ile 660	Ala	Cys	Ser	Lys	Val 665	Arg	Gly	Pro	Val	Ser 670	Gly	Ser
Pro .	Asp	Ser 675	Met	Asn	Ala	Ser	Arg 680	Leu	Ser	Gln	Pro	Gly 685	Gln	Leu	Met
Ser	Gln 690		Ser	Thr	Ala	Ser 695	Asn	Ser	Leu	Pro	Glu 700	Pro	Ala	Lys	Lys
		Glu	Leu	Val	Ala 710	Glu	Ala	His	Asn	Leu 715	Cys	Thr	Leu	Leu	Glu 720
	Ala	Ile	Gln	Asp 725	Thr	Val	Arg	Glu	Gln 730	Asp	Gln	Ser	Phe	Thr 735	Ala
Leu	Asp	Trp	Ser 740		Leu	Gln	Thr	Glu 745	Glu	Glu	Glu	His	Ser 750		Leu
Glu	Gln	Ala 755		Trp	Val	Pro	Arg 760		Arg	Asp	Pro	Pro		Ala	Thr
Met			Lys	Gly	Glu	Glu 775	Leu	Phe	Thr	Gly	Val		Pro	Ile	Leu
	770 Glu	Leu	Asp	Gly	Asp 790	Val		Gly	His	Lys 795	Phe		Val	Ser	Gly 800
785 Glu	Gly	Glu	Gly	Asp 805	Ala		Tyr	Gly	Lys 810	Leu		Leu	Lys	Phe 815	lle
Cys	Thr	Thr	Gly 820	Lys		Pro	Val	Pro	Trp		Thr	Leu	Val	Thr	Thr
Leu	Thr	Туг 835	Gly		Gln	Суз	Fhe	. Ser		туг	Pro	Asp 845	His		Lys
Gln		Asp		Phe	Lys	Ser 855	Ala		Pro	Glu	1 Gly 860	туг		l Glr	n Glu
	850 Thr		Phe	Phe		Asp		Gly	/ Asr	тут 875	Lys		r Arg	g Ala	a Glu 880
865	Two	Dhe	. Glu	. Glv	870 Asr		Let	ı Val	Asr			e Gli	ı Let	ı Ly:	s Gly
				885)				890)				89	5
			900)				905	5				91	0	u Tyr
		915	5				920)				925	5		s Asn
	930)				93	5				940	3			y Ser
Val	Gln	Let	Ala د	a Asp			r Glr	n Gli	n Asi			o Il	e Gl	y As	p Gly
945 Pro		Lei	ı T.e.	ı Pro	950 124 c		n Hi	s Tv	r Le	95 u Se		r Gl	n Se	r Al	960 a Leu
				96	5				97	0				97	5
			98	0				98	5				99	0	u Phe
Val	Thr	Al. 99		a Gl	y Ile	e Th	100		у ме	t AS	b er	u Le 100		r hÀ	3

(2) INFORMATION FOR SEQ ID NO:154:

(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 2793 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence

(B) LOCATION: 1...2790(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:154:

	_			TTT Phe					48
				GCG Ala 25					96
				ATT Ile				1	44
				тат Туг				1	.92
				AAC Asn				2	240
				CCA Pro				2	38
				GCT Ala 105				3	36
				ATG Met				3	884
				TAC Tyr				4	132

										GAG Glu					480
										TTT Phe					528
										CGG Arg					576
										AAG Lys					624
	Pro									AAA Lys 220					672
Ser										CAC His					720
														GAT Asp	768
			Glu					Asn					His	GTT Val	816
		Ala					/ Asr					Val		ATG Met	864
	: Ile					Asp					Ph€			CCA Pro	912
l Ası					туг					ي Glv				C CAT r His 320	960
				His					Al.					C CAG 1 Gln 5	1008
			Let					o Ala					1 Ph	T ACA e Thr	1056
														T GTA p Val	1104

355 360 365

GAT (1152
CTT Leu .																1200
											TGT Cys					1248
											AAA Lys					1296
											AAT Asn					1344
											AGC Ser 460					1392
											CTT Leu					1440
					Ser										CGC Arg	1488
				Arg					Phe					/ Asp	CGA Arg	1536
			Arg					Ser					Lys		AAT Asn	1584
		. Val					val					тут			CAT His	1632
	Let					Ala					s Pro				G GAT n Asp 560	1680
					ı Glı					ı Tr					A ATC r Ile 5	1728

								GAC Asp							1776
								CTA Leu							1824
								AGT Ser							1972
								ACT Thr							1920
								GAG Glu 650							1968
								ATA Ile					Pro		2016
		Ile										Asp		CCG Pro	2064
	Thr					Gly					Thr			GTG Val	2112
ıle					Asp					Gly				AGC Ser 720	2160
				, Glu					Тут					C CTG	2208
			Thr					Pro					o Thi	C CTC r Leu	2256
		Let					Glr					y Ty:		GAC O Asp	2304
	Lys					e Phe					Pr			C TAC y Tyr	2352
														G ACC s Thr	2400

785					790					795					800	
CGC (2448
CTG .																2496
CTG Leu																2544
CAG Gln																2592
									TAC Tyr							2640
					Leu					His					CAG Gln	2688
TCC Ser	GCC Ala	CTG Leu	AGC Ser 900	Lys	GAC Asp	CCC Pro	AAC Asn	GAG Glu 905	Lys	CGC Arg	GAT Asp	CAC	Met	. Val	CTG Leu	2736
			Val					/ Ile					Asp		G CTG	2784
	AAG Lys 930															2793

- (2) INFORMATION FOR SEQ ID NO:155:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 930 amino acids
 - (E) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:155:

Met Met His Val Asn Asn Phe Pro Phe Arg Arg His Ser Trp Ile Cys
1 5 10 15

Phe	Asp	Val	Asp 20	Asn	Gly	Thr	Ser	Ala 25	Gly	Arg	Ser		Leu 30	Asp	Pro
		35	Pro		Ser		40					45			
	50					55					60				
Leu 65	Ser	Pro	Lys	Ser	Met 70	Ser	Arg	Asn	Ser	Ser 75	Ile	Ala	Ser	Asp	Ile 80
His	Gly	Asp	Asp	Leu 85	Ile	Val	Thr	Pro	Phe 90	Ala	Gln	Val	Leu	Ala 95	Ser
Leu	Arg	Thr	Val	Arg	Asn	Asn	Phe	Ala 105	Ala	Leu	Thr	Asn	Leu 110	Gln	Asp
Arg	Ala	Pro		Lys	Arg	Ser	Pro	Met	Суѕ	Asn	Gln	Pro 125	Ser	Ile	Asn
Lys	Ala 130		Ile	Thr	Glu	Glu 135	Ala	Tyr	Gln	Lys	Leu 140	Ala	Ser	Glu	Thr
Leu 145	Glu	Glu	Leu	Asp	Trp 150		Leu	Asp	Gln	Leu 155	Glu	Thr	Leu	Gln	Thr 160
		Ser	Val	Ser 165	Glu	Met	Ala	Ser	Asn 170		Phe	Lys	Arg	Met 175	Leu
Asn	Arg	Glu	Leu 180		His	Leu	Ser	Glu 185	Met	Ser	Arg	Ser	Gly 190		Gln
Val	Ser	Glu 195	Phe		Ser	Asn	Thr 200		Leu	Asp	Lys	Gln 205	His	Glu	Val
Glu	11e	Pro		Pro	Thr	Gln 215	Lys	Glu	Lys	Glu	Lys 220		Lys	Arg	Pro
Met			ılle	Ser		Val	Lys	Lys	Leu			Ser	Ser	Ser	Leu 240
225	her	Ser	· Ser	· Tle	230 Pro	Ara	Phe	Glv	. Val	235 Lvs		Glu	Gln	Glu	Asp
				245					250)				255)
			260)				265					270)	: Val
		275	5				280)				285	•		Met -
	290)				295					300)			Pro
		o Thi	r Lev	ı Ile	Thr 310		Let	ı Met	Thi	r Leu 319		ı Asp	His	з Туз	His 320
305 Ala	o a Asp	o Vai	l Ala	а Туг			Asr	ı Ile	Hi:			a Asp	o Val	l Vai	l Gln
				325	5				33	0				33:	5
Sei	c Th	r Hi	s Va. 340		ı Lev	Ser	Thi	r Pro 345		a Le	1 G11	ı Ala	a va. 350		e Thr
Asj	p Le	u Gl: 35	u Ile		ı Ala	a Ala	11e	e Phe		a Se	r Al	a Ile 36		s As	p Val
Asj	р Ні 37	s Pr		y Vai	l Ser	Asr 375	ı Glı		e Le	u Il	e As: 38		r Ası	n Se	r Glu
Le			u Me	t Ty:			Se:	r Se:	r Va			u As	n Hi	s Hi	s Leu
38 גיג		ງເາ	v Ph	e Tar	390 5 Leu		ı Gl:	n Gli	u Gl	39 u As		s As	p Il	e Ph	400 e Gln
				40	5				41	0				41	5
			42	0				42	5				43	0	e Asp
Il	e Va	1 Le 43		a Th	r Asj	o Me	t Se 44		s Hi	s Me	t As	n Le		u Al	a Asp

Leu	Lys 450	Thr	Met	Val	Glu	Thr 455	Lys	Lys	Val	Thr	Ser 460	Ser	Gly	Val	Leu
Leu 465	Leu	Asp	Asn	Tyr	Ser 470	Asp	Arg	lle	Gln	Val 475	Leu	Gln	Asn	Met	Val 480
	Cys	Ala	Asp	Leu 485	Ser	Asn	Pro	Thr	Lys 490	Pro	Leu	Gln	Leu	Tyr 495	Arg
Gln	Trp	Thr	Asp		Ile	Met	Glu	Glu 505	Phe	Phe	Arg	Gln	Gly 510	Asp	Arg
Glu	Arg	Glu 515		Gly	Met	Glu	Ile 520		Pro	Met	Cys	Asp 525	Lys	His	Asn
Ala	Ser 530		Glu	Lys	Ser	Gln 535		Gly	Phe	Ile	Asp 540	Tyr	Ile	Val	His
Pro		Trp	Glu	Thr	Trp 550		Asp	Leu	Val	His 555	Pro	Asp	Ala	Gln	Asp 560
	Leu	Asp	Thr	Leu 565	Glu	Asp	Asn	Arg	Glu 570		Tyr	Gln	Ser	Thr 575	
Pro	Gln	Ser	Pro 580		Pro	Ala	Pro	Asp 585		Pro	Glu	Glu	Gly 590		Gln
Gly	Gln	Thr 595		Lys	Phe	Gln	Phe		Leu	Thr	Leu	Glu 605		Asp	Gly
Glu	Ser 610		Thr	Glu	Lys	Asp 615		Gly	Ser	Gln	Val 620	Glu	Glu	Asp	Thr
Ser 625		Ser	Asp	Ser	Lys 630	Thr	Leu	Cys	Thr	Gln 635	Asp	Ser	Glu	Ser	Thr 640
	Ile	Pro	Leu	Asp 645	Glu	Gln	Val	Glu	Glu 650	Glu	Ala	Val	Gly	Glu 655	
Glu	Glu	Ser	Gln 660		Glu	Ala	Cys	Val 665		Asp	Asp	Arg	Ser 670		qzA
Thr	Thr	Gly 675		Leu	Gln	Ser	Thr 680		Pro	Arg	Ala	Arg 685		Pro	Pro
Val	Ala 690		Met	Val	Ser	Lys 695		Glu	Glu	Leu	Phe		Gly	Val	Val
Pro 705		Leu	Val	Glu	Leu 710	Asp	Gly	Asp	Val	Asn 715		His	Lys	Phe	Ser 720
		Gly	Glu	Gly 725		Gly	Asp	Ala	Thr 730		Gly	Lys	Leu	735	Leu
Lys	Phe	Ile	Cys 740		Thr	Gly	Lys	Leu 745		Val	Pro	Trp	750		Leu
Val	Thr	Thr 755		Thr	Tyr	Gly	Val 760		. Cys	Fhe	e Ser	765		Pro	qzA o
His	Met 770		Gln	His	Asp	Phe 775		Lys	Ser	Ala	Met 780		Glu	ı Gly	7yr
Val 785		Glu	Arg	Thr	790		Phe	Lys	Asp	Asp 795		/ Asr	туг	Lys	800
Arg	Ala	Glu	Val	Lys 805		Glu	Gly	Asp	Thr 810		ı Val	Asr	n Arg	g Ile 81	e Glu 5
Leu	Lys	Gly	, Ile 820) Phe	Lys	Glu	Asp 825		/ Asr	ı Ile	e Lei	u Gly 830		s Lys
Leu	ı Glu	туя 835		Tyr	Asn	Ser	His 840		ı Val	Туз	r Ile	e Met 849		a As	o Lys
Glr	1 Lys 850		Gly	/ I16	e Lys	Val 855		n Ph€	e Lys	s Ile	e Arg 860		s Ası	n Il	e Glu
Asp 865		/ Sei	r Val	Glr	870		a Asp	His	s ТУ1	61i 87:		n Asi	n Th	r Pr	o Ile 880

Gly Asp Gly Pro Val Leu Le 885	u Pro Asp	Asn His Tyn 890	Leu Ser	Thr Gln 895	
Ser Ala Leu Ser Lys Asp Pr	o Asn Glu 905	Lys Arg Ası	His Met	Val Leu	
Leu Glu Phe Val Thr Ala Al	a Gly Ile	Thr Leu Gl	Met Asp	Glu Leu	
Tyr Lys	J 2 0				
930					
(2) INFORMATION F	OR SEQ ID	NO:156:			
(i) SEQUENCE CHARACT (A) LENGTH: 37 bas					
(B) TYPE: nucleic					
(C) STRANDEDNESS:	single				
(D) TOPOLOGY: line	ar:				
(xi) SEQUENCE DESCR	[PTION: SE	Q ID NO:156	:		
GTAAGCTTCG AACATGATGC ACG					37
(2) INFORMATION	FOR SEQ ID) NO:15/:			
(i) SEQUENCE CHARAC	reristics:				
(A) LENGTH: 34 ba					
<pre>(B) TYPE: nucleic (C) STRANDEDNESS:</pre>					
(C) STRANDEDNESS: (D) TOPOLOGY: lin					
(2)					
(xi) SEQUENCE DESCR	IPTION: SE	EQ ID NO:157	7:		
GTAAGCTTCG AACATGGAGG CAG	agggcag c <i>i</i>	AGC			34
(2) INFORMATION	FOR SEQ II	NO:158:			
(i) SEQUENCE CHARAC	TERISTICS	:			
(A) LENGTH: 34 ba					
(B) TYPE: nucleio	acid				
(C) STRANDEDNESS:	-				
(D) TOPOLOGY: lin	ear				
(xi) SEQUENCE DESCR	IPTION: S	EQ ID NO:15	8:		
GTAAGCTTCG AACATGGCTC AGC	AGACAAG C	CCG			34
(2) INFORMATION	FOR SEQ I	D NO:159:			
(i) SEQUENCE CHARAC	TERISTICS	:			
(A) LENGTH: 37 ba					
(B) TYPE: nucleio					
(C) STRANDEDNESS	single				

(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:159:	
GTGAATTCCC GTCGTGTCAG GAGAAGCATC ATCTATG	37
(2) INFORMATION FOR SEQ ID NO:160:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 24 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:160:	
GTGAATTCAA CCATGGAGCG GGCC	24
(2) INFORMATION FOR SEQ ID NO:161:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 23 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:161:	
GTGGTACCCA GTTCCGCTTG GCC	23
(2) INFORMATION FOR SEQ ID NO:162:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 24 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:162:	
GTCTCGAGGC AAGATGGCTG ACCC	24
(2) INFORMATION FOR SEQ ID NO:163:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 25 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:163:	
GTGGATCCGA GCTCTTGACT TCGGG	25
(2) INFORMATION FOR SEQ ID NO:164:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 31 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:164:	
GTAAGCTTAC ATGAGCTGGT CACCTTCCCT G	31
(2) INFORMATION FOR SEQ ID NO:165:	
(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 25 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:165:	
GTGGTACCCA TGAGGCCTGC TCCAG	25

METHOD AND APPARATUS FOR HIGH DENSITY FORMAT SCREENING FOR BIOACTIVE MOLECULES

FIELD OF THE INVENTION

The invention relates to a method and apparatus for screening large numbers of molecules for biological activities.

BACKGROUND OF THE INVENTION

Current technology is able to generate large numbers of molecules which may possess potential therapeutic value. Compounds having potentially interesting biological activity may be products of combinatorial or traditional chemistry, a natural product, proteins isolated by one- or two-dimensional gel electrophoresis, or compounds secreted from or expressed by natural or genetically modified animal, plant, microbial or fungal cells (or parts thereof), or displayed by natural or genetically modified viral or phage particles.

Screening methods have been developed which achieve very high throughputs of test compounds. Such methods are termed Ultra High Throughput Screening (UHTS). The present generation of UHTS machines rely upon essentially serial additions of test compounds, usually one test compound per discrete test well. Test well array densities range from between 96 to 3456 wells per plate. Such UHTS machines require sophisticated technologies to dispense microvolumes of many different fluids to selected locations, and also require that the detecting surface for each test molecule generally be separated from other detecting surfaces within the array.

There is a need to develop a method which allows simultaneous screening of large numbers of test compounds for biological activity and potential therapeutic use while avoiding the complications associated with dispensing multiple fluid microvolumes.

BRIEF SUMMARY OF THE INVENTION

The invention is directed to a screening method which eliminates the need for delivering microfluid volumes and allows simultaneous parallel screening of large numbers of test compounds. Accordingly, the invention is drawn to a method for screening test

compounds for bioactivity, by contacting an array of test compounds with a detector layer capable of detecting bioactivity, wherein a cell response is indicative of bioactivity.

The method of the invention is a high throughput system for parallel screening of a large number of test compounds. In one embodiment of the method of the invention, 96 to 10,000 test compounds are simultaneously screened for bioactivity in an assay; in a more specific embodiment, 96 to 3456 test compounds are simultaneously screened for bioactivity.

In a more specific embodiment, invention is drawn to a method for screening test compounds for bioactivity, comprising:

- (a) contacting a solid support comprising an array of test compounds with a liquid layer, wherein the liquid layer is in immediate contact with a detector layer and wherein each test compound comes into contact with a localized portion of the liquid layer; and
- (b) registering a response of the detector layer to the test compound, wherein a bioactive test compound is identified.

By "high throughput screening" is meant a method able to screen large number of test compounds for biological activity within a given machine time (i.e. at a rate anywhere from 100 to 100,000 compounds per hour per machine).

The term "parallel screening" refers to a method by which very many compounds are applied simultaneously to the detector layer, and similarly, signals from that detector layer are collected contemporaneously rather than sequentially.

By "array" is meant a regular two-dimensional arrangement of test compounds by which compounds are disposed at the nodes of a rectilinear grid pattern whereby a compound position can be identified by a simple 2-dimensional coordinate.

A "detector layer" means any two-dimensional system which can be used to report biologically relevant information. In one specific embodiment of the method of the invention the detector layer is a monolayer of living cells loaded with a fluorescent reporter dye such as Fluo-3.

By "bioactive" or "bioactivity" is meant an action or influence of a test compound upon the detector layer which results in a response from the detector layer that has direct biological significance or can be interpreted as being a biologically relevant response.

Bioactive agents have the ability to effect physiological parameters of living cells and tissues. Bioactivity includes inducing or suppressing the expression of a protein, activating or inhibiting transcription of a gene, and/or effecting cellular function(s) such as, for example,

intracellular movement and storage of calcium ions, and membrane transportation.

The capacity of a test compound to affect a detector layer, i.e. bioactivity, may be determined in a number of ways known to the art. In specific embodiments of the method of the invention, bioactivity is determined by changes or movements of fluorescent probes present in the detector layer which indicate changes in ionic content, cell metabolism, growth or viability. In a preferred method of the invention, living cells form the detector layer and have specific protein components tagged with a fluorescent agent, such as green fluorescent protein (GFP); changes in GFP fluorescence or distribution within cells indicate a particular cellular response which may be selected for identification of bioactivity.

The phrase "a change in fluorescence" means any change in absorption properties, such as wavelength and intensity, or any change in spectral properties of the emitted light, such as a change of wavelength, fluorescence lifetime, intensity or polarization.

A "solid support comprising an array of multiple test compounds" or similar terms, mean a fixed matrix to which test compounds have been fixed. As an example, the solid support of the invention includes a membrane or other surface comprising an array of printed test compounds. In one specific embodiment of the invention, the test compounds are deposited as discrete spots on a porous track-etched polycarbonate membrane 10 to 20 microns thickness, the spots being between 10 microns to 2 mm diameter. The quantity of compound contained in each discrete spot will depend on the concentration of the stock solution from which it was derived, and the volume of that stock solution applied to the support. In another specific embodiment of the invention, compounds are printed onto a non-porous solid support which is optically clear.

By "test compounds" is meant a fixed array of compounds to be screened for ability to effect physiological parameters of a cell or tissue. In one embodiment, the test compounds are proteins or peptides generated by combinatorial protein chemical methods known to the art. In another embodiment, the test compounds are chemical compounds generated by combinatorial chemistry methods known in the art. In another embodiments, the test compounds are chemical compounds which are naturally occurring compounds more or less purified from their native state, are the products of genetically engineered cells, or are viral or bacteriophage particles engineered to display compounds upon their surfaces (phage display).

In one embodiment, the detector layer is an undemarcated area of living cells growing on a flat culture surface. The cells on this surface may or may not be grown to confluence,

may be transformed and/or engineered cells, or directly derived from animal tissues and grown as primary cell culture.

In one embodiment, a test compound reaches the detector layer by diffusion through a porous membrane to a liquid layer immediately overlaying the detector layer. A variety of commercially available porous membranes are useful in the method of the invention. A preferred porous membrane is a track-etched polyester or polycarbonate support in which parallel channels of identical size are formed by a selective etching process following exposure of the membrane to a source of high energy ions. The method of the invention allows each test compound affixed to a solid support to come into contact with a limited fluid volume, which fluid volume is in immediate contact with the detector layer. In one embodiment, each test compound contacts the detector layer by diffusion through a liquid-containing channel directly adjacent to the detector layer.

One advantage of the method of the invention is that it allows massive parallel screening of a large array of test compounds for biological activity. When living cells are the detector layer of the invention, they are maintained under physiologically viable conditions. Provision of these conditions requires the use of solutions able to supply essential nutrients and buffer pH changes normal to the continued growth of living cells. Such solutions may be complete cell culture media (i.e. any of those commercially available, for instance from Life Technologies Ltd.), optionally supplemented with antibiotics and serum preparations for optimal cell growth conditions. Buffer solutions may also be of the type known as "chemically defined". Cells will also require controlled temperature conditions, in the range 20° to 37°C, and the provision of gases essential to continued cell growth and maintenance of buffer capacity (O₂, and optionally 5% CO₂, depending on the type of buffer being used).

These and other objectives, advantages, and features of the invention will become apparent to those persons skilled in the art upon reading the details of the method as more fully described below.

BRIEF DESCRIPTION OF THE DRAWINGS

The foregoing features of the present invention may be fully understood from the following detailed disclosure of a specific preferred embodiment in conjunction with the accompanying drawings in which:

Fig. 1 is a schematic representation of the apparatus useful in one specific embodiment of the invention: Light from a high energy light source 1 is collected and collimated by unit 2, directed through a shutter assembly 3 and passes through a excitation filter-changer 4. A light guide 5 directs excitation light into the lensing and epi-illumination optics housed in unit 7. Excitation light emerging from 7 illuminates the horizontal detector layer located in the multi-component assembly having two solid layers 10 and 11 fixed relative to a supporting stage unit 8. Layer 11 is moved vertically downward on guide pins (17 Fig. 2b) controlled by arm 12 driven by unit 13. Four sprung contacts 14 attached to 12 press upon the frame of layer 11 to drive it downwards as arm 12 descends. Specified devices (3, 4, 9, 13, 15, 16) are controlled by central processing unit 6 which issues commands and collects data and status information from the devices attached to it. Unit 6 includes a central processing unit, RAM, multi-channel serial input/output cards with onboard A/D and D/A converters, one of which cards controls the camera 16 and captures images from it.

Figs. 2a-c: Figs. 2a and 2b are side view of the test stage (not to scale); Fig. 2c is a top view of the test stage. A supporting stage 8 has a rectangular central aperture the shape and size of which is the same as the area 19 of Fig. 2c. The position of stage 8 is adjusted in the horizontal and vertical axes by the 3-axis positioner 9. Components of the test stage shown include, solution layer 18, (not shown: detector layer 20 and array of test compounds 21 in Figs 3 and 4). The array 21 is held away from the liquid layer by pins 17 which pass through holes (24 in Fig. 5) in the corners of the frame 11. Arm 12 is moved down by the drive unit 13, and the four sprung contacts 14 it bears exert pressure on the frame 11 moving it down the guide pins 17 and into close proximity to the upper surface of 10, from which it is separated by a thin liquid layer 18.

Fig. 3 is a schematic showing the relative positions of the different layers in the test-array/detector layers used in one specific embodiment. The layers are depicted in apposition, as they would appear after arm 12 has pushed component 11 down the support pins 17. An array of discrete spots of test compounds 21 on a porous membrane 19 is in contact with a liquid layer 18 overlaying the detector layer 20 which is supported by an optically transparent

solid substrate 10. The compounds fill the parallel capillary spaces in the track-etched membrane 22.

Fig. 4 is a schematic drawing of a second embodiment of the screening method of the invention. The layers are depicted in apposition, as they would appear after arm 12 has pushed component 11 down the support pins 17. A detector layer 20 supported on an optically clear porous membrane 19, and overlayed by a liquid layer 23, is placed onto an optically clear solid substrate 10 bearing an array of test compounds 21. The thin space 18 between components 19 and 10 is filled with solution from 23 which has passed through the porous membrane 19. Bioactivity is detected by measuring changes in fluoresence of the detector layer resulting from responses to the diffusion of test compounds through the porous membrane to the detector layer.

Figs. 5a-c are schematics illustrating transfer printing of an array of compounds onto a surface of a track-etched membrane. Compounds are stored in 16 separate 96-well microtitre plates and defined amounts are transferred simultaneously by a 96-pin printing head to the surface 19 (Fig. 5a). The contents of each successive 96-well plate are printed at a slightly offset position, generating an array after 4 such printing operations (Fig. 5b), and a full array of 1536 compounds after 16 printing operations (Fig. 5c).

DETAILED DESCRIPTION

Before the present method and solutions used in the method are described, it is to be understood that this invention is not limited to particular methods, components, or solutions described, as such methods, components, and solutions may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting, since the scope of the present invention will be limited only by the appended claims.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods and

materials are now described. All publications mentioned herein are incorporated herein by reference to disclose and describe the methods and/or materials in connection with which the publications are cited.

Generally, the invention is drawn to a method for high throughput screening of test compounds, by contacting a solid support comprising an array of multiple test compounds with a detector layer, wherein each test compound comes into contact with a localized liquid which is in contact with a detector layer, and detecting a response of the detector layer to the test compound, wherein a bioactive test compound is identified.

The high density format screening system (HDFS) of the invention, rests in part on the realization that the delivery of test compounds to detector surfaces can be greatly simplified by doing away with the need for complicated microfluidics. Test compounds are applied to the detector surface in a massively parallel manner, and the method is applicable to a large range of different types of test compounds.

Central to the specific embodiments of the method and apparatus of the invention, described below, is the use of living cells as detectors, their responses being signalled via changes in the fluorescent or luminescent properties of various specific probes located within. However many different types of detector systems could be used in place of cells in such a system, for example, appropriate variants of Scintillation Proximity Assay (SPA) systems (Amersham Pharmacia Biotech) and enzyme-linked immuno-sorbent assay (ELISA) systems (Amersham).

Test Compound Arrays

The array of test compounds is formatted to have the same dimensions as the detector surface. In one specific embodiment of the invention, array and detector layers have a width of 8 cm and length of 12.5 cm, so as to fit within the format of conventional 96-well or 384-well microtiter plates. Preparation of the test arrays will depend on their origin.

Test compounds held in formatted arrays. Current methods for the production of single compounds by combinatorial methods are under development which involve miniaturization and patterned arrays of tethered solid-phase substrates. Thus, test compounds generated by combinatorial methods can be used to synthesize an array directly or indirectly on a carrier sheet. In one embodiment, vapor phase solubilization is used to produce a test compound array on the synthetic substrate, followed by a printing process of the test

compound array on to an absorbent membrane. In this embodiment, the test array is the printed membrane. An attractive feature of this method is that multiple copies of the same test array can be produced at one time to be screened against multiple cell systems for specific activities which minimizes stock handling from library archives.

Currently most compounds to be screened come in 96-well format. However, the 96-well format can be altered by repeated off-set printings, to any chosen density of format that the transfer substrate and assay can support. The optimum density of compounds in the test array will depend very much on the fraction of compounds in an array which generate bioactive responses in the detector layer ("hit rate"). The hit rate will depend on how well the compound library being tested matches the targets in the assay. If the hit rate is low, e.g., 1:20,000 - 100,000 compounds tested, a test array with center to center spacing of 200 µm (giving 240,000 separate compounds in a 12 cm x 8 cm area) may be preferable, providing 2 to 10 hits per plate. At a spacing of 1 mm, 9,600 test compounds may be screened simultaneously.

The density of the format may be adjusted as required without requiring any changes in the hardware used to perform the re-formatting; rather, adjustment may be made in the degree of off-set and the number of print operations used per array.

Detection

Fluorescent imaging provides a way to monitor physiological responses of living cells in a non-invasive manner. Ion- and voltage-sensitive probes, as well as the new generation of recombinant fluorescent probes, for instance, hybrid proteins comprising fusions of green fluorescent protein variants (GFPs) to cellular proteins involved in intracellular signaling, can be used singly or in combination to report on many aspects of cellular microphysiology. Due to the strong fluorescence of GFP, the luminescence of cells expressing the probes may easily be detected and analyzed by employing a combination of fluorescence microscopy and image analysis. Furthermore, these probes described are easily introduced into cells, as they can be expressed in the cells of interest after transfection with a suitable expression vector.

Recombinant probes for second messengers and enzyme activity, such as kinase activity, are not only useful in basic research but also in screening programs aiming at identifying novel biologically active substances. As an example, any currently used screening program designed to find compounds that affect cAMP concentration and protein kinase

activity are based on receptor binding and/or immuno detection and/or reporter gene expression. The recombinant probes described herein, on the other hand, make it possible to develop an entirely new types of screening assays able to monitor immediate and transient changes of cAMP concentration and protein kinase activity in intact living cells.

The HDFS method of the invention monitors the response of cell populations to test compounds. Lens systems are currently available which can simultaneously epi-illuminate and image the fluorescence from areas in excess of 8.5 x 13 cm, the size of a standard 96-well plate. The detection method used herein collects a variety of fluorescent signals from all cells in a field, with responses from discrete areas of the field being apparent in the real image of the fluorescence from that field as formed on the surface of the photosensitive detector (imaging camera).

Delivery of Test Compounds to Detector Cells

In a first embodiment of the method of the invention, delivery of large arrays of test compounds to cells is achieved with test compounds which are present on or transferred to a porous carrier sheet. In specific embodiments, test compounds are printed on the carrier sheet, and the sheet is applied (overlayed) to a field of cells of the same area. The test compounds reach the detector cells by diffusion through a localized buffer layer immediately in contact with an area of the detector cell layer. This embodiment is shown in the schematic of Figs. 2 & 3.

Porous carrier sheet for delivery of test compounds: Test compound arrays are fixed onto the porous carrier sheet by a variety of methods known to the art. For example, an array of test compounds may be transferred and fixed to the carrier sheet by the method of contact printing, whereby an array of inert flat-ended pins (e.g. made of stainless steel) is used to transfer defined volumes of individual test compounds (in the range 50 nl to 2 μ l) in solution form to discrete points on a dry carrier sheet.

A porous membrane useful in the delivery of test compounds is a membrane constructed of a non-absorbent material with pores of regular and defined diameter which traverse the membrane directly from the upper to the lower side. The property of orthogonal capillarity is useful in these membranes to limit lateral spread of test compounds applied to the membranes as discrete spots of liquid, since it is important that the compounds remain as discrete spots upon the membrane. A variety of membranes of different thicknesses,

materials, and pore densities are commercially available from a number of manufacturers. For example, porous membranes useful in the method of the invention include a track-etched polycarbonate or polyester membrane (Corning Costar or Whatman/Polyfiltronics). These are available in thicknesses from 6 to 23 microns, with pores of 14 to 0.015 microns, at 100,000 to 1,000,000,000 pores/cm². For delivery of test compounds with maximum ease of handling and loading of test compounds, polycarbonate membranes are preferred, particularly of a thickness of greater than 10 microns, with pores between 1 and 10 microns diameter at densities of between 20,000,000 to 100,000 pores/cm², respectively. One preferred membrane is Nucleopore® from Corning Costar.

Alternative membranes useful for the delivery of compounds include cast cellulose acetate (Membra-fil®), PTFE membranes (e.g. Filinert™), and glass fiber filters, all available from Corning Costar. These thicker membranes encourage lateral spread of liquid samples applied to their surfaces, but are thicker and could thus be used to deliver larger amounts of compounds.

Track-etched and cast cellulosic membranes may also be given hydrophilic or hydrophobic surface treatments. It is useful to have membranes whose surfaces have defined wettability properties.

When the test compound is soluble, the compound will dissolve into the buffer upon contact with the buffer medium, and directly contact the detector layer immediately underlying the buffer layer. In this embodiment, the test compounds dissolve upon contact with the buffer medium, and fall vertically onto the detector layer as a result of having a higher density than the surrounding liquor. It is generally preferred that the thin buffer layer between the test compound membrane and detector layer not be stirred significantly by convection. At the detector layer, the vertical fall of a solution of test compound is expected to spread radially by displacement and diffusion. The radial extent of a measured response may thus be use as an indicator of the bio-potency of the compounds involved.

Test compounds of limited solubility, such as those expressed on the surface of a carrier system, for instance, a cell membrane, viral or phage particle, must be brought into very close proximity, including direct contact, with the detector layers.

Buffer and Detector layer. The detector layer may be a continuous or non-continuous layer of living cells. In a specific embodiment, the detector layer is a continuous cell monolayer corresponding in size to the test compound array. In more specific embodiments,

thin glass substrate, suitably tissue culture treated is preferred for fluorescent probes requiring excitation wavelengths below 400 nm.

Living cells are maintained under physiologically viable conditions, as defined by such parameters as oxygen consumption, membrane potential, mitochondrial potential and cytoplasmic ion balance. Provision of these conditions requires the use of solutions able to supply essential nutrients and buffer pH changes normal to the continued growth of living cells. Such solutions may be complete cell culture media (i.e. any of those commercially available, for instance from Life Technologies Ltd.) optionally supplemented with antibiotics and serum preparations for optimal cell growth conditions. Buffer solutions may also be of the type known as "chemically defined" (e.g. phosphate buffered saline solutions). Cells will also require controlled temperature conditions, in the range 20° to 37°C, and the provision of gases essential to continued cell growth and maintenance of buffer capacity (O₂, and optionally 5% CO₂, depending on the type of buffer being used).

Detection of bioactivity. Detection of bioactivity may be determined by a number of methods known in the art. In a preferred embodiment, detection of bioactivity is determined by cellular imaging of fluorescence. For example, imaging may be conducted of a cell layer on a clear glass substrate. A glass substrate having a surface pitted with a regular array of very shallow (approx 20 μm) depressions may be used for this purpose (Corning). This glass substrate is useful because it ensures a regular and defined spacing between the overlying test array and the cells beneath.

In one embodiment, the detector layer is an undemarcated area of living cells growing on a flat culture surface. The cells on this surface may or may not be grown to confluence, may be transformed and/or engineered cells, or directly derived from animal tissues and grown as primary cell culture. In a second embodiment of the method of the invention, the array of test compounds is laid out onto a non-porous substrate (such as thin coverglass sheet) which is transparent or optically clear. Imaging will be through this surface, and through the cell support membrane lying above. The substrate (Fig. 4, 10) should be inert and solvent tolerant. For example, borosilicate glass sheets of about 200 microns thickness, which may be further surface-treated to give either hydrophobic or hydrophilic properties as desired. This embodiment is shown in the schematic of Fig. 4.

Detector layer: In one embodiment of the invention, the detector layer is a layer of

living cells cultured on a thin porous membrane. A porous membrane useful in the culture and transfer of cells is a transparent non-absorbent membrane with pores of regular and defined diameter which traverse the membrane directly from the upper to the lower side. A porous sheet suitable for cell growth is a track-etched polyester membrane about 10 microns thick with pores between 0.015 and 5 microns diameter at densities of between 600,000,000 to 400,000 pores/cm² repectively (Nucleopore® from Corning Costar).

Delivery of test compounds to detector layer. The porous membrane which supports the detector layer, complete with the buffer medium which overlays it, is applied onto the (dry) test array. Buffer medium wets the lower surface of the porous membrane (Fig. 4, 19) and forms a continuous thin film 23 between the array of test compounds 21 and the porous membrane 19. Test compounds diffuse up through the pores to the detector layer above. In one embodiment of the invention the detector layer is a monolayer of living cells overlayed with physiological buffer solution. The invention includes the possibility that under some conditions it is desirable to have cells grow processes through the membrane to make direct contact with substances on the test array below, with the use of a membrane having an appropriate pore diameter.

Further embodiments and general considerations. Where a test array is generated as a complex mixture of components, such as from the "teabag" method of combinatorial synthesis, or from cDNA library expression systems, a separation step may first necessary. Separation of test components may be conducted in any number of ways known to the art. In one embodiment, components may be separated by the use of one- or two-dimensional separation techniques in non-denaturing gels. The resulting gels may be used directly as test arrays.

Specific separation methods will be tailored to the components involved. Any bioactive compounds from such an array would be identified from identical copies of the original test gel.

Detection of Bioactivity.

Lens and illumination system. Specialized light sources and optics are needed to illuminate and image the fluorescence coming from an area the size of a microtiter plate (96-well plate). Such a system is available from: Imaging Research Inc., St Catherines, Ontario, Canada, and consists of a high-power light source directed through a specialized lens which

acts both as a wide-field epi-illuminator and imaging device.

An illumination system useful in the HDFS device is able to deliver excitation light over an area of at least 8.5 by 13 cm at an intensity sufficient to excite measurable fluorescence from that test field (which in most cases will be living cells loaded with fluorescent reporters). The illumination may come from a scanned beam, or be wide-field for simultaneous illumination of the entire area. The imaging system will collect fluorescent light from the entire test area and bring it to focus onto a sensitive imaging photodetector, such as a cooled CCD camera chip.

Screening. The practice of screening large libraries of samples of unknown composition for the few which may contain a compound of specific biological activity is one of the more common methods of new drug discovery. The samples of unknown composition are in most cases biological material, such as plant extracts or microbial fermentation broths. Screening these for biological activity is normally accomplished by performing binding assays or, more recently, functional assays. A binding assay is an attempt to find compounds of interest by identifying those which adhere with some desired affinity to cells or cell products. This can be done using fluorescent, luminescent, or radioactive detection methods. These assays are based not on a biological response, but passive processes of adherence and displacement. They cannot be construed as functional assays or as real-time assays. Another way to determine biological activity is to measure up-regulation or down-regulation of expression of a known genc. This is done by inserting DNA which codes for something which can be readily measured into a cell's genome such that the expression of interest is coupled to expression of the inserted DNA. While this is a true functional assay, it also is not a real time assay. In addition, it is only capable of finding compounds which affect gene expression. In many cases this is not the response of interest.

The CytoSensor described in U.S. Patent No. 4,915,812 and U.S. Patent No. 5,395,503 is a commercial instrument which has been billed as a screening instrument. It is based on the detection of increased cellular proton flux by means of a semiconducting electrode. The instrument is applicable to high through-put screening, but can only detect cellular events that result in changes in extracellular pH. Again, many responses of interest are not associated with changes in extracellular pH.

The growth over the last few decades in the knowledge of cellular signaling has presented extremely rich opportunities for new ways of screening for biologically active

compounds. Armed with knowledge of the biological process which one wants to affect with a new product, it is possible to monitor the actual process as a way of looking for compounds which affect it. The development of fluorescent probe molecules which upon interaction with intracellular signaling molecules (e.g. ions, enzymes, cyclic nucleotides) change their spectral properties has enabled the real-time monitoring of dynamic biological responses within living cells. Most of these probes can be introduced non-invasively into cells and will, depending on the detection system, allow characterization of cellular events in high temporal resolution (microseconds to seconds) and high spatial resolution (nanometers to micrometers). This probe technology, in combination with the technology of cellular imaging which is described below, has had a major impact on cell biology in that it has enabled monitoring of complex, cross-reacting intracellular events that could not be unravelled by conventional invasive biochemical techniques.

Imaging of cellular functions using luminescent probes. Visualization of intracellular function using luminescent (fluorescent or bioluminescent) probes has become one of the mainstay techniques in modern cell biology. Using traditional optical microscopes with quantitative detectors in place of the human eye, both the concentration and distribution in the cell of a variety of intracellular molecules of interest can be measured. While luminescent probes can be measured in large populations of cells using other techniques, imaging is the only way to learn what is going on in single cells or small populations of cells. The imaging capabilities of the HDFS apparatus will be limited to rather low spatial resolution - fluorescent changes will be imaged from the entire field of detector layer up to 8cm by 12.5 cm. When the detector layer comprises living cells, individual cells need not be resolved in the image, only the fluorescent signals from regions in which cells are present.

The imaging times will vary depending on the responses and parameters being monitored. Signaling responses, for instance changes in the level of free calcium in cellular cytoplasm, may first be seen within seconds or minutes following delivery of test compounds to the detector layer. Such changes can be monitored by changes in the fluorescent properties of specific chemical probes, for instance Fluo-3 or Fura 2 may be used to report on cytoplasmic calcium. The way in which these changes develop within cells (time-response profile) is an important diagnostic feature of the signaling processes giving rise to them. Rapid responses are therefore recorded by sequences of images, where the time between images in a sequence is between 0.1 and 30 seconds (depending on the response being

screened for). Transcription mediated events may require minutes to hours to develop. Monitoring may be continuos or intermittent. For slow responses, two images can be sufficient to gauge the level of response, the first taken before application of test compounds, the second after a period during which the response is estimated to have reached its maximum extent.

Controls relevant to the parameters being measured can be incorporated into the test arrays, both as a check for cell responsiveness and as co-ordinate markers within the arrays. The detector layer is continuous and undemarcated, but because of the close apposition of the test array to the detector layer, the center point of a response in the detector layer corresponds to a conjugate coordinate in the test array. It is helpful to have compounds in the test array which will generate known responses at known coordinates in the detector layer. Responses at the conjugate coordinates in the detector layer act as controls for the system's response, against which responses of the detector layer to unknown compounds may be compared; the points of response to control substances also act as reference points in the detector layer from which the coordinates of other responses can be mapped. For example, when bioactivity is determined as the ability to alter the level of free calcium in cellular cytoplasm, common calcium-mobilizing agonists such as carbamylcholine or adenosine trisphosphate are included in the test array at known coordinates.

As another example, when a change in the cellular ratio of inherently fluorescent NAD(P)H/FAD is the biological parameter being assayed, metabolic inhibitors such as KCN or rotenone may be used as a control and marker compounds.

In many instances, diffusion within a thin fluid layer will be involved in many applications of the screening method of the invention, and a concentration gradient will be established from each test point. Those few compounds in a test array which have bioactivity should be detected as spreading rings of response from the focus point of diffusion, within a field of the detector showing no response. The extent of the response areas (measured over time), compared with those from control substances, will provide an indication of potency and solubility of the compound responsible, and also obviate the need to make serial dilutions of test compounds. Toxic or inhibitory substances may also be determined by causing blank sectors in response rings from known agonists. Inhibitory compounds may be determined by their actions on a (pre-)stimulated detector field. Detection of bioactive compounds may incorporate simple image processing to determine the focus, extent and potency/efficacy from

the areas of activity measured in a detector field.

Apparatus

In specific embodiments, the apparatus and method of the invention are as shown in Figs. 1-4. Fig. 1 shows a high energy light source 1, either a mercury or xenon arc lamp, light from which is collected and collimated by unit 2, directed through a shutter assembly 3 and passes through a excitation filter-changer 4. A high-quality light guide 5, either of fused quartz or a UV-compatible liquid light guide, directs excitation light into the lensing and epi-illumination optics housed in unit 7. Excitation light emerging from 7 evenly illuminates the horizontal detector layer located in the multi-component assembly labeled 10 and 11.

Further details of this assembly are shown in Figs. 2a-c, 3, and 4. The assembly comprises two solid layers of which 10 is fixed relative to the stage unit 8 which supports it, while layer 11 is moved vertically downward on guide pins (17 in Figs. 2a,b,c) to bring test compounds into contact with the detector layer. Vertical movement of 11 is controlled by arm 12 driven by unit 13. Four sprung contacts 14 attached to 12 press upon the frame of layer 11 to drive it downwards as arm 12 descends. A separate drive unit 9 controls position of the stage 8 in the horizontal plane, and also is used to adjust focus by movement along the vertical axis.

Fluorescent light emitted by the detector layer is collected by lensing unit 7, passes through an emission filter-changer 15 and is brought to focus on the photosensitive surface of an imaging detector housed in unit 16.

Specified devices (3, 4, 9, 13, 15, 16) are controlled by a central processing unit 6 which issues commands to, and collects data and status information from the devices attached to it. Collected data (images) can also be analyzed by unit 6, or passed to a subsidiary analysis station (not shown). Unit 6 comprises: central processing unit (Intel Pentium chip, or better), RAM, multi-channel serial input/output cards with onboard A/D and D/A converters, one of which cards controls the camera 16 and captures images from it, also a video controller card, VDU, and hard disk memory units.

Figs. 2a,b,c are schematic diagrams of the test stage, which includes a supporting stage 8 with large rectangular central aperture, the shape and size of which is the same as the area labeled 19. The position of stage 8 is adjusted in the horizontal and vertical axes by the

3-axis positioner 9. These diagrams are drawn for the specific embodiment in which the detector layer is a layer of living cells growing on the upper surface of the solid transparent component 10, which also serves to contain the liquid layer 18 which overlays the cells in the detector layer and provides them with necessary nutrients and conditions to keep them alive. The printed array of test compounds 21 is borne on a sheet of track-etched membrane 19 held by a rectangular rigid frame 11. At the beginning of the screening assay, the array 21 is not in contact with the fluid layer 18. The array 21 is held away from the liquid layer by pins 17 which pass through holes 24 in the corners of the frame 11 and which, by friction or "click-stops", prevent it from falling (Fig. 2a). At the appropriate moment, arm 12 is moved down by the drive unit 13 and the four sprung contacts it bears 14 exert pressure on the frame 11 moving it down the guide pins 14 and into the liquid 18 below to a position where it is in very close proximity to the underlying layer of detector cells 20 grown on top of the solid substrate 10 (Fig. 2b). Throughout this procedure, the entire area of the detector layer corresponding to the size and shape of area 19 is illuminated and imaged from below by the additional apparatus shown in Fig. 1.

The apparatus can also be used in a second embodiment of the screening method of the invention, where the test array is laid out on the upper surface of component 10, and components 11 and 19 are a frame and thin transparent track-etched membrane, respectively. In this specific embodiment, the frame 11 is sufficiently deep to contain culture liquid as required to sustain the detector layer of living cells growing on the upper surface of the membrane 19.

Figs. 3 and 4 are schematics to show the relative positions of the different layers in the test-array/detector layers used in the specific embodiments of the invention. Fig. 3 shows the arrangement in which an array of discrete spots of test compounds 21 on a porous membrane 19 is in contact with a liquid layer 18 overlaying the detector layer 20 which is supported by an optically transparent solid substrate 10. The compounds fill the parallel capillary spaces 22 in the track-etched membrane 19. Bioactivity is detected by measuring changes in fluorescence in the detector layer 20 resulting from responses to the diffusion of test compounds through the porous membrane to the detector layer.

Fig. 4 is a schematic drawing of a second embodiment of the screening method in which a detector layer 20 supported on an optically clear porous membrane 19, and overlayed

by a liquid layer 23, is placed onto an optically clear solid substrate 10 bearing an array of test compounds 21. The thin space 18 between components 19 and 10 is filled with solution from 23 which has passed through the porous membrane 19. Bioactivity is again detected by measuring changes in fluorescence of the detector layer resulting from responses to the diffusion of test compounds through the porous membrane to the detector layer.

Fig. 5 is a schematic illustrating the way in which an array of 1536 compounds can be created on a membrane surface, such as would be useful in the first embodiment described above, by simple transfer printing. Compounds are stored in 16 separate 96-well microtiter plates and defined amounts are transferred simultaneously by a 96-pin printing head to the surface 19. The contents of each successive 96-well plate are printed at a slightly offset position, generating an array as shown in Fig. 5b after 4 such printing operations, and a full array of 1536 compounds (Fig. 5c) after 16 printing operations. The holes 24 in frame 11 are used to position and guide the completed array on the pins 17 indicated in Figs. 2b and 2c. The process illustrated in Fig. 5 can also be used to transfer an array of test compounds to a solid surface such as would be useful for component 10 in the second embodiment of the method described above.

EXAMPLE

Example 1. Screening of 1536 Test Compounds for Bioactivity.

The following description of the use of one embodiment of the apparatus of the invention in the screening method disclosed. An array of test compounds are supplied in 96-well microtiter plates, as is common practice for compounds produced by methods commonly known as combinatorial chemistry, or for compounds extracted from natural sources. In this example, the compounds are provided in soluble form, and the concentrations and solvents used have previously been tested for compatibility with the apparatus. In this example, 1536 compounds are tested simultaneously against a known cellular target, specifically a G-protein coupled receptor (GPCR) of the Gq type expressed in a transformed cell line. Gq GPCRs give clearly identifiable changes in intracellular calcium when activated.

First, physiologically viable living cells are cultured to a near confluent monolayer in a transparent culture dish (10, Fig. 2a-c) in appropriate culture medium and conditions.

Immediately prior to being used in the experiment, the cells are loaded with the fluorescent

indicator of free cytoplasmic calcium concentration, Fluo-3 (from Molecular Probes, Oregon). This is accomplished by incubating the cells with a 2 to 5 μ M solution of Fluo-3 acetoxymethyl ester (AM) for a period of 10 to 15 minutes, followed by a series of solution exchanges to wash away excess Fluo-3 AM.

The method of transfer of compounds to the track-etched membrane Fig. 2a-c 19 is illustrated in Fig. 5. In this example, 1536 compounds are printed as an array 21 on a single track-etched membrane 19, from sixteen individual 96-well microtiter plates in the following manner: A 96-pin printing head is used to transfer defined volumes of compounds (in the range 0.05 to 0.5 µl of each compound), one compound per pin, from each 96-well plate in turn (with wash steps between source plates to avoid cross-contamination). Each 96-point print to the membrane occurs in an offset grid, such that 16 print operations are made sequentially on the same membrane and the printed spots of compounds remain discrete and separated from each other (three of these spots are indicated in Fig. 5a, 21). Fig. 5a shows the result of a single 96-point print operation, Fig. 5b after four such operations, and Fig. 5c the finished array after 16 print operations. In this way, just sixteen print operations (and sixteen intermediate wash steps for a single print head) are sufficient to transfer 1536 compounds to a single test array. The procedure can be readily automated, and multiple copies of each printed sheet made for multiple tests.

Completed arrays are fixed to the pins 17 (Figs. 2b-c) projecting from the culture dish 10 such that they are supported some small distance above the thin fluid layer 18 covering the living cells which form the detector layer. Once the test array is fixed in place over the Fluo-3-loaded cells, the entire assembly is placed onto the test stage as shown in Fig. 2a.

The following events are synchronized by sequential instructions from the computer processing unit 6. First, the test stage is centered over the lensing unit 7 (Fig. 1) and the detector layer it supports is brought into focus by the motor unit 9. Fluo-3 is excited by light of 490 nm, and its fluorescent emissions are collected in the range 505-540 nm. The intensity of emission is increased when the dye binds free calcium. Thus the computer brings a 490 nm band-pass excitation filter into line of the light path coming from units 1 and 2 using the filter changer unit 4. At the same time, a band-pass emission filter for the range 505-540 nm is positioned in the imaging path by unit 15. The shutter 3 is opened for a pre-determined exposure period (typically 50 to 500 milliseconds), and during this time the whole area of the

detector layer is illuminated with 490 nm light. Fluorescent emission from the Fluo-3 in the cells is collected by the lens 7 and focused into the camera. The camera captures the image and sends it to the processing unit 6 where it is stored and displayed. At regular intervals thereafter, images are captured in sequence by repeatedly opening the shutter 3. Intervals between successive images are typically in the range 0.5 to 30 seconds, depending on the speed of the response expected. Intervals of 0.5 to 2 seconds are usual and sufficient to sample the dynamics of most changes in cellular calcium. At a predetermined time during this continuing sequence of images, the test array is pushed down the guide pins 17 by the actuating arm 12 and its sprung contacts 14, driven by unit 13. In close apposition to the cells in the detector layer, the test array begins to release the compounds it carries. The compounds dissolve into the liquid layer, and these fall vertically downwards onto the cells below. Because there is only a thin liquid layer between the membrane of the test array and the cells below, there is insignificant intermixing of adjacent test compounds. If a test compound activates cells below it bearing Gq GPCRs, these cells will respond with an immediate increase in free cytoplasmic calcium, and the fluorescence signal from the Fluo-3 dve they contain will increase. The sequence of images collected during the period of the response (which is typically of 1 to 10 minutes duration) will reveal which cells have so responded, and their position in the area of the detector layer will be correlated with the identity of the compound in the array above. An analysis of the entire area of each image in the sequence, performed on-line by the processing unit 6, yields the following information: the identity of the compound eliciting the response, the profile of the response with time, the intensity of the response, and also the potency of the compound with reference to a chosen standard. The latter information is contained in the radius of the area of cells responding within a particular time, and can be compared directly to a known standard which is included in the array at known points. The use of standard compounds at known points in the array also provides a control for the experiment, and helps to identify coordinates in the detector layer from which other responses can be mapped.

At the end of the screening assay, the sequence of images is stopped, the actuating arm 12 raised, and the test assembly removed. The next assembly is then moved in and the sequence begun afresh. Assembling the test units and exchanging them on the test stage can be automated by appropriate robotic control (not shown in the diagrams).

One of the advantages of the method of the invention is that the method does not require that either the components of the detector layer (e.g. living cells), or the different test compounds, be isolated from one another within discrete chambers or compartments, as is common to all high throughtput screening procedures currently in use or development. The method also removes the need to dispense microvolumes of test compounds during the period of the assay itself. Delivery of test compounds to detector layers is either by direct contact or by simple diffusion across thin liquid films. Delivery and detection becomes a (massively) parallel process.

CLAIMS

What is claimed is:

- 1. A method for screening test compounds for bioactivity, comprising:
- (a) contacting an array of test compounds with a detector layer; and
- (b) detecting a detector layer response, wherein a response is indicative of bioactivity.
- 2. The method of claim 1, wherein the detector layer is comprised of physiologically viable cells.
- 3. The method of claim 2, wherein the detector layer is supported by an optically clear substrate.
- 4. The method of claim 3, wherein the reactive sensing surface is held stationary in the field of view of the optical detector and the sample surface is moved into contact with it during the course of the measurement.
- 4. The method of claim 1, wherein the detection of step (b) is a change in a fluorescence or luminescence property of the cell.
- 5. The method of claim 4, wherein detection is determined with an illumination system capable of exciting the fluorescence of the reactive surface with any of a number of previously selected wavelengths with defined order and of defined time duration.
- 6. The method of claim 2, wherein the physiologically viable cells form a monolayer.
- 7. The method of claim 1, wherein the test compounds are generated on a solid support by combinatorial chemistry.
- 8. The method of claim 1, wherein the test compound array is generated by one- or two-dimensional gel electrophoresis.

- 9. A method for high throughput screening of test compounds for bioactivity, comprising:
- (a) contacting a solid support comprising an array of multiple test compounds with a cell layer, wherein each test compound comes into contact with a localized liquid which is in contact with a detector layer; and
- (b) detecting a response of the detector layer to the test compound, wherein a response is indicative of a bioactive compound.
- 10. A method for simultaneously exposing an array of test compounds with a reactive sensing surface, comprising the steps of:
- (a) contacting an array of test compounds on a solid matrix with a porous membrane which is in contact with a liquid layer overlaying a reactive sensing surface layer; and
- (b) allowing the test compounds to diffuse through the porous membrane to the liquid layer overlaying the reactive sensing surface.
- 11. An apparatus for screening an array of test compounds for bioactivity, comprising:
 - (a) a solid support comprising an array of test compounds;
 - (b) a porous membrane; and
- (c) a detector layer layer, wherein a liquid layer is between the porous membrane and detector layer layer, and wherein the test compounds contact the detector layer layer by diffusion through the porous membrane.

METHOD AND APPARATUS FOR HIGH DENSITY FORMAT SCREENING FOR BIOACTIVE MOLECULES

Abstract

A method and apparatus for screening an array of test compounds for bioactivity by contacting an array of test compounds with a detector layer capable of detecting bioactivity, and detecting a detector layer response. The detector layer is comprised of physiologically viable cells. The method and apparatus allow a large number of test compounds to be simultaneously assayed in parallel.

Fig. 1 Schematic view of equipment; not to scale

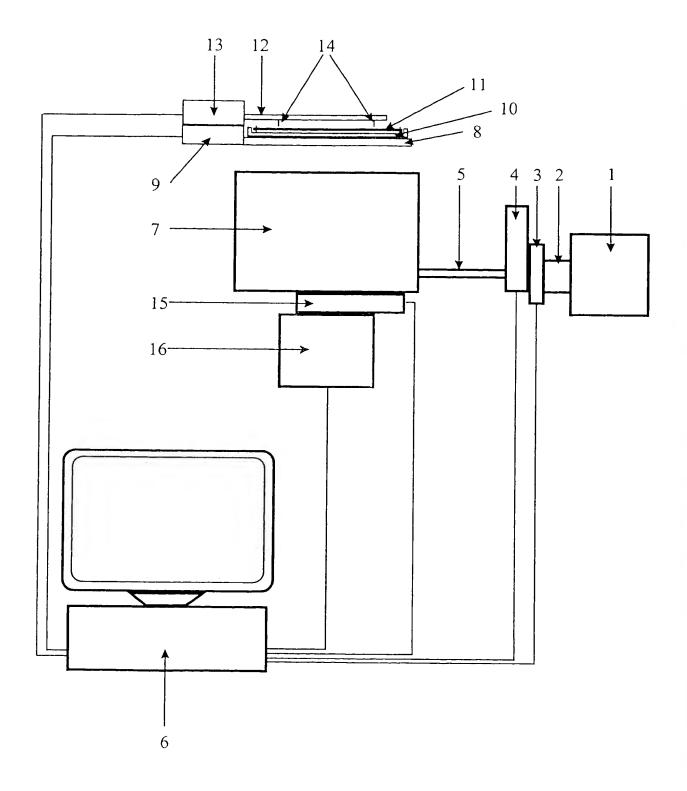
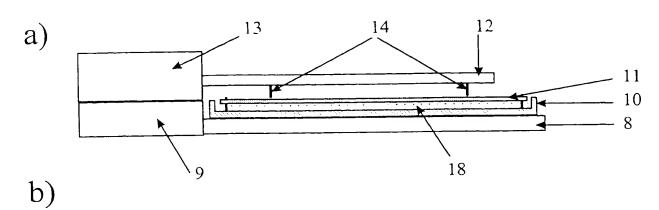
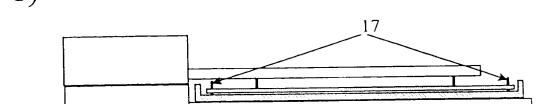
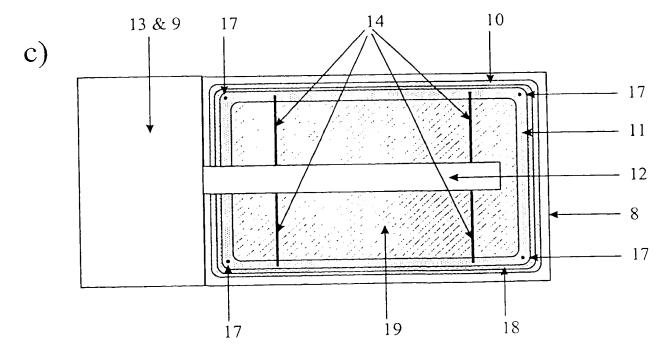


Fig. 2

Side views of test stage; not to scale

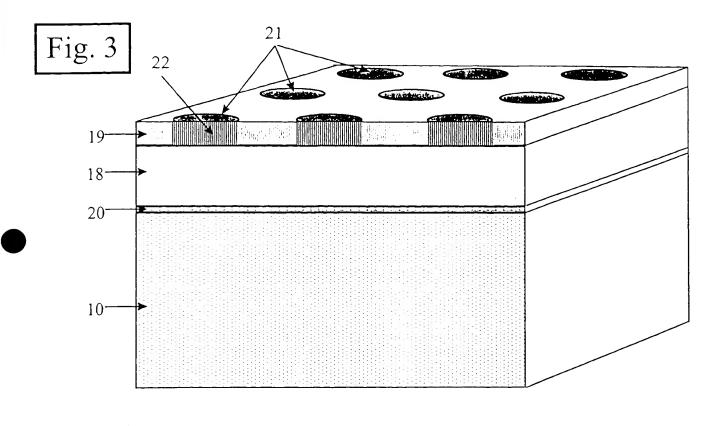


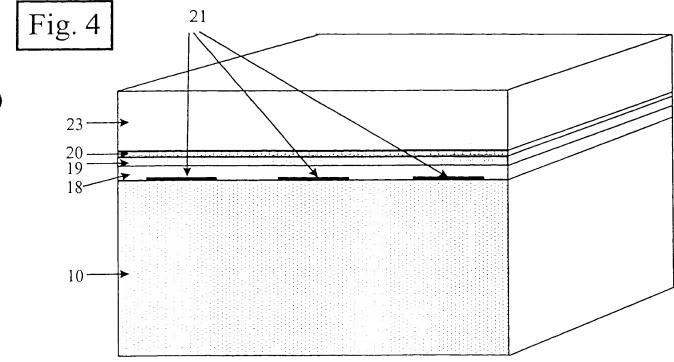




Top view of test stage; not to scale

3-D sectional representations of portions of the test-array/detector layers: not to scale





Changes in intracellular cAMP visualised using a cAMP-dependent protein kinase-green fluorescent protein hybrid.

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ABSTRACT

A novel method to monitor changes in intracellular cAMP concentration ([cAMP]) within intact living cells has been developed based on a fusion of the catalytic subunit of cAMP-dependent protein kinase to green fluorescent protein (GFP). In stably transfected unstimulated fibroblasts, fusion protein fluorescence was highly concentrated in aggregates throughout the cytoplasm and absent in the nucleus. Stimulation with the adenylate cyclase activator forskolin caused the release of tagged catalytic subunits from the cytoplasmic aggregates within minutes, resulting in an increasingly homogeneous distribution of GFP fluorescence throughout the cytoplasm. The observed redistribution was completely reversible: removal of forskolin led to the return of fluorescence to the cytoplasmic aggregates. Spot-photobleach measurements showed that the rate of exchange of GFP-labelled catalytic subunits at these aggregates increased in proportion to [cAMP]. The localisation of the fusion protein was also sensitive to receptor stimulation. In fibroblasts stably expressing the G-protein coupled glucagon receptor, generation of an increased [cAMP], through glucagon stimulation resulted in a redistribution of tagged catalytic subunit similar to that observed after forskolin addition. Conversely, in fibroblasts overexpressing the G-protein coupled α2a adrenoreceptor, addition of norepinephrine after forskolin stimulation led to a reversal of the fusion protein redistribution.

INTRODUCTION

The cAMP-dependent protein kinase (cAK)¹ is a ubiquitous serine/threonine protein kinase. cAK is recognised as the only mediator of intracellular cAMP signals in eukaryotes², with the exception of certain ion channels³. The cAK holoenzyme is an R_2C_2 tetramer consisting of a regulatory (R) dimer and two catalytic (C) subunits². Presently, four isoforms of the regulatory subunit (RI α , RI β , RII α and RII β) and three isoforms of the catalytic subunit (C α , C β and C γ) have been described². Splice variants of C α and C β ⁴ and possible R heterodimers, as reported for RI α and RI β ⁵, add to the complexity of the cAK holoenzyme. Although the C γ isoform is unique with respect to substrate specificity, inhibition and tissue distribution⁶, few reports suggest different roles for C α and C β isoforms of the catalytic subunit⁷. In contrast, the RI and RII subunits are reported to be distinct. The cAKI (RI₂C₂) holoenzyme is thought to be mainly soluble and cytoplasmic² although RI is reported to be associated with

sarcoplasmic membranes⁸ and also with a detergent-resistant structure in mammalian sperm⁹. cAKII (RII₂C₂) on the other hand is thought to be particulate and RII has been reported to bind to a number of intracellular components, most notably Golgi membranes^{10,11} and centrosomes^{10,11} but also mitochondria¹², nuclei^{13,14} and cytoskeletal components^{11,12}. RII subunits interact with a family of proteins called A-kinase anchoring proteins (AKAP)¹⁵ and this may also be true of RI subunits¹⁶. The AKAP-RII subunit interaction is presumed to be responsible for localising the cAKII tetramer at these intracellular sites. The NH₂-terminus of the C subunit is myristoylated¹⁷, a post-translational modification usually associated with membrane insertion. However, the C subunit does not appear to be membrane attached and while myristoylation may increase the thermostability of the protein, the possible role of myristoylation in its targeting or substrate specificity is still not clear¹⁸.

The C subunit in the assembled tetramer is believed, although not unanimously¹⁹, to be catalytically inactive. Activation of cAK is physiologically mediated through G_s-protein coupled plasma membrane receptors. G_s-protein activation leads to activation of adenylate cyclases, which generate cAMP. Binding of two molecules of cAMP to each R subunit causes the release and activation of the C subunits. Dissociated C subunits phosphorylate cytoplasmic substrates^{20,21} and have been shown to relocalise to the nucleus²². The nuclear redistribution mechanism of C subunits may be by simple diffusion through nuclear pores²¹. To date a large number of cytoplasmic and a few nuclear cAK substrates have been reported. An incomplete list of 25 *in vitro* substrates²³ includes several enzymes involved in basic metabolism such as phosphorylase kinase, glycogen synthase and fructose bisphosphatase. Nuclear C subunit regulates transcription of genes under control of the cAMP response element (CRE) by phosphorylating the continuously bound CRE binding protein, (CREB)^{24,25}.

Several factors decrease the level of cAK activity. Stimulation of plasma membrane bound G_i-protein coupled receptors inhibits adenylate cyclases and cAMP is continuously being broken down by a variety of phosphodiesterases. Despite the importance of the cAMP/cAK signalling pathway, there is no easy method to monitor intracellular cAMP concentrations ([cAMP]_i) in intact living cells. The current method of choice involves fluorescence resonance energy transfer (FRET) between microinjected fluorescently labelled R and C subunits²⁶. In the work described herein, the Cα subunit was tagged with a highly fluorescent variant of green fluorescent protein (GFP) containing F64L and S65T amino acid substitutions (GFP^{LT}) (International

Publication No. WO97/11094). This approach provides a transfectable probe for monitoring the intracellular trafficking of C subunits in response to changes in [cAMP], and represents the first easy method to evaluate changes in [cAMP], in intact living cells in response to extracellular signals.

Results

GFP^{LT} tagged C had the expected molecular weight.

Lysates of glucagon receptor-transfected baby hamster kidney cells (BHK/GR) stably expressing the C-GFP^{LT} fusion protein were characterised by Western blot analysis using polyclonal antibodies directed against the NH₂-terminus of C α (Fig. 1). In a separate experiment, lysates of BHK cells, transiently expressing either of the two fusion proteins, were characterised by Western blot analysis using polyclonal antibodies that recognise GFP (data not shown). Taken together, these experiments show that C-GFP^{LT} fusion protein is recognised as a unique protein of the expected size by the anti-C α antibody in stably transfected cells and that both fusion proteins have the same molecular weight.

The fusion protein localised to cytoplasmic aggregates.

The localisation of the two fusion proteins, when transiently expressed in Chinese hamster ovary (CHO) cells, was very different. While GFP^{LT}-C was evenly distributed throughout the cytoplasm (Fig. 2A), C-GFP^{LT} was found in highly fluorescent aggregates in the cytoplasm (Fig. 2B). These distinct patterns for the two fusions was also seen in transiently transfected human embryonic kidney (HEK293) and BHK/GR cells (data not shown). For unknown reasons it was not possible to make stable transfectants expressing the GFP^{LT}-C fusion, whereas this procedure was straightforward with the C-GFP^{LT} fusion. The distribution of GFP^{LT}-C in transiently transfected CHO cells did not change when [cAMP], was raised by the addition of 50 µM forskolin (n=6, data not shown). The following results are therefore based only on work with the C-GFP^{LT} fusion.

Increased [cAMP]_i caused the release of fusion protein from cytoplasmic aggregates.

Within 2-3 minutes of treatment of CHO/C-GFP^{LT} cells with forskolin, C-GFP^{LT} fluorescence dispersed from the bright aggregates and filled the cytoplasm (Fig. 3A, 1 μM forskolin), remaining in this distribution for as long as forskolin was present (cells were followed up to two hours). The probe did not enter the nuclear compartment to any clearly observable extent. Higher doses of forskolin increased the rate and extent of probe redistribution. The responses depicted in Figure 3B-G have all been quantified from image data, as described in the experimental protocol. Table 1 gives a comparison of the average temporal profiles of fusion protein redistribution in response to the three forskolin concentrations shown in Figure 3B. Addition of 1 mM dibutyryl cAMP (dbcAMP) (n=6), a membrane permeable cAMP analogue, which is not degraded by phosphodiesterases, caused a similar but slower response (Fig. 3C). Addition of 100 μM 3-isobutyl-1-methylxanthine (IBMX) (n=4), a cell permeable phosphodiesterase inhibitor, caused a similar, slow response (Fig. 3D), even in the absence of adenylate cyclase stimulation. Addition of buffer (n=2) had no effect (data not shown). As a control for the behaviour of the fusion protein, GFP1.7 alone was expressed in CHO cells and these also given 50 μM forskolin (n=5); the uniform diffuse distribution characteristic of GFP in these cells was unaffected by such treatment (data not shown).

To test the reversibility of the fusion protein redistribution, CHO/C-GFP^{LT} cells were treated with 10 μ M forskolin (n=2) and washed repeatedly (5-8 times) with 37°C buffer. Although the plant terpenoid forskolin is lipophilic, it is possible to remove its effect by washing with aqueous buffer²². In these experiments, fusion protein began to return to its prestimulatory localisation within 2-3 min (Fig. 3E). In fact the fusion protein returned to a pattern of fluorescent cytoplasmic aggregates virtually indistinguishable from that observed before forskolin stimulation. To test whether the return of fusion protein to the cytoplasmic aggregates reflected a decreased [cAMP], cells were treated with a combination of 10 μ M forskolin and 100 μ M IBMX (n=2); when washed repeatedly (5-8 times) with 37°C buffer containing 100 μ M IBMX the fusion protein did not return to its prestimulatory localisation after removal of forskolin (Fig. 3E).

To test the probe's response to receptor activation of adenylate cyclase, stably transfected BHK/GR,C-GFP^{LT} cells were exposed to glucagon stimulation. In these cells, addition of 100 nM glucagon (n=2) caused the release of C-GFP^{LT} from the cytoplasmic aggregates and a resulting permanent redistribution of the fusion protein to

a more even cytoplasmic distribution within 2-3 min (Fig. 3F). Similar but less pronounced effects were seen at lower glucagon concentrations (n=2, data not shown). Addition of buffer (n=2) had no effect over time (data not shown). CHO/C-GFP^{LT} cells, transiently transfected with the α 2a adrenoreceptor (AR α 2a), were treated with 10 μ M forskolin then, in the continued presence of forskolin, exposed to 10 μ M norepinephrine to stimulate the exogenous adrenoreceptors. This treatment led to reaggregation of C-GFP^{LT} within the fluorescent structures, consistent with a receptor-induced decrease in [cAMP] (Fig. 3G).

Rate of recovery from photobleach of C-GFP^{LT} aggregates is dependent on forskolin concentration.

Photobleach measurements were made to confirm that changes seen in the distribution of C-GFP^{LT} fluorescence were a result of changes in the rate of turnover of C-GFP^{LT} upon the aggregates. The fluorescence of an entire C-GFP^{LT} aggregate within a cell could be effectively bleached within 2 to 5 seconds by a stationary laser beam at full intensity. After bleaching, aggregates recovered their fluorescence, indicating a dynamic exchange of C-GFP^{LT} at these loci (Fig. 4A). The rate of recovery from spot photobleach was highly reproducible at each particular concentration of forskolin even in different cells (Fig. 4B). Both the extent and rate of recovery increased with the forskolin treatment given. Most recovery curves required at least two exponentials to fit them adequately. Given the limits of the experimental procedure, the curves are used here only to estimate half-times of recovery. To an approximation, half times for recovery can be estimated directly from the slope of reciprocal plots of the fluorescence displacement for the first few time points²⁷. Values for half times estimated within the first 3.0 seconds of recovery (Fig. 4C) are plotted as a dose response curve in Figure 5, giving an estimated ½-maximal concentration for forskolin of about 3 μM

Fusion protein redistribution correlated with [cAMP]i

As described above, the time it took for a response to come to completion was inversely related to the forskolin dose (Table 1). In addition the extent of a response was also dose dependent. In an automated imaging system we stimulated CHO/C-GFP^{LT} cells with 5 increasing doses of forskolin (n=8). Images were analysed with the same algorithm used

to construct Figure 3B-G. From the results shown in Figure 5, a half maximal stimulation was observed at 1.7 μ M forskolin by this method. In parallel, CHO/C-GFP^{LT} cells were stimulated with 8 increasing concentrations of forskolin (n=N) and the relative amount of cAMP produced was measured in a scintillation proximity assay (SPA). The ½-maximal concentration for forskolin in the SPA assay was determined to be 9.3 μ M (Fig. 5).

Co-localisation of C-GFP^{LT} with labelled ceramide distributions

Figure 6A is an overlay of green and red fluorescence emissions from CHO/C-GFP^{LT} cells stained with BODIPY[®] FL C₅-ceramide (ceramide-FL). The green channel contains the ceramide-FL and GFP^{LT} fluorescence; the red channel shows only the ceramide-FL excimer emission. The ceramide-FL probe preferentially accumulates in Golgi membranes²⁸. This is most obvious in images formed from the red excimer emissions of the FL-ceramide. The GFP^{LT}-bright structures do not stain with the ceramide probe indicating that they are clearly distinct from Golgi membranes.

Structure of the GFP^{LT} -bright aggregates

Figure 6B shows an iso-surface rendering of 25 deconvolved and reconstructed through-focus wide-field images of a single large C-GFP^{LT} aggregate. Each aggregate appears to have the structure of a convoluted tubule or glomerulus, and this is more obvious in the stereo pair (Fig. 6C) derived from the same data set from which the iso-surface rendering was made. It is not completely clear whether each structure is formed from a single fully connected tubule or a small number of discrete tubules in close apposition. The structure is however clearly compact and more complex and structured than a simple amorphous aggregation of C-GFP^{LT} molecules. Figure 6B-C is typical of the larger aggregates which are of the order of 2 to 4 μm across. The more numerous smaller aggregates (less than 1 μm across) appear to share the same underlying structural component(s) as their larger counterparts.

Discussion

The aim of the present study was to develop a transfectable probe for monitoring changes in [cAMP],. Since cAK is by far the major intracellular effector for cAMP², a measure of its activation should closely reflect physiologically relevant changes in [cAMP].

 NH_2 - and COOH-terminal fusions of C subunit were made to a highly fluorescent variant of GFP. Only the C-GFP^{LT} fusion responded to changes in [cAMP], The three-dimensional structure of the C subunit^{29,30} reveals that both the NH_2 - and COOH-termini, while far apart, are both located opposite the catalytic cleft and close to the surface of the protein. Comparison with the closely related cGMP-dependent protein kinase, whose R and C subdomains are contained within the same polypeptide chain in R-C order³¹, suggests that the R subunit of cAK may be expected to interact with the NH_2 -terminal region of the C subunit. Furthermore, the surface of the C subunit in the NH_2 -terminal region is hydrophobic²⁹, supportive of a protein-protein interaction in this area. An NH_2 -terminal GFP^{LT} tag would also prevent post-translational myristoylation (of the NH_2 -terminus) of the C subunit as reported specifically for mouse $C\alpha^{18}$, while the C- GFP^{LT} fusion may well be myristoylated. These factors may explain the very different behaviours of the NH_2 - and COOH-terminal fusions of C subunit to GFP^{LT} .

There are reasons to believe, that the C-GFP^{LT} fusion protein behaves like the endogenous kinase both with regard to localisation and activation kinetics. Li *et al.* (1996)¹¹ have, for instance, reported that RII subunits occur as "intensely fluorescent spots" within perinuclear cytoplasm. Skålhegg *et al.* (1997)³² also reported a granular distribution of RII in both human B and T lymphocytes. Also, the time frame of fusion protein redistribution in response to forskolin addition reported here, corresponds well to the observation of dissociation of microinjected RI α_2 C α_2 holoenzyme in response to forskolin within 1-2 minutes²⁶ and the dissociation of endogenous RII₂C₂ in response to forskolin observed by immunofluorescence after less than 5 min²².

In contrast with previous work with microinjected RIIa₂Ca₂ holoenzyme and Ca subunit²⁴, we did not observe any translocation of C-GFP^{LT} to the nucleus. A possible explanation could be the increased size of the fusion protein relative to endogenous C subunit. Nuclear pores are thought to allow passage by diffusion of globular proteins of less than 45-60 kDa³³. The putative size limit of 45-60 kDa may adequately explain the exclusion of the fusion protein (68 kDa), yet passage of endogenous C subunit (41 kDa).

Consistent with this, a microinjected 65 kDa fusion protein of glutathione S-transferase and mouse Ca subunit (GST-C) was excluded from the nucleus²¹.

That the C-GFP^{LT} fusion can be released by dbcAMP or treatments which increase [cAMP], suggests that it must recognise and attach to endogenous R subunits (or some subset of the same) and therefore that these R subunits are naturally collected at or on the structures seen in Figures 3A and 6. Reversal of elevated [cAMP], e.g. by removal of forskolin or stimulation of G-coupled receptors, results in rapid return of fluorescence to the original prestimulatory locations within cytoplasm. These anchoring structures therefore appear to be persistent features within the cytoplasm of CHO/C-GFP^{LT} cells. Similar structures and C-GFP^{LT} behaviour were also found in transfected BHK cells.

The distribution of fluorescence between aggregates and cytoplasm should reflect the position of a dynamic equilibrium within each cell, determined principally by [cAMP]. This is confirmed by results from spot-photobleach measurements. The rate of fluorescence recovery of aggregates following photobleach measures the net rate of turnover of C subunits at these sites. The rate of recovery is the sum of on and off rates for the association of catalytic with regulatory subunits at these loci, both of which will be governed principally by the concentration of cAMP within the cell (the off rate being governed directly by [cAMP]; the on rate being dependent on the concentration of free C-GFP^{LT} in the cytoplasm). Most aggregates completely disappear after full stimulation with forskolin. However, often one aggregate remains, and this is always the biggest and brightest from the unstimulated cell. Nevertheless, as photobleaching can demonstrate, there is active turnover of C-GFP^{LT} even at these large fluorescent aggregates which remain in fully stimulated cells. As a further observation, there appears to be considerable mobility of catalytic subunits within the structure of an aggregate, since a stationary laser beam (approx. 0.5-1.0 µm diameter) is able to bleach fluorescence from an entire aggregate of 2-3 µm diameter in 2 to 5 seconds.

The lack of colocalisation of C-GFP^{LT} and ceramide fluorescence, the position of aggregates within the cell and their unusual form, suggest that these structures are definitely not associated with Golgi, but may well be constructed of membrane tubules with C-GFP^{LT} on the outer surface. Although we have been unable as yet to ascertain the identity of these structures, we have ruled out Golgi membranes. They may however be membranous since fusion protein is apparently freely mobile on them, possible tubular judging by the 3-D recontructed image, and clearly the catalytic subunits are able to

bind to and release from R subunits with ease, suggesting that the latter are anchored to the surface of these structures. They are also persistent within the cytoplasm, and found in all cells transfected thus far with the C-GFP^{LT} construct (CHO, HEK293 and BHK).

Figure 5 gives a comparison of an SPA assay conducted in parallel with two different forskolin dose response experiments using the cAK fusion protein. These experiments showed a direct correlation of three parameters: level of [cAMP], turnover rate of C-GFP^{LT} at cytoplasmic aggregates, and overall degree of fusion protein redistribution. Data from these three greatly varying methods agree on an ½-maximal concentration for forskolin of between 1.7 to 9.3 μM in this system. As these results show, the cAK fusion protein represents a novel and reliable probe by which dynamic changes in [cAMP], can be measured in intact living cells as they respond to extracellular signals.

Experimental protocol

Hybrid cDNA construction

Hybrid cDNAs encoding NH_2 - and COOH-terminal fusions of murine $C\alpha$ subunit³⁴ to GFP^{LT} were inserted into the multiple cloning site of the pZeoSV (Invitrogen Corp., San Diego, CA, USA) mammalian expression vector, generating the fusion constructs C-GFP^{LT} and GFP^{LT}-C. Briefly, cDNAs encoding C and GFP^{LT} were amplified by PCR 5'-C. primers: following using TTGGACACAGCTTTGGACACCCTCAGGATATGGGCAACGCCGCCGCCCCC 3'-C. GTCATCTTCTCGAGTCTTTCAGGCGCGCCCAAACTCAGTAAACTCCTTGCCA 5'-GFPLT, CAC TTGGACACAGCTTTGGACACGGCGCGCCATGAGTAAAGGAGAAGAACTTT 3'-GFPLT. and TC GTCATCTTCTCGAGTCTTACTCCTGAGGTTTGTATAGTTCATCCATGCCATGT . HindIII/AscI restriction endonuclease digested C subunit PCR amplification product and AscI/XhoI digested GFPLT PCR product were ligated with the HindIII/XhoI digested vector for the generation of the C-GFP^{LT} fusion construct. Correspondingly the GFP^{LT}-C construct was generated by ligating HindIII/Bsu36I digested GFP^{LT} PCR product and Bsu36I/XhoI digested C subunit PCR product with the HindIII/XhoI digested vector. To generate a similar construct which allowed the expression of GFP^{LT} alone, the GFP^{LT} PCR product was digested with HindIII/XhoI and ligated with the HindIII/XhoI digested vector.

Cell cultures

CHO cells were transfected with the vectors containing hybrid cDNA for the C-GFP^{LT} or the GFP^{LT}-C fusion proteins using the calcium phosphate precipitate method in HEPES-buffered saline³⁵. Stable transfectants were selected using 1000 μg Zeocin/ml (Invitrogen) in the growth medium (DMEM with 1000 mg glucose/l, 10 % foetal bovine serum (FBS), 100 μg penicillin-streptomycin mixture ml⁻¹, 2 mM L-glutamine purchased from Life Technologies Inc., Gaithersburg, MD, USA). Untransfected CHO cells were used as the control. To assess the effect of glucagon on fusion protein redistribution, the constructs were stably expressed in BHK/GR cells (Novo Nordisk, Bagsværd, Denmark) overexpressing the human GR. Untransfected BHK/GR cells were used as the control. Expression of GR was maintained with 500 μg G418/ml (*Neo* marker) and C-GFP^{LT} was maintained with 500 μg Zeocin/ml (*Sh ble* marker). CHO cells were also simultaneously co-transfected with vectors containing cDNAs for C-GFP^{LT} and the human ARα2a (ATCC). Transfected cells are referred to as *e.g.* CHO/C-GFP^{LT} cells in the text.

For fluorescence microscopy, cells were allowed to adhere to Lab-Tek chambered coverglasses (Nalge Nunc Int., Naperville, IL, USA) for at least 24 hours and cultured to about 80% confluence. Prior to experiments, the cells were cultured over night without selection pressure in HAM's F12 medium with glutamax (Life Technologies), 100 μ g penicillin-streptomycin mixture ml¹ and 0.3 % FBS. This medium has low autofluorescence enabling fluorescence microscopy of cells straight from the incubator.

Immunoblotting

Samples containing 10 μg of protein, determined according to the method of Bradford³⁶ using the Bio-Rad Protein Assay (Bio-Rad Laboratories, Hercules, CA, USA), were added to SDS sample buffer³⁵ and run on precast 7.5 % SDS-PAGE gels with a 4 % stacking gel (Bio-Rad). The proteins were transferred to PH79 nitrocellulose membranes (Scleicher & Schuell GmbH., Dassel, Germany) for an hour at 4°C using a Bio-Rad Transfer Blot apparatus (80 V). Non-specific adhesion was blocked by

incubating the membranes over night in 3 % bovine serum albumin Fraction V (Sigma Chemical Company, St. Louis, MO, USA) in Tris-buffered saline (TBS) containing 50 mM Tris pH 7.5 and 0.15 M NaCl and for an hour in 3 % skim milk powder (Difco Laboratories, Detroit, MI, USA) in TBS with 0.1 % Tween20 (TBST). The membranes were incubated for an hour in TBST with 3 % skim milk powder and the primary polyclonal rabbit anti-Cα antibody (Upstate Biotechnology Inc., Lake Placid, NY, USA), which was raised against a peptide corresponding to a 16 amino acid N-terminal stretch of human Ca, diluted 1:1000. After 4 washes of 5 min each with TBST, antibody (horse radish peroxidase-conjugated donkey anti-rabbit secondary immunoglobulin from Amersham International plc, Buckinghamshire, UK) diluted 1:5000 in TBS with 3 % skim milk powder was added and incubated for an hour. After 4 washes in TBST and one in TBS, immunoreactivity was detected by enhanced chemiluminescence (ECL) as described by the manufacturer (Amersham) and exposed on Biomax® MR film (Eastman Kodak Company, Rochester, NY, USA). All the steps were performed at room temperature unless otherwise stated.

Time-lapse recording of fusion protein movement.

Cells were cultured in HAM's F12 medium as described above. The chambers were placed on a temperature regulated microscope stage and kept at 37°C. Fluorescence images were captured using an Axiovert 135 inverted light microscope (Carl Zeiss, Oberkochen, Germany) equipped with a Fluar x40, NA 1.3 oil immersion objective (Zeiss) and a cooled (-40°C) CH1 charged coupled device (CCD) camera (Photometrics Ltd., Tucson, AZ. USA). The microscope was equipped with a 470±20 nm excitation filter, a 505 nm dichroic mirror and a 515±15 nm emission filter (Delta Lys & Optik, Lyngby, Denmark). The excitation light source was a 100W HBO arc lamp.

Redistribution of the C-GFP^{LT} fusion protein was quantified using an image analysis program custom written in LabVlEW (National Instruments, Austin, TX, USA). Fluorescent aggregates are segmented from each image using an automatically found threshold based on maximisation of the information measure between the object and the background. The *a priori* entropy of the image histogram is used as the information measure³⁷. The area occupied by aggregates in each image is calculated by counting pixels in the segmented areas. The value thus obtained for each image in a series, or treatment pair, is normalised to the value found for the first (unstimulated) image

collected. A value of zero (0) indicates no redistribution of fluorescence from the starting condition. A value of one (1) by this method equals full redistribution.

Spot photobleaching

A Zeiss LSM 410 with x40 Fluar (as above) was used in spot scan mode at 488 nm to bleach individual fluorescent C-GFP^{LT} aggregates within CHO cells variously treated with forskolin. Fluorescence recovery at the locus of each aggregate was monitored immediately after bleach with successive small-area raster scans just large enough to include most of the cell in which the aggregate lay. Nominal output of the laser at 488 nm, before launch into the microscope, was 10 mW. Subsequent raster scans were also run with the laser at full intensity and without a confocal aperture to allow the first to be made within 0.2 seconds of bleach, and for each scan to be completed within 0.3 seconds (100 x 100 pixels per scan). The recovery of fluorescence for the majority of bleach experiments was measured over a period of 215 seconds, recorded in three consecutive blocks of 10 scans having successive intervals between frames of 0.5, 1 and 5 seconds, and a final set of 15 scans each 10 seconds apart. A single scan collected prior to each bleach exposure served both to establish depth of bleach and to estimate maximum recoverable fluorescence in each experiment. Bleach recovery scans (8-bit images) were analysed using IPlab Spectrum software (Signal Analytics Corp., Vienna, VI, USA). A small region of interest (ROI) of between 6x6 to 10x10 pixels was used to define the area for which fluorescence recovery would be monitored in each experiment, and the average fluorescence within that ROI was measured for successive frames in each time series. The measurement ROIs were slightly larger than the bleached C-GFPLT aggregates to allow for cytoplasmic movements during the measurement period. The total average fluorescence within each frame was also measured to allow fluorescence recovery within C-GFP^{LT} aggregates to be corrected for the minor effects of photobleaching caused by the series of measurement scans.

Results of the spot-bleach experiments are presented as normalised values of displacement from photobleach, $\Delta F(t)$, versus time t:

$$\Delta F(t) = [F(\infty) - F(t)]/[F(\infty) - F(0)]$$

where
$$F(\infty) = F_i \cdot R_i / R_i$$

 $F(\infty)$ being the maximum recoverable fluorescence within a measurement ROI calculated from the pre-bleach intensity of the target aggregate, F_i , corrected for total loss of fluorophores within the cell, R_i/R_i , during the bleach exposure and recovery periods.

SPA

CHO/C-GFP^{LT} cells were cultured in HAM's F12 medium as described above, but in 96-well plates. The medium was exchanged with Ca²⁺-HEPES buffer containing 100 μM IBMX. The cells were stimulated with different concentrations of forskolin for 10 min. Reactions were stopped with addition of NaOH to 0.14 M and the amount of cAMP produced was measured with the cAMP-SPA kit, RPA538 (Amersham) as described by the manufacturer.

Automated imaging

A Diaphot300 microscope (Nikon Corp., Tokyo, Japan) coupled to a camera based on the SITe back illuminated 512 x 512 CCD camera (Princeton Instruments Inc., Trenton, NJ, USA) and integrated with a digital data acquisition system using LabVIEW software was configured to allow automated focusing and image-based analyses in 96-well plates. CHO/C-GFP^{LT} cells were cultured as described above but in 96-well plates and kept at 37°C throughout the experiments. A fluorescence micrograph of the same field of cells, initially chosen at random, was acquired before and 30 min after forskolin stimulation and analysed as described above.

Endomembrane labelling with fluorescently tagged ceramides

Golgi membranes in CHO/C-GFP^{LT} cells were labelled with ceramide-FL (Molecular Probes Inc., Eugene, OR, USA) at 0.5 μ M for 20 minutes before washing. Ceramide-FL excited at 480 nm normally emits in the green at about 510 nm, but when concentrated (as in Golgi membranes) the fluorophore forms excimers, resulting in a shift in the emission maximum to greater than 600 nm³⁸. Images were collected at both 520 \pm 10 nm and beyond 570 nm, allowing good separation of GFP^{LT} and ceramide-FL signals.

Structure of the GFP^{LT}-bright aggregates

Through-focus images of individual C-GFP^{LT} aggregates were collected from chilled cells with a x63 NA 1.4 oil-immersion objective. The built-in focus motor of the Zeiss LSM 410 was used to advance the objective 0.2 µm between images, 25 images per data set. Effective pixel size in the images was 65.6 nm. Data sets were corrected for bleaching and fluctuations in illumination intensity. Out-of-focus information in the images was removed using iterative, constrained, three-dimensional deconvolution (DeltaVision from Applied Precision Inc., Seattle, WA, USA) based on a theoretically calculated point-spread function. The deconvolved images were then reconstructed into a 3-D rotational projection of 40 images (9 degrees between images) using the method of maximum intensity ray-tracing (DeltaVision, Applied Precision, Inc., Seattle, USA). Two adjacent images in this set, re-sized and pixel-smoothed, were used to create the stereo pair shown in Figure 6C. An iso-surface rendering of the 3-D reconstruction was created using Milan software (BitPlane AG, Zurich, Switzerland) (Fig. 6B).

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References

- 1. Walsh, D.A., Perkins, J.P., and Krebs, E.G. 1968. An adenosine 3'.5'-monophosphate-dependant protein kinase from rabbit skeletal muscle. *J. Biol. Chem.* 243:3763-3765.
- 2. Taskén, K., Skålhegg, B.S., Taskén, K.A., Solberg, R., Knutsen, H.K., Levy, F.O., Sandberg, M., Ørstavik, S., Larsen, T., Johansen, A.K., Vang, T., Schrader, H.P., Reinton, N.T.K., Torgersen, K.M., Hansson, V., and Jahnsen, T. 1997. Structure, function, and regulation of human cAMP-dependent kinases. *Adv. Second Messenger Phosphoprotein Res.* 31:191-204.
- Kingston, P.A., Zufall, F., and Barnstable, C.J. 1996. Rat hippocampal neurons express genes for both rod retinal and olfactory cyclic nucleotide-gated channels: Novel targets for cAMP/cGMP function. *Proc. Natl. Acad. Sci. USA* 93:10440-10445.
- 4. Taskén, K., Solberg, R., Foss, K.B., Skålhegg, B.S., Hansson, V., and Jahnsen, T. 1995. *The Protein Kinase Facts Book: Protein-Serine Kinases* (Eds., Hardie, G., and Hanks, S.), Academic Press Ltd., London, UK, pp 58-63
- Taskén, K., Skålhegg, B.S., Solberg, R., Andersson, K.B., Taylor, S.S., Lea, T., Blomhoff, H.K., Jahnsen, T., and Hansson, V. 1993. Novel isozymes of cAMPdependent protein kinase exist in human cells due to formation of RIα-RIβ heterodimeric complexes. J. Biol. Chem. 268:21276-21283.
- Beebe, S.J., Salomonsky, P., Jahnsen, T., and Li, Y. 1992. The Cγ subunit is a unique isozyme of the cAMP-dependent protein kinase. J. Biol. Chem. 267:25505-25512.
- 7. Gamm, D.M., Baude, E.J., and Uhler, M.D. 1996. The major catalytic subunit isoforms of cAMP-dependent protein kinase have distinct biochemical properties *in vitro* and *in vivo*. *J. Biol. Chem.* **271:**15736-15742
- 8. Robinson, M.L., Wallert, M.A., Reinitz, C.A., and Shabb, J.B. 1996. Association of the type I regulatory subunit of cAMP-dependent protein kinase with cardiac myocyte sarcolemma. *Arch. Biochem. Biophys.* 330:181-187.
- 9. Moos, J., Peknicová, J., Geussová, G., Philimonenko, V., and Hozák, P. 1998. Association of protein kinase A type I with detergent-resistant structures of mammalian sperm cells. *Mol. Reprod. Dev.* 50:79-85.

- 10. Nigg, E.A., Schäfer, G., Hilz, H., and Eppenberger, H.M. 1985. Cyclic-AMP-dependent protein kinase type II is associated with the Golgi complex and with centrosomes. *Cell* 41:1039-1051.
- 11. Li, Y., Ndubuka, C., and Rubin, C.S. 1996. A kinase Anchor protein 75 targets regulatory (RII) subunits of cAMP-dependent protein kinase II to the cortical actin cytoskeleton in non-neuronal cells. *J. Biol. Chem.* 271:16862-16869.
- 12. Pariset, C., and Weinman, S. 1994. Differential localization of two isoforms of the regulatory subunit RIIα of cAMP-dependent protein kinase in human sperm: Biochemical and cytochemical study. *Mol. Reprod. Dev.* **39:**415-422.
- 13. Shmyrev, I.I., Grozdova, I.D., Kondratyev, A.D., Mamayeva, E.G., and Severin, E.S. 1990. Immunofluorescence localization of the regulatory subunit type II of cAMP-dependent protein kinase in PC12 and 3T3 cells in different proliferative states. *Mol. Cell. Biochem.* 93:47-52.
- 14. Zhang, Q., Carr, D.W., Lerea, K.M., Scott, J.D., and Newman, S.A. 1996. Nuclear localization of type II cAMP-dependent protein kinase during limb cartilage differentiation is associated with a novel developmentally regulated A-kinase anchoring protein. *Dev. Biol.* 176:51-61.
- 15. Coghlan, V.M., Bergeson, S.E., Langeberg, L., Nilaver, G., and Scott, J.D. 1993. *A-K*inase Anchoring Proteins: a key to selective activation of cAMP-responsive events? *Mol. Cell. Biochem.* 127/128:309-319.
- 16. Huang, L.J.-S., Durick, K., Weiner, J.A., Chun, J., and Taylor, S.S. 1997. D-AKAP2, a novel protein kinase A anchoring protein with a putative RGS domain. *Proc. Natl. Acad. Sci. USA* 94:11184-11189.
- 17. Carr, S.A., Biemann, K., Shoji, S., Parmelee, D.C., and Titani, K. 1982. *n*-Tetradecanoyl is the NH₂-terminal blocking group of the catalytic subunit of cyclic AMP-dependent protein kinase from bovine cardiac muscle. *Proc. Natl. Acad. Sci. USA* **79:**6128-6131.
- 18. Adie, E.J., Thomas, P.H., Munday, M.R., and Clegg, R.A. 1995. Subcellular targeting of recombinant and mammalian Cα subunits of cAMP-dependent protein kinase. *Biochem. Soc. Trans.* 23:451S.
- 19. Yang, S., Fletcher, W.H., and Johnson, D.A. 1995. Regulation of cAMP-dependent protein kinase: Enzyme activation without dissociation. *Biochemistry* **34:**6267-6271.

- 20. Hardie, D.G. 1991. Biochemical Messengers: Hormones, Neurotransmitters and Growth Factors, Chapman & Hall, London, UK
- 21. Harootunian, AT., Adams, SR., Wen, W., Meinkoth, J.L., Taylor, S.S., and Tsien, R.Y. 1993. Movement of the free catalytic subunit of cAMP-dependent protein kinase into and out of the nucleus can be explained by diffusion. *Mol. Biol. Cell* **4:**993-1002.
- 22. Nigg, E.A., Hilz, H., Eppenberger, H.M., and Dutly, F., 1985. Rapid and reversible translocation of the catalytic subunit of cAMP-dependent protein kinase type II from the Golgi complex to the nucleus. *EMBO J.* **4:**2801-2806.
- 23. Walsh, D.A., and Van Patten, S.M. 1994. Multiple pathway signal transduction by the cAMP-dependent protein kinase. *FASEB J.* 8:1227-1236.
- 24. Mellon, P.L., Clegg, C.H., Correll, L.A., and McKnight, G.S. 1989. Regulation of transcription by cyclic AMP-dependent protein kinase. *Proc. Natl. Acad. Sci. USA* **86:**4887-4891.
- 25. Hagiwara, M., Brindle, P., Harootunian, A., Armstrong, R., Rivier, J., Vale, W., Tsien, R., and Montminy, M.R. 1993. Coupling of hormonal stimulation and transcription via the cyclic AMP-responsive factor CREB is rate limited by nuclear entry of protein kinase A. *Mol. Cell. Biol.* 13:4852-4859.
- 26. Adams, S.R., Harootunian, A.T., Buechler, Y.J., Taylor, S.S., and Tsien, R.Y. 1991. Fluorescence ratio imaging of cyclic AMP in single cells. *Nature* **349**:694-697.
- 27. Elson, E.L., and Qian, H. 1989. Interpretation of fluorescence correlation spectroscopy and photobleaching recovery in terms of molecular interactions. *Methods Cell Biol.* **30:**307-332.
- 28. Lipsky, N.G., and Pagano, R.E. 1985. A vital stain for the Golgi apparatus. *Science* 288:745-747.
- 29. Knighton, D.R., Zheng, J., Ten Eyck, L.F., Ashford, V.A., Xuong, N.-h., Taylor, S.S., and Sowadski, J.M. 1991. Crystal structure of the catalytic subunit of cyclic adenosine monophosphate-dependent protein kinase. *Science* 253:407-414.
- 30. Knighton, D.R., Zheng, J., Ten Eyck, L.F., Xuong, N.-h., Taylor, S.S., and Sowadski, J.M. 1991. Structure of a peptide inhibitor bound to the catalytic subunit of cyclic adenosine monophosphate-dependent protein kinase. *Science* 253:414-420.

- 31. Scott, J.D. 1991. Cyclic nucleotide-dependent protein kinases. *Pharmacol. Ther.* **50:**123-145.
- 32. Skålhegg, B.S., Keryer, G., Torgersen, K.M., Aandahl, E.M., Levy, F.O., Jahnsen, T., Hansson, V., and Taskén, K. 1997. Function and localization of cAMP-dependent protein kinase type I and II in lymphoid cells. *FEBS special meeting* 1997: Cell signalling mechanisms, Elsevier Science, The Netherlands, Abstract P6-123
- 33. Akey, C.W. 1995. Structural plasticity of the nuclear pore complex. J. Mol. Biol. 248:273-293.
- 34. Uhler, M.D., Carmichael, D.F., Lee, D.C., Chrivia, J.C., Krebs, E.G., and McKnight, G.S. 1986. Isolation of cDNA clones coding for the catalytic subunit of mouse cAMP-dependent protein kinase. *Proc. Natl. Acad. Sci. USA* 83:1300-1304.
- 35. Sambrook, J., Fritsch, E.F., and Maniatis, T. 1989. *Molecular Cloning: A Laboratory Manual, 2nd Ed.*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, USA
- 36. Bradford, M.M. 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* 72:248-254.
- 37. Kapur, J.N., Sahoo, P.K., and Wong, A.K.C. 1985. A new method for gray-level picture thresholding using the entropy of the histogram. *Computer Vision, Graphics, and Image Processing* **29:**273-285.
- 38. Haugland, R.P. 1996. *Handbook of fluorescent probes and research chemicals*, 6th Ed. Molecular Probes, Oregon, USA, p281.

Figure legends

Table 1. Time from initiation of a response to half maximal $(t_{1/2 max})$ and maximal (t_{max}) C-GFP^{LT} redistribution. The data was extracted from curves such as shown in Figure 3B. All $t_{1/2 max}$ and t_{max} values are given as mean±SD and are based on a total of 26-30 cells from 2-3 independent experiments for each forskolin concentration. Since the observed redistribution is sustained over time, the t_{max} values were taken as the earliest time point at which complete redistribution is reached. Note that the values do not relate to the degree of redistribution.

Figure 1. Western blot analysis of lysates containing C-GFP^{LT} fusion proteins. Total lysates of BHK/GR,C-GFP^{LT} (A) and control BHK/GR (B) cells were probed with an anti-Cα antibody. 500 ng of purified bovine C subunit (C) was included as a positive control and to identify the endogenous C subunit. Although the antibody clearly reacts unspecifically with several proteins in the total lysates, the fusion protein (f) is recognised as a specific band, migrating with an apparent size of 60 kDa, in the transfected cells (A). The endogenous C subunit (e) migrated as predicted by its molecular weight of 41 kDa. It is possible to compare the expression levels of endogenous hamster C subunit and overexpressed mouse fusion proteins in these blots since the immunogenic peptide is conserved between these two species.

Figure 2. Fluorescence micrographs of CHO cells expressing C subunit fusion proteins. The two fusion proteins of the C subunit of cAK show distinct localisation patterns. A. The NH₂-terminal GFP^{LT}-C fusion protein is localised almost evenly throughout the cytoplasm. B. The COOH-terminal C-GFP^{LT} fusion protein is highly concentrated in cytoplasmic aggregates, often in one large and several minor structures per cell. Scale bar 10 μm.

Figure 3. Time-lapse analyses of fluorescence redistribution in CHO/C-GFP^{L1} cells treated with various agonists. The raw data of each experiment consisted of 60 fluorescence micrographs acquired at regular intervals including several images acquired before the addition of agonist. Six of these images are shown (A) for the typical response to 1 μ M forskolin, taken at the time points indicated. The time point t=0 corresponds to the image acquired immediately before the cells were challenged with agonist. Scale bar

10 μm. The charts (B-G) each show a quantification of the responses in each time series. The total area of the highly fluorescent aggregates (see Experimental Protocol) is plotted versus time for each experiment. (B) Redistribution time profiles of the C-GFP^{LT} fusion following treatment of cells with various concentrations of forskolin. (C) Response following addition of 1 mM dbcAMP. (D) The effect of 100 μM IBMX on the fusion protein distribution. (E) Demonstrates the reversibility of the forskolin-induced redistribution of C-GFP^{LT}, where 10 μM forskolin (open arrow) is followed shortly by repeated washings with buffer (dark arrow). In a parallel experiment, treatment with 10 μM forskolin plus 100 μM IBMX is followed by repeated washing with buffer containing 100 μM IBMX. (F) BHK/GR,C-GFP^{LT} cells treated with 100 nM glucagon. (G) CHO/C-GFP^{LT} cells transiently transfected with the ARα2a were pretreated with 10 μM forskolin (open arrow) to increase [cAMP], then given 10 μM norepinephrine in the continued presence of forskolin.

Figure 4. (A) Four frames from the recovery sequence following spot photobleach of a large aggregate (arrow) in a CHO/C-GFP^{LT} cell exposed to 25 μM forskolin. Times are seconds after bleach. (B) Normalised displacement curves of the fluorescence recovery process in cells exposed to various levels of forskolin. Measurement points are averages±sem (n=4). (C) Linear fits to the first five points of the normalised recovery curves shown in (B). The slope of each line is used as an estimate of the half-time of recovery from bleach at each forskolin concentration.

Figure 5. Parallel dose response analyses of forskolin effects in CHO/C-GFP^{LT} cells on: [cAMP], elevation (□), the rate of recovery from spot photobleach (Δ) and induced change in C-GFP^{LT} redistribution (•). [cAMP], was measured by SPA assay, analysing the effects of buffer or 8 increasing concentrations of forskolin in these cells. The graph shows a trace of the mean±sem expressed in arbitrary units (n=4 for each data point). Half times for recovery from spot photobleach were estimated from the first 5 time points of the mean value (n=4) curves in Figure 4B. Changes induced in C-GFP^{LT} distribution were quantified as described (Experimental Protocol) using fluorescence micrographs taken of the same field of cells prior to and 30 min after the addition of forskolin. The graph shows a trace of the mean±sem at each forskolin concentration (n=8 for each data point). The fitted curves indicate ½-maximal concentration values for

forskolin as: 1.7 μ M, image-based assay (\square); 3.0 μ M, spot photobleach assay (Δ); 9.3 μ M, SPA (\bullet).

Figure 6. (A) Two images of CHO/C-GFP^{LT} cells stained with ceramide-FL, in emission ranges of 520 ± 10 nm and >570 nm, have been superimposed to demonstrate the distinct separateness of Golgi membranes (orange) and C-GFP^{LT} fluorescence (green). Scale bar is $10 \mu m$. (B) An iso-surface rendering of a single large C-GFP^{LT} aggregate (similar to that arrowed in 6A). The image is a reconstruction from 25 through-focus images deconvolved and processed as described (Experimental Protocol). Scale bar $1 \mu m$. (C) Stereo pair of the reconstructed images used to generate the iso-surface seen in (B). Each image is smoothed for presentation, the structure originally being 35 pixels high by 27 wide in this orientation. Scale bar $1 \mu m$.

Figure 1

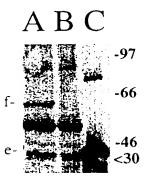
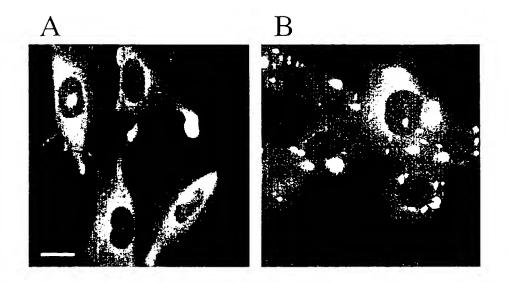
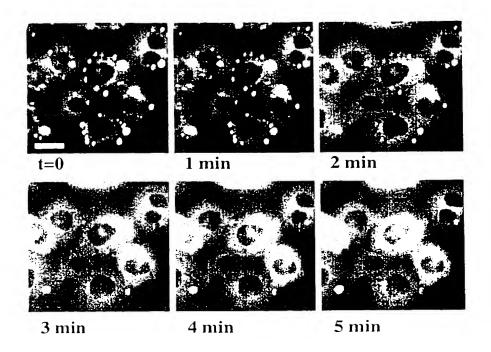
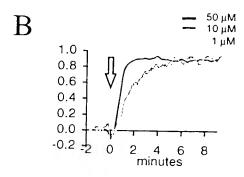


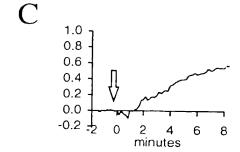
Figure 2

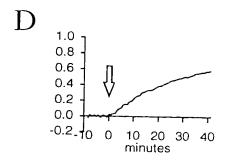


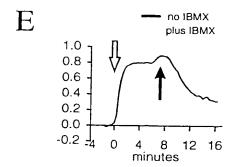
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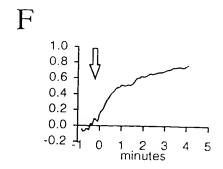












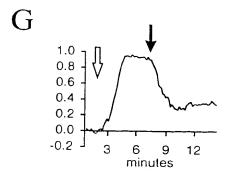
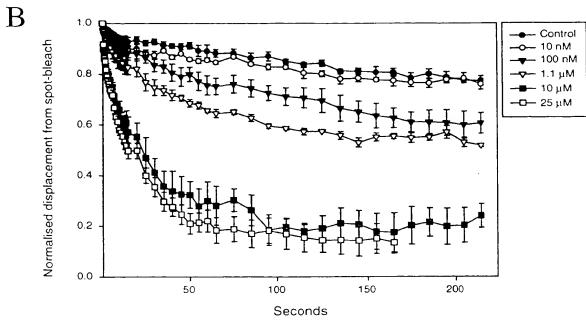
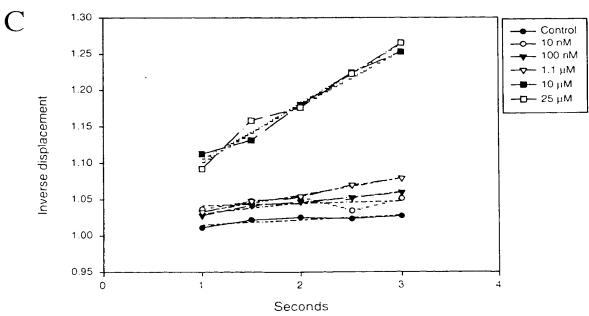


Figure 4

A
0.5
6.0
20.0
165.0





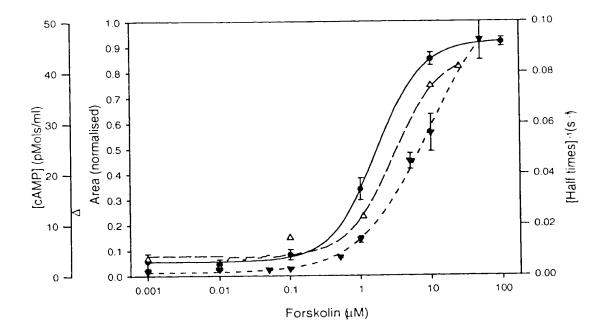
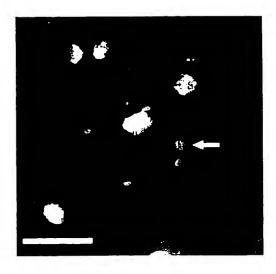
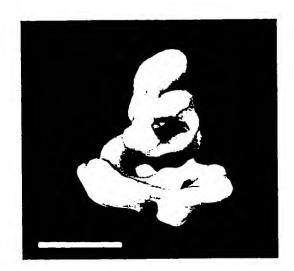


Figure 6

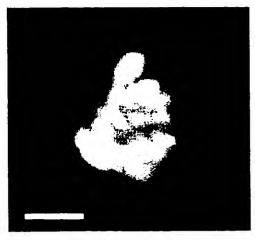
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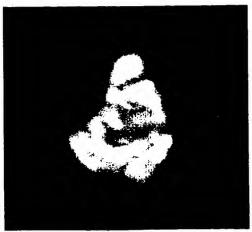
В





 \mathbf{C}





Left

Right

Table 1

[foral-1: 1/ >		
[forskolin]/µM	$t_{1/2max}/s$	t _{max} / s
1	115±21	310±31
10	69±14	224±47
50	47±10	125±28

k/ E			